# Response Manipulation vs Diurnal Observation: A Direct Comparison of Two Methods for Quantifying Stomatal Behavior in Tropical Forests

Kenneth J. Davidson, Justin Lawson, Shawn P. Serbin, Qianyu Li, & Alistair Rogers

Brookhaven National Laboratory, Environmental Plant Science Sciences Department, Upton, NY, United States

Stony Brook University, Department of Ecology and Evolution, Stony Brook, NY, United States

Abstract #

AGU FALL MEETING
New Orleans | 11-15 December 2023

## BACKGROUND

**Terrestrial Land Biosphere**

One of the largest uncertainties in current model predictions of plant carbon uptake and water cycling is how stomatal conductance (*g*$_s$) responds to changes in environmental conditions (Medlyn et al. 2011). Most models of *g*$_s$ are based on the optimization hypothesis that stomata maximize the amount of carbon gained per unit water lost (Cowan & Farquhar 1977). The most widely used model is the Medlyn et al. (2011) model, which has the form:

$$g_s = g_0 + 1.6\left(1 + \frac{g_1}{\sqrt{D}}\right)\frac{A}{C_a}$$

In this model, *g*$_0$ is residual stem flux and can be set to 0 for most work on the scale of a plot or canopy because flux rates are so small relative to the other fluxes (Wolfe et al. 2016). *A* is net photosynthesis, *C*$_a$ is the ambient CO$_2$ concentration, and *D* is vapor pressure deficit. The slope parameter *g*$_1$ is the only parameter that must be fitted to data and is proportional to the ratio of the marginal water cost of carbon gain to the marginal carbon cost of water loss (Wolf et al. 2016).

**Prevailing view on stomatal effects on climate**

Terrestrial plants take up carbon dioxide from the atmosphere for photosynthesis and release water vapor through stomata. Stomatal conductance (*g*$_s$) is the rate at which water vapor and CO$_2$ diffuse through stomata. Stomatal conductance is a key component of the water balance of plants and ecosystems, and it is also a key component of the carbon balance of plants and ecosystems.

**Stomatal changes interpretation?**

Traditionally, leaf-level stomatal conductance (*g*$_s$) is measured using gas exchange systems, such as the LI-6800 (LI-COR, Inc., Lincoln, NE, USA). These systems measure the rate of water vapor and CO$_2$ exchange between a leaf and the atmosphere. Stomatal conductance is then calculated from these measurements. Gas exchange systems can provide accurate measurements of *g*$_s$, but they can be time-consuming and labor-intensive. Additionally, they can only measure *g*$_s$ on a few leaves at a time, which can limit the representativeness of the measurements.

**Objective**

The objective of this research was to compare the response to stomatal conductance (*g*$_s$) to changes in environmental conditions between two methods:

1. Diurnal time course, one-point, leaf-level gas exchange measurements (LI-6800 with 6800-01A chamber)
2. Response curves, leaf-level gas exchange measurements (LI-6800 with 6800-01A chamber)
3. Time-integrated stomatal conductance (*g*$_s$) measurements, using carbon isotope discrimination ($\Delta^{13}C$) and leaf gas exchange measurements

## METHODS

**Study Site.** The study was conducted at the Brookhaven National Laboratory Forest Research Plot (BNL FRP) (40° 52' 0.12" N, 72° 57' 0.12" W) in the Pine Barrens of the Northeastern US (Hardwood-Conifer) forest (Fig. 1). Vegetation is a mixture of oak-hickory forest, the dominant species are *Quercus velutina* (black oak), *Quercus alba* (white oak), *Carya glabra* (pignut hickory), and *Pinus rigida* (pitch pine). The soil is a sandy loam (Carlson).

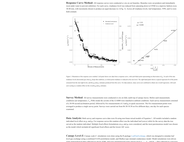


Figure 1. Location of study site in the Pine Barrens of the Northeastern US.

**Canopy Access.** The Brookhaven National Laboratory Forest Research Plot (BNL FRP) (40° 52' 0.12" N, 72° 57' 0.12" W) in the Pine Barrens of the Northeastern US (Hardwood-Conifer) forest (Fig. 1). Vegetation is a mixture of oak-hickory forest, the dominant species are *Quercus velutina* (black oak), *Quercus alba* (white oak), *Carya glabra* (pignut hickory), and *Pinus rigida* (pitch pine). The soil is a sandy loam (Carlson).

**Study Species.** Six trees were selected for each site, and two sun-exposed canopy leaves per tree were measured (Figure 2). For each tree, the diameter at breast height (DBH) was measured, and the height was measured using a laser rangefinder.

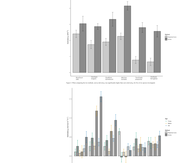


Figure 2 Sampled trees from the study site. Trees were selected to represent the range of diameters at breast height (DBH) and canopy positions (dominant, co-dominant, intermediate, and suppressed) found in the stand. Two sun-exposed canopy leaves were measured per tree.

**Gas Exchange.** Leaf gas exchange was conducted using the LI-6800 (LI-COR) and 6800-01A chamber system (LI-COR Biosciences, Lincoln, NE, USA). All gas exchange was conducted using ambient CO$_2$ (~400).

**Response Curve Methods.** All response curves were conducted on sun-exposed canopy leaves. Response curves were conducted using the following sequence of reference CO$_2$ concentrations: 400, 300, 200, 150, 100, 400, 400, 500, 600, 700, 800, 900, 1000, 1100, 1200, 1300, 1400, 1500, 1600, 1700, 1800, 1900, 2000, 2100, 2200, 2300, 2400, 2500. The response curves were conducted at a leaf temperature of 25 °C, a flow rate of 500 µmol s$^{-1}$, and a relative humidity of 50%.

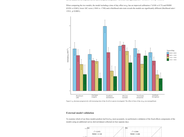


Figure 3 Example response curve of net photosynthesis (*A*) and stomatal conductance (*g*$_s$) to changes in reference CO$_2$ concentration. The response curve was conducted on a sun-exposed canopy leaf of a co-dominant tree. The response curve was conducted at a leaf temperature of 25 °C, a flow rate of 500 µmol s$^{-1}$, and a relative humidity of 50%.

**Stomatal Method.** Stomatal conductance was measured on both sides of each leaf (Hypo- Hemi and Amphistomatous leaves). Stomatal conductance was measured at 1-second intervals for 5 minutes. Stomatal conductance was averaged over the last 30 seconds of the measurement.

**Flow Analysis.** Flow rate was measured after each time point of the response curve. Flow rate was measured at 1-second intervals for 30 seconds. Multiple flow rate measurements were averaged to obtain a single flow rate value for each time point.

**Carbon Isotope Discrimination.** Leaf carbon isotope discrimination ($\Delta^{13}C$) was measured using the LI-6800 system. The $\Delta^{13}C$ was calculated using the following equation:

$$\Delta^{13}C = \frac{\xi(\delta_{13}C_{out} - \delta_{13}C_{in})}{\delta_{13}C_{in} + 1000}$$

where $\xi$ is the ratio of the flow rate of CO$_2$ in the sample cell to the flow rate of CO$_2$ in the reference cell, $\delta_{13}C_{out}$ is the $\delta^{13}C$ of the CO$_2$ in the sample cell, and $\delta_{13}C_{in}$ is the $\delta^{13}C$ of the CO$_2$ in the reference cell.

**Time Flow.** Flow rate was measured after each time point of the response curve. Flow rate was measured at 1-second intervals for 30 seconds. Multiple flow rate measurements were averaged to obtain a single flow rate value for each time point.

## STOMATAL MODEL RESULTS

**Comparison of response curve and time-of-day stomatal parameters**

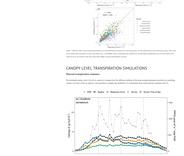


Figure 4 Comparison of stomatal conductance ($g_s$) and net photosynthesis ($A$) measured using response curves and time-of-day measurements. The response curve was conducted on a sun-exposed canopy leaf of a co-dominant tree. The response curve was conducted at a leaf temperature of 25 °C, a flow rate of 500 µmol s$^{-1}$, and a relative humidity of 50%. The time-of-day measurements were conducted on the same leaf as the response curve.

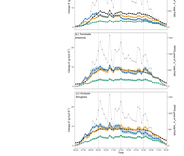


Figure 5 Comparison of stomatal conductance ($g_s$) and net photosynthesis ($A$) measured using response curves and time-of-day measurements. The response curve was conducted on a sun-exposed canopy leaf of a co-dominant tree. The response curve was conducted at a leaf temperature of 25 °C, a flow rate of 500 µmol s$^{-1}$, and a relative humidity of 50%. The time-of-day measurements were conducted on the same leaf as the response curve.

**Investigation of a law of day effect**

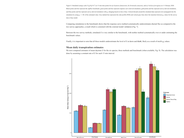


Figure 6 Comparison of stomatal conductance ($g_s$) and net photosynthesis ($A$) measured using response curves and time-of-day measurements. The response curve was conducted on a sun-exposed canopy leaf of a co-dominant tree. The response curve was conducted at a leaf temperature of 25 °C, a flow rate of 500 µmol s$^{-1}$, and a relative humidity of 50%. The time-of-day measurements were conducted on the same leaf as the response curve.

**Stomatal model validation**

Figure 7 Comparison of stomatal conductance ($g_s$) and net photosynthesis ($A$) measured using response curves and time-of-day measurements. The response curve was conducted on a sun-exposed canopy leaf of a co-dominant tree. The response curve was conducted at a leaf temperature of 25 °C, a flow rate of 500 µmol s$^{-1}$, and a relative humidity of 50%. The time-of-day measurements were conducted on the same leaf as the response curve.

## CANOPY LEVEL TRANSPIRATION SIMULATIONS

**Diurnal transpiration estimates**

Figure 8 Comparison of stomatal conductance ($g_s$) and net photosynthesis ($A$) measured using response curves and time-of-day measurements. The response curve was conducted on a sun-exposed canopy leaf of a co-dominant tree. The response curve was conducted at a leaf temperature of 25 °C, a flow rate of 500 µmol s$^{-1}$, and a relative humidity of 50%. The time-of-day measurements were conducted on the same leaf as the response curve.

Figure 9 Comparison of stomatal conductance ($g_s$) and net photosynthesis ($A$) measured using response curves and time-of-day measurements. The response curve was conducted on a sun-exposed canopy leaf of a co-dominant tree. The response curve was conducted at a leaf temperature of 25 °C, a flow rate of 500 µmol s$^{-1}$, and a relative humidity of 50%. The time-of-day measurements were conducted on the same leaf as the response curve.

**Seasonal model validation**

Figure 10 Comparison of stomatal conductance ($g_s$) and net photosynthesis ($A$) measured using response curves and time-of-day measurements. The response curve was conducted on a sun-exposed canopy leaf of a co-dominant tree. The response curve was conducted at a leaf temperature of 25 °C, a flow rate of 500 µmol s$^{-1}$, and a relative humidity of 50%. The time-of-day measurements were conducted on the same leaf as the response curve.

## CANOPY LEVEL TRANSPIRATION SIMULATIONS

**Diurnal transpiration estimates**

Figure 11 Comparison of stomatal conductance ($g_s$) and net photosynthesis ($A$) measured using response curves and time-of-day measurements. The response curve was conducted on a sun-exposed canopy leaf of a co-dominant tree. The response curve was conducted at a leaf temperature of 25 °C, a flow rate of 500 µmol s$^{-1}$, and a relative humidity of 50%. The time-of-day measurements were conducted on the same leaf as the response curve.

Figure 12 Comparison of stomatal conductance ($g_s$) and net photosynthesis ($A$) measured using response curves and time-of-day measurements. The response curve was conducted on a sun-exposed canopy leaf of a co-dominant tree. The response curve was conducted at a leaf temperature of 25 °C, a flow rate of 500 µmol s$^{-1}$, and a relative humidity of 50%. The time-of-day measurements were conducted on the same leaf as the response curve.

## CONCLUSIONS AND DISCUSSION

**Summary of Key results**

1. Photosynthesis was the most sensitive to stomatal effects (Fig. 3), but the stomatal impact on plant water-use efficiency was relatively small (Fig. 4).
2. Stomatal limitations to photosynthesis were relatively small (Fig. 5), and the stomatal impact on plant water-use efficiency was relatively small (Fig. 6).
3. Differences in the diurnal course of stomatal conductance between the two methods (Fig. 7) were due to the stomatal response to changes in vapor pressure deficit (*D*).
4. Changes in the stomatal response to *D* (*g*$_1$) were the largest source of error in the stomatal models (Fig. 8).
5. Using a single value for *g*$_1$ for the entire growing season (Fig. 9) resulted in large errors in the stomatal models.
6. Some leaf temperatures were too cool for a stomatal conductance. Since the air pressure is very low, this can be a problem for the LI-6800 system.

**Conclusions**

1. Stomatal conductance is sensitive to current conditions but is relatively insensitive to past conditions.
2. Leaf-level gas exchange measurements can be used to estimate canopy-level transpiration.
3. The stomatal model used here is the Medlyn et al. (2011) model, which is widely used in land surface models.

**Lack of Significant *g*$_1$ Correlation**

Stomatal conductance (*g*$_s$) is a measure of the rate of water vapor diffusion through stomata. The Medlyn et al. (2011) model is a widely used model of *g*$_s$ that relates *g*$_s$ to the net photosynthesis rate (*A*), the vapor pressure deficit (*D*), and the slope parameter *g*$_1$. The slope parameter *g*$_1$ is proportional to the ratio of the marginal water cost of carbon gain to the marginal carbon cost of water loss. The lack of significant correlation between *g*$_1$ and leaf gas exchange parameters in this study suggests that other factors may be more important in determining stomatal conductance in the species and environment studied here. It is also possible that the relationship between *g*$_1$ and leaf gas exchange parameters is more complex than a simple linear correlation. For example, the relationship may be nonlinear or it may be affected by other factors, such as leaf temperature or humidity.

**Differences in key functional methods**

The Medlyn et al. (2011) model of stomatal conductance (*g*$_s$) is a widely used model that relates *g*$_s$ to the net photosynthesis rate (*A*), the vapor pressure deficit (*D*), and the slope parameter *g*$_1$. The slope parameter *g*$_1$ is proportional to the ratio of the marginal water cost of carbon gain to the marginal carbon cost of water loss. The lack of significant correlation between *g*$_1$ and leaf gas exchange parameters in this study suggests that other factors may be more important in determining stomatal conductance in the species and environment studied here. It is also possible that the relationship between *g*$_1$ and leaf gas exchange parameters is more complex than a simple linear correlation. For example, the relationship may be nonlinear or it may be affected by other factors, such as leaf temperature or humidity.

**Sensitivities that drive flux**

Stomatal conductance (*g*$_s$) is a measure of the rate of water vapor diffusion through stomata. The Medlyn et al. (2011) model is a widely used model of *g*$_s$ that relates *g*$_s$ to the net photosynthesis rate (*A*), the vapor pressure deficit (*D*), and the slope parameter *g*$_1$. The slope parameter *g*$_1$ is proportional to the ratio of the marginal water cost of carbon gain to the marginal carbon cost of water loss. The lack of significant correlation between *g*$_1$ and leaf gas exchange parameters in this study suggests that other factors may be more important in determining stomatal conductance in the species and environment studied here. It is also possible that the relationship between *g*$_1$ and leaf gas exchange parameters is more complex than a simple linear correlation. For example, the relationship may be nonlinear or it may be affected by other factors, such as leaf temperature or humidity.

**Limits to method due to implications for canopy scale models**

Stomatal conductance (*g*$_s$) is a measure of the rate of water vapor diffusion through stomata. The Medlyn et al. (2011) model is a widely used model of *g*$_s$ that relates *g*$_s$ to the net photosynthesis rate (*A*), the vapor pressure deficit (*D*), and the slope parameter *g*$_1$. The slope parameter *g*$_1$ is proportional to the ratio of the marginal water cost of carbon gain to the marginal carbon cost of water loss. The lack of significant correlation between *g*$_1$ and leaf gas exchange parameters in this study suggests that other factors may be more important in determining stomatal conductance in the species and environment studied here. It is also possible that the relationship between *g*$_1$ and leaf gas exchange parameters is more complex than a simple linear correlation. For example, the relationship may be nonlinear or it may be affected by other factors, such as leaf temperature or humidity.

This presentation by Dr. Davidson Lawson

## DISCLOSURES

**Publication Disclosures**

**Publication Disclosures**

**Publication Disclosures**

## AUTHOR INFORMATION

Kenneth J. Davidson [1], Justin Lawson [1], Shawn P. Serbin [1], Qianyu Li [1], & Alistair Rogers [1]

[1] Brookhaven National Laboratory, Environmental Plant Science Sciences Department, Upton, NY, United States

[2] Stony Brook University, Department of Ecology and Evolution, Stony Brook, NY, United States

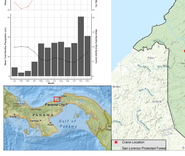
Presenting author's email address: kdavidson@bnl.gov

## ABSTRACT

Traditionally, leaf-level stomatal conductance (*g*$_s$) is measured using gas exchange systems, such as the LI-6800 (LI-COR, Inc., Lincoln, NE, USA). These systems measure the rate of water vapor and CO$_2$ exchange between a leaf and the atmosphere. Stomatal conductance is then calculated from these measurements. Gas exchange systems can provide accurate measurements of *g*$_s$, but they can be time-consuming and labor-intensive. Additionally, they can only measure *g*$_s$ on a few leaves at a time, which can limit the representativeness of the measurements.

An alternative approach is to use time-integrated measurements of stomatal conductance. One such method is the carbon isotope discrimination ($\Delta^{13}C$) method. This method uses the fact that the ratio of $^{13}C$ to $^{12}C$ in plant tissue is affected by the stomatal conductance. The $\Delta^{13}C$ method can provide a more integrated measure of stomatal conductance over time, but it can also be more complex and expensive than gas exchange systems.

The objective of this study was to compare the response to stomatal conductance (*g*$_s$) to changes in environmental conditions between two methods: diurnal time course, one-point, leaf-level gas exchange measurements (LI-6800 with 6800-01A chamber) and response curves, leaf-level gas exchange measurements (LI-6800 with 6800-01A chamber). The study was conducted at the Brookhaven National Laboratory Forest Research Plot (BNL FRP) (40° 52' 0.12" N, 72° 57' 0.12" W) in the Pine Barrens of the Northeastern US (Hardwood-Conifer) forest. The results will be presented and discussed at the conference.

## REFERENCES

1. Ainsley, M. C., Medlyn, B. E., Pendall, E., Ellsworth, D. S., Barton, C. V. M., Kruger, E. L., Duursma, R. A. (2011). Testing the optimal stomatal theory with measurements from mature Pinus taeda trees in elevated [CO2]. Tree Physiology, 31(12), 1333–1344. https://doi.org/10.1093/treephys/tpr104
2. Bonan, G. B., Williams, M., Fisher, R. A., & Oleson, K. W. (2014). Modeling stomatal conductance in the earth system: linking leaf water-use efficiency and water transport along the soil–plant–atmosphere continuum. Geoscientific Model Development, 7(5), 2193–2222. https://doi.org/10.5194/gmd-7-2193-2014
3. Cowan, I. R., & Farquhar, G. D. (1977). Stomatal function in relation to leaf metabolism and environment. Symposia of the Society for Experimental Biology, 31, 471–505.
4. Farquhar, G. D., Ehleringer, J. R., & Hubick, K. T. (1989). Carbon isotope discrimination and photosynthesis. Annual Review of Plant Physiology and Plant Molecular Biology, 40(1), 503–537. https://doi.org/10.1146/annurev.pp.40.060189.002443
5. Farquhar, G. D., & Sharkey, T. D. (1982). Stomatal conductance and photosynthesis. Annual Review of Plant Physiology, 33(1), 317–345. https://doi.org/10.1146/annurev.pp.33.060182.001533
6. Franks, P. J., Drake, P. L., & Beerling, D. J. (2009). Plasticity in maximum stomatal conductance constrained by negative correlation between stomatal size and density: an analysis using Eucalyptus globulus. Plant, Cell & Environment, 32(12), 1737–1748. https://doi.org/10.1111/j.1365-3040.2009.02031.x
7. Gerlein-Safdi, C., & Ruddell, B. L. (2017). Analysis of the stomatal conductance approach to the law of the optimum plant water use. Water Resources Research, 53(3), 2616–2630. https://doi.org/10.1002/2016WR019716
8. Hetherington, A. M., & Woodward, F. I. (2003). The role of stomata in sensing and driving environmental change. Nature, 424(6951), 901–908. https://doi.org/10.1038/nature01843
9. Jarvis, P. G. (1976). The interpretation of the variations in leaf water potential and stomatal conductance found in canopies in the field. Philosophical Transactions of the Royal Society of London. B, Biological Sciences, 273(927), 593–610. https://doi.org/10.1098/rstb.1976.0035
10. Katul, G., Palmroth, S., & Oren, R. (2009). Leaf stomatal responses to vapour pressure deficit under current and CO2-enriched atmosphere explained by the economics of gas exchange. Plant, Cell & Environment, 32(8), 968–979. https://doi.org/10.1111/j.1365-3040.2009.01977.x
11. Lawson, T., & Blatt, M. R. (2014). Stomatal size, speed, and responsiveness impact on photosynthesis and water use efficiency. Plant Physiology, 164(4), 1556–1570. https://doi.org/10.1104/pp.114.237107
12. Leuning, R. (1995). A critical appraisal of a combined stomatal-photosynthesis model for C3 plants. Plant, Cell & Environment, 18(4), 339–355. https://doi.org/10.1111/j.1365-3040.1995.tb00370.x
13. Medlyn, B. E., Duursma, R. A., Eamus, D., Ellsworth, D. S., Prentice, I. C., Barton, C. V. M., Crous, K. Y., De Angelis, P., Freeman, M., & Wingate, L. (2011). Reconciling the optimal and empirical approaches to modelling stomatal conductance. Global Change Biology, 17(6), 2134–2144. https://doi.org/10.1111/j.1365-2486.2010.02375.x
14. Medlyn, B. E., Duursma, R. A., Eamus, D., Ellsworth, D. S., Prentice, I. C., Barton, C. V. M., Crous, K. Y., De Angelis, P., Freeman, M., & Wingate, L. (2011). Reconciling the optimal and empirical approaches to modelling stomatal conductance. Global Change Biology, 17(6), 2134–2144. https://doi.org/10.1111/j.1365-2486.2010.02375.x
15. Oren, R., Sperry, J. S., Katul, G. G., Pataki, D. E., Ewers, B. E., Phillips, N., & Schäfer, K. V. R. (1999). Survey and synthesis of intra- and interspecific variation in stomatal sensitivity to vapour deficit. Plant, Cell & Environment, 22(12), 1515–1526. https://doi.org/10.1046/j.1365-3040.1999.00513.x
16. Wolf, A., Anderegg, W. R. L., & Pacala, S. W. (2016). Optimal stomatal behavior with competition for water and risk of hydraulic impairment. Proceedings of the National Academy of Sciences, 113(46), E7222–E7230. https://doi.org/10.1073/pnas.1603372113