

Abstract

In the predominantly oxic, upland soils, periods of high wetness trigger anaerobic processes such as iron (Fe) reduction within the soil microsites, with implications for organic matter decomposition, the fate of pollutants, and nutrient cycling. In fluctuating O_2 conditions, Fe reduction is maintained by the re-oxidation of ferrous iron, which renews the electron acceptor, Fe^{III} , for microbial Fe reduction. To characterize such processes, it is fundamental to relate the redox cycling of iron between the two redox states to the hydro-climatic conditions. Here, we link iron cycling to soil moisture variability through a model of iron-redox dynamics and find the hydrologic regime that maximizes Fe reduction, under non-limiting organic carbon availability. Away from the optimal cycle, the duration of the oxic or the anoxic phase limits the regeneration of Fe^{III} or its reduction rate, respectively. We relate the average duration of the oxic and anoxic intervals to the frequency and mean depth of precipitation events that drive the dynamics of soil moisture, effectively linking iron cycling to the hydrologic regime. We then compare a tropical (Luquillo CZO) and a subtropical (Calhoun CZO) forest to provide insights into the soil moisture control on iron-redox dynamics in these ecosystems. The tropical site maintains a high potential for iron reduction throughout the year, due to quick and frequent transitions between oxic and anoxic conditions, whereas the subtropical site is strongly affected by seasonality, which limits iron reduction to winter and early-spring months with higher precipitation and lower evaporative demand.

Plain Language Summary

Iron (Fe) plays a critical role in terrestrial ecosystems, influencing from the carbon cycle to the mobilization of contaminants and the formation of colloidal particles. It thus important to understand and quantify its biogeochemical cycle in relation to the environmental factors that drive it, for example the oxygen content in the soil pores. Here, we couple its redox cycle, consisting of Fe reduction and subsequent Fe oxidation, to the in-situ rainfall and soil moisture variability and show that the cycle is faster for a specific hydro-climate. These results represent an important step towards predicting the potential for iron redox cycling across different climate and identify the climatic regions where the Fe biogeochemical cycle may participate more actively in ecosystem functioning.

1 Introduction

The iron (Fe) biogeochemical cycle is an important component of terrestrial ecosystems, where it is implicated in the decomposition of the organic matter (E. Herndon et al., 2017; Bhattacharyya et al., 2018; Calabrese & Porporato, 2019; Vermeire et al., 2019; Han et al., 2019; Zheng et al., 2019; LaCroix et al., 2019), the formation of colloids (Stucki, 2011; Henderson et al., 2012; Wang et al., 2019) and mobilization of contaminants (Borch et al., 2009; Bishop et al., 2014; Couture et al., 2015; Yu et al., 2016). Predicting the variations in Fe reduction rates as a function of the hydro-climatic requires linking processes from the pedon to the watershed scale, but this has been challenging because of the numerous factors that affect the Fe redox chemistry.

The fundamental constraint on the Fe redox dynamics is the reduction of Fe^{III} , which has slower kinetics than the oxidation of Fe^{II} (Lovley, 1991; Ginn et al., 2017; Chen & Thompson, 2017). During anoxic conditions, Fe^{III} -reducing microorganisms rely on the availability of Fe^{III} -oxides as an electron acceptor, reducing it to ferrous iron (Fe^{II}) in order to decompose the organic matter (Lovley, 1991; Roden & Wetzels, 1996; Dubinsky et al., 2010). The rate of Fe^{III} reduction thus depends on a suitable organic substrate (LaRowe & Van Cappellen, 2011), the activity of Fe-reducers, as well as the abundance of Fe^{III} electron acceptor relative to other more thermodynamically favorable ones (e.g., O_2 , Mn^{IV}).

66 The energy yield obtained from oxidizing organic matter coupled to Fe as electron
67 acceptor is lower than the energy yield obtained when coupled to O₂. Thus, Fe reduc-
68 tion is strongly dependent upon the availability of an easily degradable substrate (LaRowe
69 & Van Cappellen, 2011), whereas those substrates that require more energy to oxidize
70 (i.e., have higher ΔG values for the C oxidation half reaction) can become thermody-
71 namically unfavorable for microbial Fe^{III} reduction. The abundance and activity Fe-reducers
72 is critical for predicting Fe reduction rates. Laboratory and field observations both have
73 shown that Fe reduction is faster when the soil has experienced Fe reduction in the re-
74 cent past (Buettner et al., 2014; Barcellos, Cyle, & Thompson, 2018), suggesting increased
75 Fe-reducers activity in these conditions. Lastly, higher reduction rates are driven mostly
76 by recently oxidized Fe^{II} (Weiss et al., 2004, 2005; Thompson et al., 2006). The avail-
77 ability of Fe^{III} electron acceptor can in fact be quantified through measurements of short
78 range ordered Fe^{III} minerals.

79 The above arguments suggest that Fe reduction rates are strongly controlled by the
80 characteristics of the soil oxic/anoxic cycles. In fact, in well-aerated soils (oxic conditions),
81 iron mostly remains in its oxidized state (Fe^{III}) and aerobic respiration is the main mech-
82 anism of carbon decomposition, whereas in nearly constantly anoxic environments, such
83 as wetlands or paddy soils, iron may persist in its reduced state (Fe^{II}) and other metabolisms
84 typical of low redox potentials may be triggered, i.e., fermentation or methanogenesis
85 (Morel et al., 1993; Brady & Weil, 2016). Between these extreme scenarios, a contin-
86 uous transitions between oxic and anoxic conditions (e.g., wet tropical soils, river banks,
87 fluctuating water tables), which spurs the formation of degradable organic substrates,
88 higher activity of Fe reducers, and the continuous regeneration of Fe^{III}, may favor high
89 rates of iron reduction (Calabrese & Porporato, 2019).

90 The main environmental factor controlling the transitions between oxic and anoxic
91 conditions is the soil water content (Todd-Brown et al., 2012; Brady & Weil, 2016), as
92 this determines the activity of aerobic bacteria and the fraction of air-filled volume. Ex-
93 perimental studies show that soil moisture may be a proxy for oxygen content, because
94 this remains relatively high ($\approx 20\%$) for water contents up to the soil field capacity and
95 then nonlinearly declines to $\approx 0\%$ as the soil approaches saturation (Hall et al., 2013;
96 Barcellos, OConnell, et al., 2018). A quantification of the fate and redox changes of soil
97 iron, necessary for the understanding of the global carbon cycle and related climate dy-
98 namics (Colombo et al., 2014; Oertel et al., 2016; Zheng et al., 2019), then needs to be
99 carried out in relation to how hydro-climatic variability can induce changes in soil aer-
100 ation and redox potential.

101 Towards this goal, we derive the relationship between the average Fe reduction rate
102 and the length of exposure to oxic and anoxic conditions, which is related to the hydro-
103 logic regime. By means of a mechanistic iron-redox model, we explore the interaction
104 between the timescales of the biogeochemistry (i.e., the reaction rates) and of the changes
105 in environmental conditions (i.e., oxic/anoxic cycle) and highlight the existence of a max-
106 imum average Fe reduction rate at an intermediate anoxic/oxic intervals ratio. We then
107 relate the oxic/anoxic cycle to the hydro-climatic fluctuations and link the characteris-
108 tics of the cycle to the statistical properties of the soil moisture dynamics and precip-
109 itation, in terms of its frequency and mean rainfall depth. This effectively links iron re-
110 duction to the in-situ hydro-climatic variability, for which measurements are readily ob-
111 tained through direct or remote sensing techniques. Applying the framework to soils from
112 a humid tropical forest (Luquillo CZO) and a subtropical forest (Calhoun CZO), we ex-
113 plore the iron-redox dynamics in these different ecosystems and discuss its control on the
114 carbon cycle and plant primary productivity.

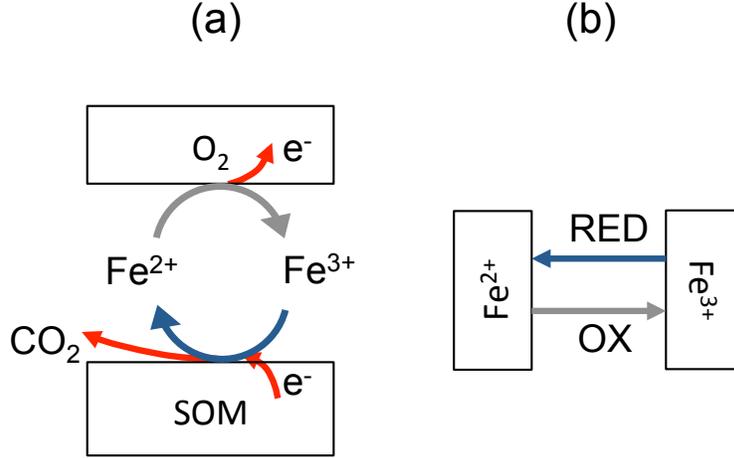


Figure 1. (a) Schematic representation of the soil iron cycle, emphasizing the electron transport from the soil organic matter (SOM) to the atmospheric oxygen by means of iron Fe^{II}, which operates as an ‘electron carrier’. See description in Section 2. (b) Schematics of the iron redox cycle described by equation (1).

2 Optimal oxic/anoxic cycle for Fe reduction

The soil iron cycle, summarized in Figure 1(a), has an anoxic phase, in which Fe^{III} is utilized as an electron acceptor to decompose organic matter and an oxic phase, when oxygen oxidizes Fe^{II}, thus regenerating the Fe^{III} pool. Going back and forth between the two oxidation states, iron operates as an ‘electron carrier’ between the soil organic matter and the atmospheric oxygen (Figure 1 (a)), so that the decomposition depends on the rate at which electrons can be transported from the organic matter to oxygen. Decomposition by iron reduction in fact needs a continuous supply of iron Fe^{III}, which after having been reduced to Fe^{II} during an anoxic phase needs to be regenerated (i.e., re-oxidized) during the subsequent oxic phase. It is thus clear that the hydro-climate generating the oxic/anoxic cycles exerts a major control on the rate of iron cycling.

Consider the top soil layer containing organic matter and refer to the total iron content in the oxidized and reduced states as Fe^{II} and Fe^{III} , respectively. The total content of reducible iron is constant and equal to $Fe^{TOT} = Fe^{II} + Fe^{III}$. Since our focus is on the maximum rates, we assume that the availability of the organic substrate and microbes does not limit the reactions, so that the regeneration of Fe^{III} electron acceptor and presence/absence of anoxic conditions limit the reaction. The hydrologic cycle will thus govern the reaction rates in this framework. The soil is subject to an oxic/anoxic cycle of duration T that begins with the anoxic phase of duration $\tau_a = fT$ (Figure 2), whereas the oxic phase lasts for $\tau_o = (1 - f)T$, f being the anoxic fraction. During the anoxic phase, only iron reduction occurs (no oxidation allowed), with a consequent increase of Fe^{II} . During the oxic phase, iron reduction stops and Fe^{II} is oxidized to Fe^{III} (Figure 1(b)). Such dynamics are described by the following mass balance equation,

$$\frac{dFe^{II}}{dt} = RED - OX, \quad (1)$$

where $RED = k_R(Fe^{TOT} - Fe)$ and $OX = k_O Fe^{II}$, k_R and k_O being the reduction and oxidation rate constants, respectively. Note that these expressions do not contain a dependence on the amount of substrate and microbial activity, as we are focusing exclusively on the hydrologic regime. However, the rate constants do explicitly depend on the time, t , in that during the anoxic phase $k_O = 0$, while during the oxic phase $k_R =$

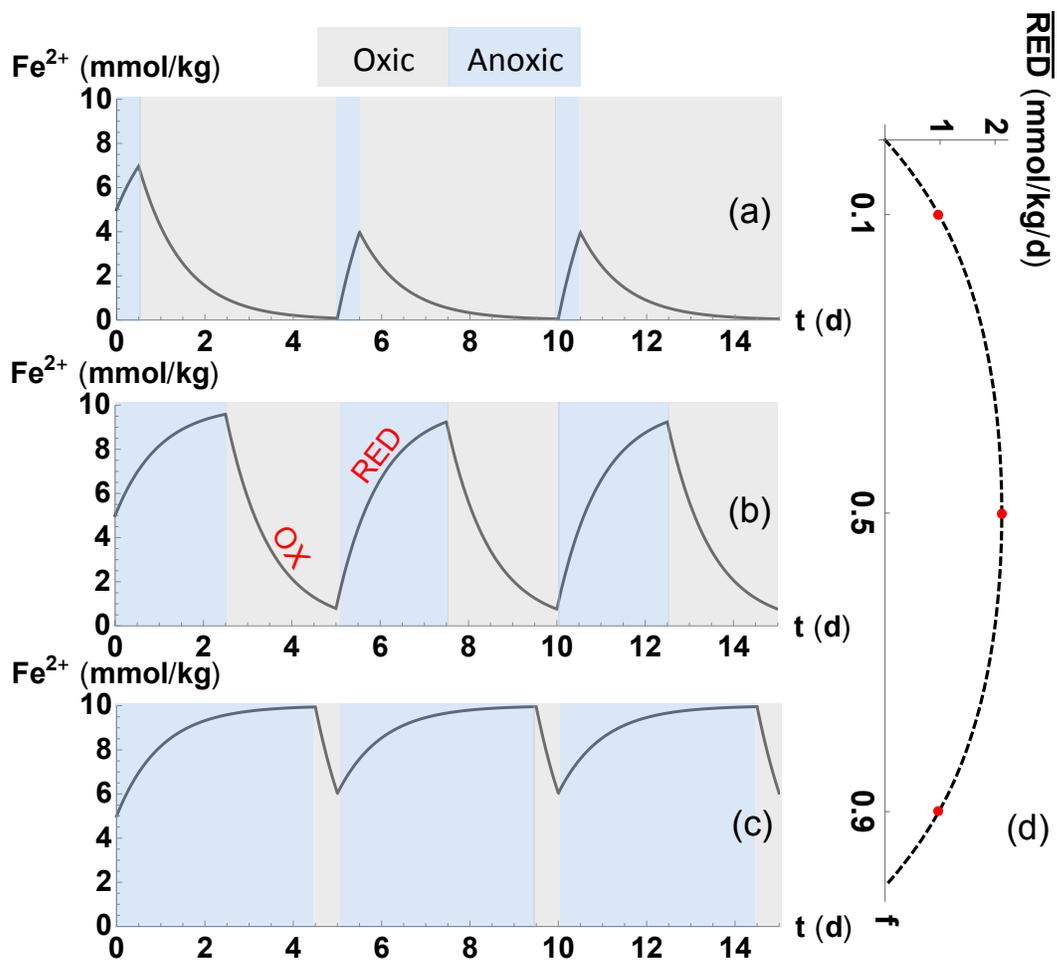


Figure 2. Time evolution of Fe^{II} content for different values of the ratio of anoxic/oxic intervals, $f = 0.1$ in (a), $f = 0.5$ in (b) and $f=0.9$ in (c). The period of the anoxic/oxic cycle is fixed, $T = 5$ days. (d): (Dashed line) Average reduction rate, $\overline{RED} = 1/T \int_T RED(t)dt$, as a function of f , for $T = 5$ days. (Red points) Average reduction rate for f equal to 0.1, 0.5, and 0.9.

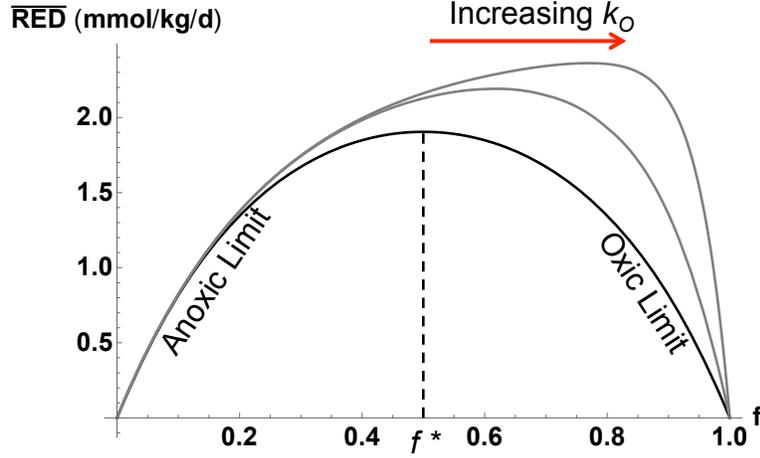


Figure 3. Average reduction rate, $\overline{RED} = 1/T \int_T RED(t)dt$, for different values of the reduction and oxidation rate constants, as a function of f and $T = 5$ days. The reduction rate constant $k_R = 1$ mmol/kg/d, while the oxidation rate constants are, from left to right, 1, 2 and 5 mmol/kg/d.

143 0. Solving equation (1) for sufficiently long time such that the initial condition has no
 144 longer influence, the stationary solution for a given oxic/anoxic cycle (shown in Figure
 145 2(a)) is given by an exponential decay during the oxic phase,

$$Fe^{II}(t) = Fe_0^{II} e^{-k_O t}, \quad (2)$$

146 where Fe_0^{II} is the iron content at the end of the preceding anoxic phase and t is the time
 147 elapsed since the beginning of the oxic phase. On the contrary, during the anoxic phase
 148 Fe^{II} increases, approaching exponentially Fe^{TOT} ,

$$Fe^{II}(t) = Fe^{TOT} - Fe_0^{II'} e^{-k_O t}, \quad (3)$$

149 $Fe_0^{II'}$ being the iron content at the end of the preceding oxic phase and t the time elapsed
 150 since the beginning of the anoxic phase.

151 In an extreme scenario, in which conditions are set to be always oxic ($f = 0$, Fig-
 152 ure 2), iron content persists in its oxidized state, $Fe^{II}(t) = 0$, and the average reduc-
 153 tion rate, which can be defined as $\overline{RED} = 1/T \int_T RED(t)dt$, goes to zero. On the other
 154 hand, for a scenario of constant anoxic conditions ($f = 1$, Figure 2) iron persists in its
 155 reduced state, $Fe^{II}(t) = Fe^{TOT}$, and again the reduction rate $\overline{RED} = 0$. This argu-
 156 ment suggests that a maximum reduction rate \overline{RED}^* exists at an intermediate value of
 157 f , f^* . Solving equation (1) for different values of f , the different Fe^{II} trajectories are shown
 158 in Figure 2, and computing the average reduction rate per cycle, see Figure 2(d), illus-
 159 trates the anoxic/oxic cycle for which the \overline{RED} is maximum.

160 The optimal f^* at which the maximum Fe^{III} is achieved depends on the reaction
 161 rate constants, k_R and k_O . For simplicity, Figure 2 demonstrates that, in the hypothet-
 162 ical condition in which $k_R = k_O$, the resulting $f^* = 0.5$. For higher k_R or k_O , shorter
 163 anoxic or oxic phases are needed to reduce or oxidize the same amount of iron, respec-
 164 tively. As the ratio of anoxic/oxic time moves away from the optimal f , the oxic/anoxic
 165 cycles is favoring either the reduction ($f > f^*$) or the oxidation ($f < f^*$), leading to
 166 an inhibition of Fe cycling. When $f > f^*$, the iron-redox cycle is limited by the regen-
 167 eration of Fe^{III} electron acceptor, as essentially there is not enough time to oxidize enough
 168 sufficient iron to use in the following anoxic phase. On the contrary, when $f < f^*$ the
 169 iron-redox cycle is limited by the Fe^{III} reduction, such that the anoxic phase is too short
 170 to reduce substantial amounts of iron (Figure 3).

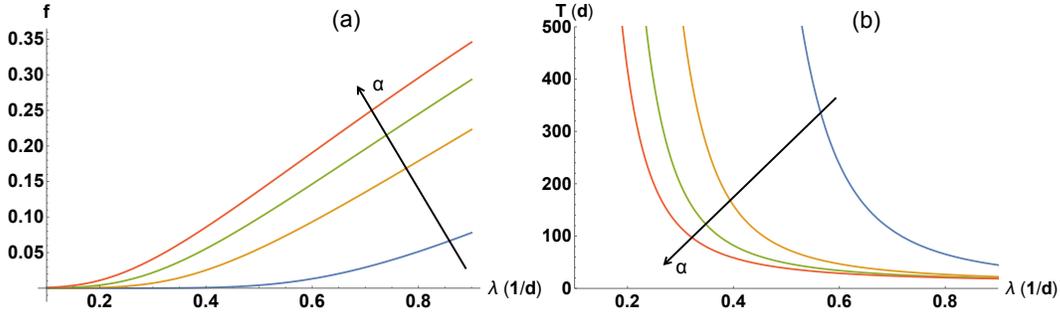


Figure 4. (a) Anoxic fraction of the cycle, f , and (b) duration of the cycle, T , as a function of the frequency of precipitation events, λ , and for different values of mean precipitation depth, α . The values of α , from blue to red line, are 5, 8, 10, and 12 mm. The probability density function for the soil moisture was computed from (Laio et al., 2001), for a silty clay loam soil, average porosity $n = 0.48$, potential evapotranspiration $PET = 4$ mm/d, and hydraulic conductivity $k_h = 14$ cm/d.

171 3 Soil moisture control on oxic/anoxic cycle

172 Under field conditions, the frequency and depth of the rainfall events, evapo-transpiration
 173 from soil and plants, and soil properties altogether determine the evolution of the soil
 174 water and oxygen content, causing the soil to undergo transitions between oxic and anoxic
 175 conditions. Since oxygen content exhibits a first dependence on soil moisture (Hall et
 176 al., 2013; Calabrese & Porporato, 2019), the average duration of the oxic and anoxic phases,
 177 for a given hydro-climate, can be obtained by analyzing the specific time series of soil
 178 moisture. Fixing the soil moisture threshold \hat{s} above which soil conditions can be con-
 179 sidered anoxic (there are enough anoxic soil microsites to activate anaerobic processes),
 180 the average time spent in oxic conditions τ_o then can be calculated as the average time
 181 of each excursion below the threshold \hat{s} . The average time of each excursion above \hat{s} will
 182 be τ_a , the average duration of a oxic/anoxic cycle $T = \tau_a + \tau_o$, and in turn the anoxic
 183 fraction of time $f = \tau_a/\tau$.

184 We show the relationship between the anoxic fraction f , cycle length T and the fre-
 185 quency and mean depth of precipitation (λ and α , respectively) in Figure 4. The curves
 186 are drawn for constant soil properties (typical of a silty clay loam) and potential evap-
 187 otranspiration ($PET = 4$ mm/d) using a stochastic water balance that provides the
 188 statistical properties of soil moisture based on rainfall statistics (Laio et al., 2001) (see
 189 Appendix A). Because of the high water losses at soil moisture above field capacity, the
 190 fraction of time spent in anoxic conditions is generally lower than the one spent in oxic
 191 conditions, such that the values of f are below 0.5 (Figure 4). As can be expected, soils
 192 are in anoxic conditions on average longer (higher values of f) for high rainfall frequen-
 193 cies accompanied by high average rainfall depths. On the contrary, the whole duration
 194 of the cycle, T , decreases with λ as the excursion from oxic to anoxic is more likely to
 195 occur. For the realistic range of mean rainfall depth α and frequency λ explored, the length
 196 of the full anoxic and oxic cycle T decreases with α , again because it becomes more likely
 197 that the soil moisture threshold \hat{s} is crossed. However, for very high mean rainfall depth
 198 α and frequency λ the trend in Figure 4 may be inverted as the soil switches to very wet
 199 conditions that are in anoxic conditions ($s > \hat{s}$) for most of the time.

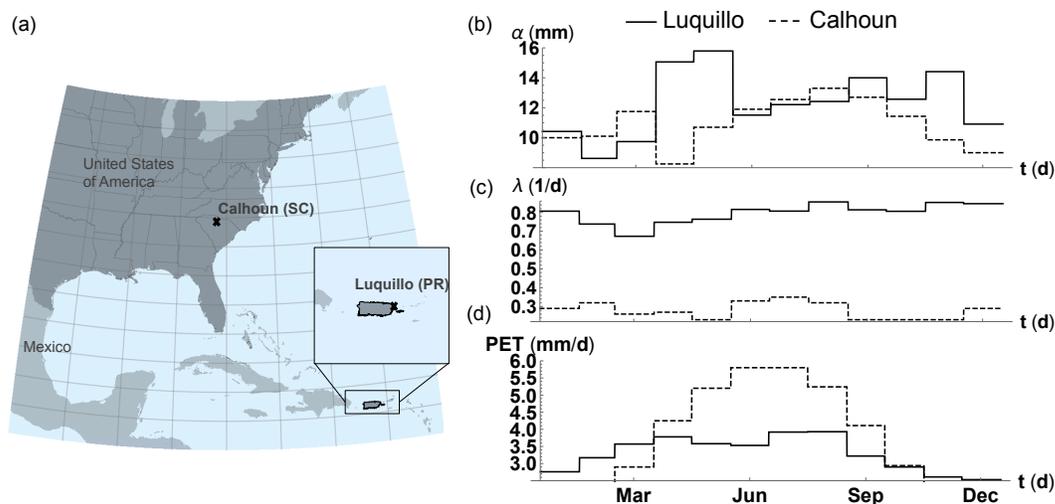


Figure 5. (a) Geographic location of the Luquillo and Calhoun Experimental Forests. (b) Mean depth and (c) frequency of precipitation events per each month. In Luquillo, rainfall statistics are available in Heartsill-Scalley et al. (2007) and Calabrese and Porporato (2019), while in Calhoun they were obtained combining multiple sources ("<http://criticalzone.org/calhoun/data/datasets/>" and "<https://www.usclimatedata.com/climate/south-carolina/united-states/3210/>"). (d) Monthly average potential evapotranspiration. Values were acquired from the CRU climate dataset (Harris et al., 2014) for Luquillo and from the ORNL DAAC archives data (Vogel & Sankarasubramanian, 2005) for Calhoun.

4 Case studies

4.1 Study areas

The comprehensive hydrological and biogeochemical observations at the tropical forest in Luquillo (Puerto Rico) and at the subtropical forest in Calhoun (South Carolina), which are part of the Critical Zone research network sponsored by the US National Science Foundation, allow us to readily apply the above framework to compare the soil iron dynamics and the potential for iron reduction in these different environments. In Luquillo, we focus on the Bisley watershed, where many Fe cycling studies have been performed. At that site, the mean annual precipitation is about 3.5 m and the vegetation belongs to the Tabonuco forest type (Scatena, 1989). Soils are predominantly Ultisols, formed from volcanic parent material, and belong to the silty clay loam textural class. Calhoun has mean annual precipitation of approximately 1250 mm and vegetation includes mixed hardwood and pine trees. Here soils are also predominantly Ultisols, formed from a granite-gneiss bedrock, and belong to the silt loam textural class (Richter & Markewitz, 2001).

Monthly averaged mean depth and frequency of precipitation as well as potential evapotranspiration for the two sites are illustrated in Figure 5. While Luquillo has a humid tropical climate with only a mild seasonality (slightly reduced rainfall in the winter season), Calhoun has a subtropical climate with marked seasonality in both precipitation and evaporative demand, June and July being the wettest months with also a peak in potential evapotranspiration. Geochemical analysis showed that Luquillo and Calhoun soils have approximately 150 and 45 mmol, respectively, of short-range ordered or low-crystallinity Fe^{III} phases per kilogram of soil (Ginn et al., 2017; Barcellos, Cyle, & Thompson, 2018; Barcellos, 2018). Soil incubation experiments with soil samples from both sites amended with substrate and microbes revealed that reduction rate constants are of the

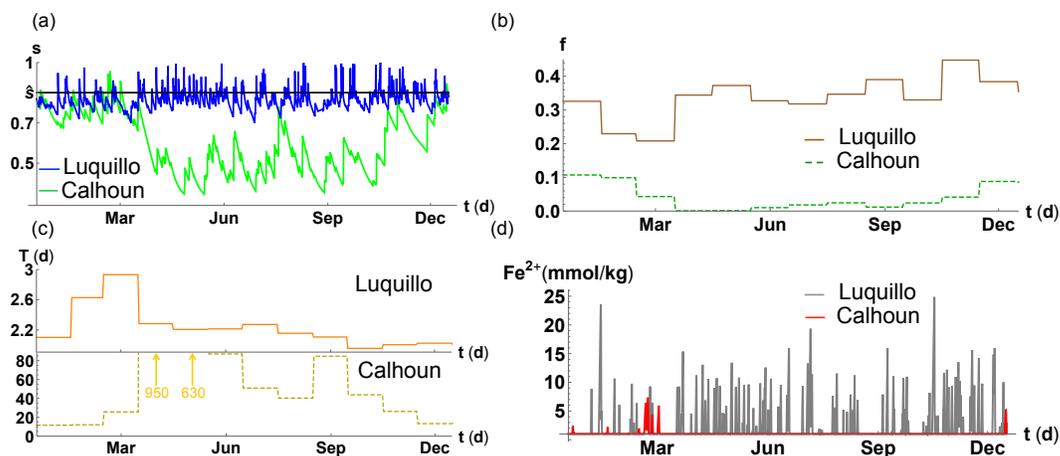


Figure 6. (a) Temporal evolution of soil moisture, simulated by means of the stochastic model in (Laio et al., 2001), in Luquillo (blue line) and Calhoun (green line) over the course of a year. Soils are silty clay loams and silty loams in Luquillo and Calhoun, respectively, with porosity of 0.48. Soil hydrologic properties for the simulation of the soil moisture dynamics from Fernandez-Illescas et al. (2001). Soils are considered to have sufficient anoxic microsites to support Fe reduction for soil moisture levels above $\hat{s} = 0.85$ in Luquillo and $\hat{s} = 0.75$ in Calhoun. (b) Anoxic fraction of the cycle, f , and (c) duration of the cycle, T , for each month computed by means of equations (??) and (??) in the Appendix. (d) Temporal evolution of Fe^{II} in Luquillo (gray line) and Calhoun (red line) over the course of a year, simulated through equation (1). The reduction and oxidation rate constants are $k_R = 0.1$ and $k_O = 10$ mmol/kg/d, respectively.

225 order of 10^{-1} d^{-1} , while the oxidation rate constants at 21% O_2 are of the order of 10
 226 d^{-1} (Chen & Thompson, 2017; Ginn et al., 2017).

227 4.2 Oxic/anoxic cycles and iron reduction

228 To calculate the temporal dynamics of potential iron reduction (when limited only
 229 by the hydrologic regime), we solved equation (1) coupled to a soil water balance that
 230 generates a time series of soil moisture levels based on the frequency and mean depth
 231 of precipitation events (Figure 6). For Luquillo these rainfall statistics are available in
 232 Heartsill-Scalley et al. (2007) and Calabrese and Porporato (2019), while in Calhoun they
 233 were obtained combining multiple sources ("<http://criticalzone.org/calhoun/data/datasets/>"
 234 and "<https://www.usclimatedata.com/climate/south-carolina/united-states/3210>"). The
 235 average anoxic fraction f and cycle length T of the oxic/anoxic cycles are then computed
 236 for each month from the probability density function of soil moisture (see Appendix A).
 237 Note that for each month the parameters f and T are computed assuming stationary
 238 climatic conditions. For each month their values thus correspond to oxic/anoxic cycles
 239 that would occur if the climatic conditions were stationary and typical of that specific
 240 month. As a consequence, it can happen that the value of T is greater than the dura-
 241 tion of the month, e.g., $T = 80$ days in Calhoun in September. Of course, these large
 242 values of T for a particular month only indicate that it is very unlikely to observe full
 243 redox cycles (an Fe oxidation event and an Fe reduction event) in that given month, typ-
 244 ically because soil moisture remains below the threshold set.

245 In Luquillo, the soil moisture frequently crosses the \hat{s} threshold, generating redox
 246 cycles of only a few days (2-3 days) throughout the year (Figure 6(a) and (c)). Similarly,
 247 the calculated anoxic fraction f remains practically constant during the year and approx-
 248 imately equal to 0.3 (Figure 6(b)). The mild seasonality here is almost not visible in the

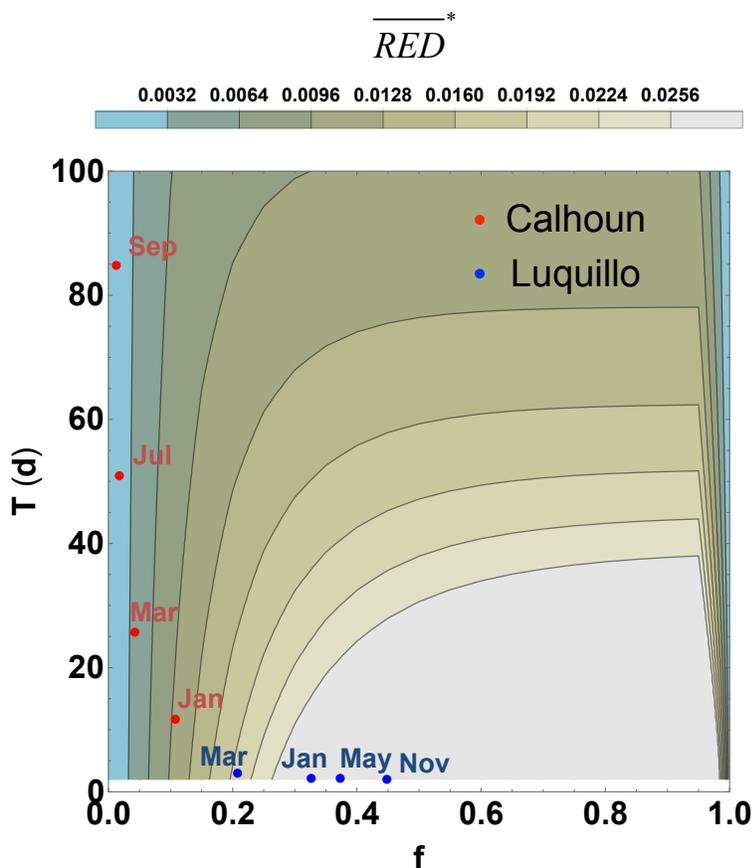


Figure 7. Average and normalized reduction rate per cycle ($\overline{RED}^* = \overline{RED}/Fe^{TOT}$) as a function of the anoxic fraction of the cycle, f , and length of the cycle, T . Corresponding \overline{RED}^* values in different months for Luquillo (blue dots) and Calhoun (red dots). Note that values of T longer than 30 days indicate that a full oxic/anoxic cycle is not expected for that month.

249 iron dynamics. There are continuous heavy rainfall events that bring soil moisture above
 250 the soil moisture threshold \hat{s} , stimulating iron reduction, following which the soil returns
 251 oxic conditions, triggering rapid Fe^{II} oxidation. As a result, throughout the year Fe^{II} dy-
 252 namics appears as a series of rapid redox cycles (a reduction followed by the oxidation)
 253 initiated by heavy precipitation (Figure 6(d)).

254 The estimated temporal patterns of the oxic/anoxic cycles and iron reduction are
 255 rather different in Calhoun. Compared to Luquillo, the full redox cycles in Calhoun are
 256 longer and are affected by the climatic seasonality. Conditions are most favorable for iron
 257 cycling only in the late Fall through the early Spring, when the evaporative demand is
 258 low. In December and January, for example, the ratio f reaches values of 0.1, much lower
 259 than the 0.45 reached in Luquillo in November, while the duration of the cycle reduces
 260 to about 10 days (Figure 6(b) and (c)). This results in only a few, long iron redox cycles
 261 (Figure 6(d)). From the Spring until the following Fall, the climate favors oxic condi-
 262 tions, with f tending to 0 and T reaching values of over 900 days. After the rapid ox-
 263 idation in April/May, iron is in fact likely to remain in its oxidized state until the ar-
 264 rival of the following Fall season (Figure 6(d)).

265 With the above considerations on the different characteristics of the oxic/anoxic
 266 cycles in Luquillo and Calhoun, the pace of iron cycling can be quantified by means of

267 the average reduction rate. To use an index of iron reduction that depends only on the
 268 oxic/anoxic cycle and that can be used to compare the two study sites, we computed the
 269 average reduction rate \overline{RED} , Fe reduced per cycle divided by the duration of the cycle,
 270 and normalized it by the total short range ordered Fe content, $\overline{RED}^* = \overline{RED}/Fe^{TOT}$.
 271 The term \overline{RED}^* is thus analyzed as a function of the fraction in anoxic conditions f and
 272 the overall duration of the cycle T (Figure 7). For the reduction and oxidation rate con-
 273 stants found in these forests, the optimum oxic/anoxic cycle has an anoxic fraction $f \approx$
 274 0.8 and a cycle length T shorter than 10 days. The humid tropical climate in Luquillo
 275 guarantees a high potential for iron reduction throughout the year, with only slightly less
 276 favorable conditions in the spring (blue dots in Figure 5(d)). Differently from Luquillo,
 277 the seasonal climate in Calhoun is largely reflected in the soil redox conditions. As we
 278 have seen above, the Fall until the beginning of the Spring is the period with the high-
 279 est potential for iron reduction. Indeed, January has an almost optimal oxic/anoxic cy-
 280 cle for iron reduction. On the contrary, conditions are far from favorable in the late Spring
 281 and in the summer, when due to lower precipitation and high evapotranspiration, respec-
 282 tively, oxic conditions tend to persist.

283 5 Discussion

284 The high potential of iron reduction predicted here, which is consistent with pre-
 285 vious reported rates (Yang & Liptzin, 2015; Hall et al., 2013; Barcellos, Cyle, & Thomp-
 286 son, 2018), suggests that a large portion of soil organic matter potentially can be decom-
 287 posed through iron reduction rather than aerobic decomposition, making iron fundamen-
 288 tal for the functioning of these ecosystems. In humid tropical forests, such as Luquillo,
 289 iron redox cycles occur throughout the year because the high rainfall can continuously
 290 sustain fluctuations in oxic/anoxic conditions. Experimental (Dubinsky et al., 2010) and
 291 modeling (Calabrese & Porporato, 2019) studies in fact estimated that up to 40% of or-
 292 ganic matter decomposition could be attributed to iron reduction. In subtropical forests,
 293 such as Calhoun in the Southeastern Piedmont in USA, the marked climatic seasonal-
 294 ity greatly controls iron reduction, which is favored only in those months with higher rain-
 295 fall and lower evaporative demand. This finding is supported by recent experiments in
 296 Calhoun, where potential of iron reduction was measured over the course of a year by
 297 means of steel IRIS (Indicator of Reduction of Iron in Soils) probes (Hodges et al., 2019).

298 Other than impacting directly the carbon cycle, iron redox cycling also affects the
 299 productivity of plants (Colombo et al., 2014; Calabrese & Porporato, 2019), because iron
 300 is a micro-nutrient essential for plants to support their physiological processes (e.g., pho-
 301 tosynthesis) and is directly related to the bioavailability in soils of phosphorous, which
 302 tends to be adsorbed to iron oxides with high surface area (Miller et al., 2001; Chacon
 303 et al., 2006; Gross et al., 2018; E. M. Herndon et al., 2019; Khan et al., 2019). It is thus
 304 the in-situ hydro-climate that controls the availability of these two nutrients. Wet con-
 305 ditions favor iron reduction and thus more Fe^{II} and P in the soil solution. If conditions
 306 are constantly wet, however, iron may almost completely dissolve in water, causing plants
 307 to uptake large amounts of iron that result toxic to their cells (Foy et al., 1978). Dry con-
 308 ditions, on the contrary, make these nutrients unavailable because they favor iron oxi-
 309 dation, with formation of iron oxyhydroxides (Fe^{III}), on which phosphorous can be strongly
 310 adsorbed. Interestingly, plants are able to cope with iron deficient or toxic conditions
 311 by affecting, for example, the redox potential in the rhizosphere (Guerinot & Yi, 1994;
 312 Rout & Sahoo, 2015), thus exerting a feedback on the iron redox dynamics. Variations
 313 in rainfall frequency and depth, resulting from climate changes, call for modeling frame-
 314 works able to quantify the coupled soil iron-plants dynamics, which is of interest not only
 315 to predict the future response of natural ecosystems, but also to refine irrigation and fer-
 316 tilization strategies in agroecosystems, to limit the utilization of resources while guar-
 317 anteeing food security (FAO, 2015).

318 A deeper understanding of iron cycling is required also to reduce the uncertainty
 319 in the carbon budget, particularly in tropical and temperate forests (Bailey et al., 2018;
 320 Kramer & Chadwick, 2018). The former contain some 20% of global soil carbon and have
 321 among the highest rates of greenhouse gas emissions (Jobbágy & Jackson, 2000; Malhi
 322 & Grace, 2000), while the latter (e.g., the eastern USA) have high potential for soil and
 323 plant carbon storage, given that these forests have been recovering after historically be-
 324 ing deforested (Bonan, 2008). Because iron cycling in upland soils is driven by the wet-
 325 ting and drying of soils upon the intermittent arrival of precipitation, for a realistic and
 326 more accurate description of these dynamics, Earth system and climate models need to
 327 account for the temporal oxic/anoxic transitions driven by hydro-climatic fluctuations,
 328 especially when investigating future soil carbon storage and its feedback on the long-term
 329 climate.

330 6 Conclusions

331 Through the minimalist model of iron redox dynamics presented here, we relate
 332 the potential rates of iron reduction to the hydro-climatic variability through its influ-
 333 ence on changes in soil moisture and predicted oxic and anoxic conditions. The study
 334 showed that hydro-climatic variability may favor or inhibit iron reduction, depending
 335 on how closely the resulting oxic/anoxic cycle approaches the ‘optimal’ oxic/anoxic cy-
 336 cle. By relating the rate of the soil iron cycle to hydro-climatic fluctuations, this anal-
 337 ysis also paves the way for a global identification of hot spots of iron reduction, in which
 338 climatic features are highly favorable, and for prediction of future trends in organic mat-
 339 ter decomposition. We also believe that these results represent an important step toward
 340 an improved representation of biogeochemical processes, especially anaerobic processes
 341 (e.g., Zheng et al. (2019) and Calabrese and Porporato (2019)), in Earth system mod-
 342 els (e.g., Hurrell et al. (2013)).

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347 References

- 348 Bailey, V. L., Bond-Lamberty, B., DeAngelis, K., Grandy, A. S., Hawkes, C. V.,
 349 Heckman, K., . . . others (2018). Soil carbon cycling proxies: Understand-
 350 ing their critical role in predicting climate change feedbacks. *Global change*
 351 *biology*, *24*(3), 895–905.
- 352 Barcellos, D. (2018). *Biogeochemical cycling of iron and carbon in humid (sub)*
 353 *tropical forest soils under fluctuating redox conditions* (Unpublished doctoral
 354 dissertation). University of Georgia.
- 355 Barcellos, D., Cyle, K. T., & Thompson, A. (2018). Faster redox fluctuations can
 356 lead to higher iron reduction rates in humid forest soils. *Biogeochemistry*,
 357 *137*(3), 367–378.
- 358 Barcellos, D., OConnell, C., Silver, W., Meile, C., & Thompson, A. (2018). Hot
 359 spots and hot moments of soil moisture explain fluctuations in iron and carbon
 360 cycling in a humid tropical forest soil. *Soil Systems*, *2*(4), 59.
- 361 Bhattacharyya, A., Campbell, A. N., Tfaily, M. M., Lin, Y., Kukkadapu, R. K., Sil-
 362 ver, W. L., . . . Pett-Ridge, J. (2018). Redox fluctuations control the coupled
 363 cycling of iron and carbon in tropical forest soils. *Environmental science &*
 364 *technology*, *52*(24), 14129–14139.
- 365 Bishop, M. E., Glasser, P., Dong, H., Arey, B., & Kovarik, L. (2014). Reduction and
 366 immobilization of hexavalent chromium by microbially reduced Fe-bearing clay

- 367 minerals. *Geochimica et Cosmochimica Acta*, 133, 186–203.
- 368 Bonan, G. B. (2008). Forests and climate change: forcings, feedbacks, and the cli-
369 mate benefits of forests. *science*, 320(5882), 1444–1449.
- 370 Borch, T., Kretzschmar, R., Kappler, A., Cappellen, P. V., Ginder-Vogel, M.,
371 Voegelin, A., & Campbell, K. (2009). Biogeochemical redox processes and
372 their impact on contaminant dynamics. *Environmental science & technology*,
373 44(1), 15–23.
- 374 Brady, N. C., & Weil, R. R. (2016). *The nature and properties of soils*. Pearson.
- 375 Buettner, S. W., Kramer, M. G., Chadwick, O. A., & Thompson, A. (2014). Mo-
376 bilization of colloidal carbon during iron reduction in basaltic soils. *Geoderma*,
377 221, 139–145.
- 378 Calabrese, S., & Porporato, A. (2019). Impact of ecohydrological fluctuations on
379 iron-redox cycling. *Soil Biology and Biochemistry*.
- 380 Chacon, N., Silver, W. L., Dubinsky, E. A., & Cusack, D. F. (2006). Iron reduction
381 and soil phosphorus solubilization in humid tropical forests soils: the roles of
382 labile carbon pools and an electron shuttle compound. *Biogeochemistry*, 78(1),
383 67–84.
- 384 Chen, C., & Thompson, A. (2017). Ferrous iron oxidation under varying po2 levels:
385 the effect of fe (iii)/al (iii) oxide minerals and organic matter. *Environmental*
386 *science & technology*, 52(2), 597–606.
- 387 Colombo, C., Palumbo, G., He, J.-Z., Pinton, R., & Cesco, S. (2014). Review on iron
388 availability in soil: interaction of fe minerals, plants, and microbes. *Journal of*
389 *Soils and Sediments*, 14(3), 538–548.
- 390 Couture, R.-M., Charlet, L., Markelova, E., Made, B., & Parsons, C. T. (2015). On-
391 off mobilization of contaminants in soils during redox oscillations. *Environmental*
392 *science & technology*, 49(5), 3015–3023.
- 393 Dubinsky, E. A., Silver, W. L., & Firestone, M. K. (2010). Tropical forest soil mi-
394 crobial communities couple iron and carbon biogeochemistry. *Ecology*, 91(9),
395 2604–2612.
- 396 FAO, I. (2015). Status of the worlds soil resources (swsr)—main report. *Food and*
397 *agriculture organization of the United Nations and intergovernmental technical*
398 *panel on soils, Rome, Italy*, 650.
- 399 Fernandez-Illescas, C. P., Porporato, A., Laio, F., & Rodriguez-Iturbe, I. (2001).
400 The ecohydrological role of soil texture in a water-limited ecosystem. *Water*
401 *Resources Research*, 37(12), 2863–2872.
- 402 Foy, C., Chaney, R. t., & White, M. (1978). The physiology of metal toxicity in
403 plants. *Annual review of plant physiology*, 29(1), 511–566.
- 404 Ginn, B., Meile, C., Wilmoth, J., Tang, Y., & Thompson, A. (2017). Rapid iron re-
405 duction rates are stimulated by high-amplitude redox fluctuations in a tropical
406 forest soil. *Environmental Science & Technology*, 51(6), 3250–3259.
- 407 Gross, A., Pett-Ridge, J., & Silver, W. (2018). Soil oxygen limits microbial phospho-
408 rus utilization in humid tropical forest soils. *Soil Systems*, 2(4), 65.
- 409 Guerinot, M. L., & Yi, Y. (1994). Iron: nutritious, noxious, and not readily avail-
410 able. *Plant Physiology*, 104(3), 815.
- 411 Hall, S. J., McDowell, W. H., & Silver, W. L. (2013). When wet gets wetter: decou-
412 pling of moisture, redox biogeochemistry, and greenhouse gas fluxes in a humid
413 tropical forest soil. *Ecosystems*, 16(4), 576–589.
- 414 Han, L., Sun, K., Keiluweit, M., Yang, Y., Yang, Y., Jin, J., . . . Xing, B. (2019).
415 Mobilization of ferrihydrite-associated organic carbon during fe reduction:
416 Adsorption versus coprecipitation. *Chemical Geology*, 503, 61–68.
- 417 Harris, I., Jones, P. D., Osborn, T. J., & Lister, D. H. (2014). Updated high-
418 resolution grids of monthly climatic observations—the cru ts3. 10 dataset.
419 *International journal of climatology*, 34(3), 623–642.
- 420 Heartsill-Scalley, T., Scatena, F. N., Estrada, C., McDowell, W., & Lugo, A. E.
421 (2007). Disturbance and long-term patterns of rainfall and throughfall nu-

- 422 trient fluxes in a subtropical wet forest in puerto rico. *Journal of Hydrology*,
 423 *333*(2-4), 472–485.
- 424 Henderson, R., Kabengi, N., Mantripragada, N., Cabrera, M., Hassan, S., & Thomp-
 425 son, A. (2012). Anoxia-induced release of colloid-and nanoparticle-bound
 426 phosphorus in grassland soils. *Environmental science & technology*, *46*(21),
 427 11727–11734.
- 428 Herndon, E., AlBashaireh, A., Singer, D., Chowdhury, T. R., Gu, B., & Graham, D.
 429 (2017). Influence of iron redox cycling on organo-mineral associations in arctic
 430 tundra soil. *Geochimica et Cosmochimica Acta*, *207*, 210–231.
- 431 Herndon, E. M., Kinsman-Costello, L., Duroe, K. A., Mills, J., Kane, E. S.,
 432 Sebestyen, S. D., . . . Wullschleger, S. D. (2019). Iron (oxyhydr) oxides serve
 433 as phosphate traps in tundra and boreal peat soils. *Journal of Geophysical*
 434 *Research: Biogeosciences*, *124*(2), 227–246.
- 435 Hodges, C., Mallard, J., Markewitz, D., Barcellos, D., & Thompson, A. (2019).
 436 Seasonal and spatial variation in the potential for iron reduction in soils of the
 437 southeastern piedmont of the us. *Catena*, *180*, 32–40.
- 438 Hurrell, J. W., Holland, M. M., Gent, P. R., Ghan, S., Kay, J. E., Kushner, P. J.,
 439 . . . others (2013). The community earth system model: a framework for col-
 440 laborative research. *Bulletin of the American Meteorological Society*, *94*(9),
 441 1339–1360.
- 442 Jobbágy, E. G., & Jackson, R. B. (2000). The vertical distribution of soil organic
 443 carbon and its relation to climate and vegetation. *Ecological applications*,
 444 *10*(2), 423–436.
- 445 Khan, I., Fahad, S., Wu, L., Zhou, W., Xu, P., Sun, Z., . . . others (2019). Labile
 446 organic matter intensifies phosphorous mobilization in paddy soils by microbial
 447 iron (iii) reduction. *Geoderma*, *352*, 185–196.
- 448 Kramer, M. G., & Chadwick, O. A. (2018). Climate-driven thresholds in reactive
 449 mineral retention of soil carbon at the global scale. *Nature Climate Change*,
 450 *8*(12), 1104.
- 451 LaCroix, R. E., Tfaily, M. M., McCreight, M., Jones, M. E., Spokas, L., & Keiluweit,
 452 M. (2019). Shifting mineral and redox controls on carbon cycling in seasonally
 453 flooded mineral soils. *Biogeosciences*, *16*(13), 2573–2589.
- 454 Laio, F., Porporato, A., Ridolfi, L., & Rodriguez-Iturbe, I. (2001). Plants in water-
 455 controlled ecosystems: active role in hydrologic processes and response to
 456 water stress: Ii. probabilistic soil moisture dynamics. *Advances in Water*
 457 *Resources*, *24*(7), 707–723.
- 458 LaRowe, D. E., & Van Cappellen, P. (2011). Degradation of natural organic matter:
 459 a thermodynamic analysis. *Geochimica et Cosmochimica Acta*, *75*(8), 2030–
 460 2042.
- 461 Lovley, D. R. (1991). Dissimilatory fe (iii) and mn (iv) reduction. *Microbiological re-*
 462 *views*, *55*(2), 259–287.
- 463 Malhi, Y., & Grace, J. (2000). Tropical forests and atmospheric carbon dioxide.
 464 *Trends in Ecology & Evolution*, *15*(8), 332–337.
- 465 Miller, A. J., Schuur, E. A., & Chadwick, O. A. (2001). Redox control of phosphorus
 466 pools in hawaiian montane forest soils. *Geoderma*, *102*(3-4), 219–237.
- 467 Morel, F. M., Hering, J. G., et al. (1993). *Principles and applications of aquatic*
 468 *chemistry*. John Wiley & Sons.
- 469 Oertel, C., Matschullat, J., Zurba, K., Zimmermann, F., & Erasmi, S. (2016).
 470 Greenhouse gas emissions from soilsa review. *Chemie der Erde-Geochemistry*,
 471 *76*(3), 327–352.
- 472 Richter, D. D., & Markewitz, D. (2001). Understanding soil change. *Understanding*
 473 *Soil Change*, by Daniel D. Richter, Jr and Daniel Markewitz and Foreword
 474 by William A. Reiners and Pedro Sánchez, pp. 272. ISBN 0521771714. Cam-
 475 bridge, UK: Cambridge University Press, June 2001., 1.
- 476 Roden, E. E., & Wetzal, R. G. (1996). Organic carbon oxidation and suppression of

- 477 methane production by microbial Fe (iii) oxide reduction in vegetated and un-
478 vegetated freshwater wetland sediments. *Limnology and Oceanography*, 41(8),
479 1733–1748.
- 480 Rout, G. R., & Sahoo, S. (2015). Role of iron in plant growth and metabolism. *Re-*
481 *views in Agricultural Science*, 3, 1–24.
- 482 Scatena, F. N. (1989). An introduction to the physiography and history of the
483 bisley experimental watersheds in the Luquillo mountains of Puerto Rico. *Gen.*
484 *Tech. Rep. SO-72. New Orleans, LA: US Dept of Agriculture, Forest Service,*
485 *Southern Forest Experiment Station. 22 p., 72.*
- 486 Stucki, J. W. (2011). A review of the effects of iron redox cycles on smectite proper-
487 ties. *Comptes Rendus Geoscience*, 343(2-3), 199–209.
- 488 Thompson, A., Chadwick, O. A., Rancourt, D. G., & Chorover, J. (2006). Iron-oxide
489 crystallinity increases during soil redox oscillations. *Geochimica et Cosmochim-*
490 *ica Acta*, 70(7), 1710–1727.
- 491 Todd-Brown, K. E., Hopkins, F. M., Kivlin, S. N., Talbot, J. M., & Allison, S. D.
492 (2012). A framework for representing microbial decomposition in coupled
493 climate models. *Biogeochemistry*, 109(1-3), 19–33.
- 494 Vermeire, M.-L., Bonneville, S., Stenuit, B., Delvaux, B., & Cornélis, J.-T. (2019). Is
495 microbial reduction of Fe (iii) in podzolic soils influencing C release? *Geoderma*,
496 340, 1–10.
- 497 Vogel, R., & Sankarasubramanian, A. (2005). *Monthly climate data for selected*
498 *USGS HCDN sites, 1951-1990, r1*. ORNL Distributed Active Archive Center. Re-
499 trieved from http://daac.ornl.gov/cgi-bin/dsviewer.pl?ds_id=810 doi:
500 10.3334/ORNLDAAAC/810
- 501 Wang, Y., Liu, C., Peng, A., & Gu, C. (2019). Formation of hydroxylated poly-
502 chlorinated diphenyl ethers mediated by structural Fe (iii) in smectites. *Chemo-*
503 *sphere*, 226, 94–102.
- 504 Weiss, J. V., Emerson, D., & Megonigal, J. P. (2004). Geochemical control of micro-
505 bial Fe (iii) reduction potential in wetlands: comparison of the rhizosphere to
506 non-rhizosphere soil. *FEMS Microbiology Ecology*, 48(1), 89–100.
- 507 Weiss, J. V., Emerson, D., & Megonigal, J. P. (2005). Rhizosphere iron (iii) deposi-
508 tion and reduction in a l-dominated wetland. *Soil Science Society of America*
509 *Journal*, 69(6), 1861–1870.
- 510 Yang, W. H., & Liptzin, D. (2015). High potential for iron reduction in upland soils.
511 *Ecology*, 96(7).
- 512 Yu, H.-Y., Li, F.-B., Liu, C.-S., Huang, W., Liu, T.-X., & Yu, W.-M. (2016). Iron
513 redox cycling coupled to transformation and immobilization of heavy metals:
514 implications for paddy rice safety in the red soil of south China. In *Advances in*
515 *agronomy* (Vol. 137, pp. 279–317). Elsevier.
- 516 Zheng, J., Thornton, P. E., Painter, S. L., Gu, B., Wulfschleger, S. D., & Graham,
517 D. E. (2019). Modeling anaerobic soil organic carbon decomposition in arctic
518 polygon tundra: insights into soil geochemical influences on carbon mineraliza-
519 tion. *Biogeosciences*, 16(3), 663–680.