

1Title: Prevalence and intensity of avian malaria in a quail hybrid zone

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3Running Head: Parasitism in a quail hybrid zone

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28

29Authorship Contributions

30AMR performed data analyses, constructed the first draft of the manuscript, and revised the
31manuscript based on coauthors' suggestions. JMG established the study system of hybridizing
32quail, the study site and field methodology, collected blood samples, identified individuals as
33California quail, Gambel's quail, or hybrid quail, and collated the precipitation data. JBW
34conducted parasite quantification. CNK created figures. JMG and JBW were responsible for the
35initial project conceptualization, under the supervision of Rosemary Grant. All authors were
36involved in the later stages of project development and revision of the written manuscript.

37

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39Prevalence and intensity of avian malaria in a quail hybrid zone

40

41Abstract

42Hybridization is a common and important stage in species formation in plants and animals. The
43evolutionary consequences of hybridization depend not only on reproductive compatibility
44between sympatric species, but also on factors like vulnerability to each other's predators and
45parasites. We examine infection patterns of the blood parasite *Haemoproteus lophortyx*, a
46causative agent of avian malaria, at a site in the contact zone between California quail
47(*Callipepla californica*) and Gambel's quail (*C. gambelii*). We tested whether species identity,
48sex, and year predicted infection status and intensity. While we found no effect of sex on the
49status or intensity of infection, we found differences in infection status and intensity across
50species and between years. The prevalence of infection in California and hybrid quail was lower
51than in Gambel's quail. Once infected, however, California and hybrid quail had higher infection
52intensities than Gambel's quail. California and hybrid quail exhibited no significant differences
53in prevalence or intensity of infection. These findings suggest that infection by *H. lophortyx* has
54the potential to influence species barrier dynamics in this system, however, more work is
55necessary to determine the exact evolutionary consequences of this blood parasite.

56

57**Keywords:** *Haemoproteus lophortyx*, blood parasite, *Callipepla*, Galliform, species barriers

58Introduction

59 Understanding the proximate and ultimate mechanisms underlying the maintenance of
60species barriers in localities where closely related congeners overlap is a major focus of
61evolutionary biology. Recently, parasitism has been put forth as a mechanism that may work to
62promote the formation, preservation, or breakdown of species barriers (reviewed in
63Theodosopoulos et al. 2019). Although tests of this hypothesis may be conducted in the lab (e.g.,
64Goldberg et al. 2005; Hedrick et al. 2006; Brucker and Bordenstein 2013; González et al. 2014;
65Liang et al. 2018), hybrid zones provide scientists the opportunity to study the effects of factors
66influencing gene flow, such as parasitism, in a natural setting (Harrison and Larson 2014;
67Kenney and Sweigart 2016; Mořkovský et al. 2018; Theodosopoulos et al. 2019). Because
68hybridization may have major implications for both host species as well as their parasites, it is
69important to examine coevolutionary dynamics between the two across hybrid zones (Hafner et
70al. 1998; Tompkins et al. 2003; Reullier et al. 2005; Theodosopoulos et al. 2019).

71 From the viewpoint of parasites, host hybrid zones can function as population sinks,
72population sources, or as “bridges” that enable the colonization of a new host species (Whitham
731989; Floate and Whitham 1993; Strauss 1994). Parasite specificity can either limit or facilitate
74expansion of a parasite’s range into a new host species, with parasites with lower specificity
75moving more easily between host species (Bensch et al. 2000; Ricklefs and Fallon 2002;
76Ricklefs et al. 2004; Reullier et al. 2005). Nevertheless, because hybrids may have a range of
77phenotypes intermediate to that of their parental species, hybrid zones may allow parasites of one
78parental species to evolve to colonize the other parental species, even in cases where host
79specificity is relatively high (Floate and Whitham 1993).

80 From the viewpoint of hosts, parasites may alter the direction and magnitude of gene
81 exchange between two hybridizing species, and differential parasitism between hybrids and their
82 parental species may either reinforce or degrade host species barriers (e.g., Derothe et al. 2001;
83 Parris 2004; Goldberg et al. 2005; Hedrick et al. 2006; Brucker and Bordenstein 2013; González
84 et al. 2014; Guttel and Ben-Ami 2014; Maynard et al. 2016; Eastwood et al. 2017; Liang et al.
85 2018; reviewed in Theodosopoulos et al. 2019). If hybrids experience reduced fitness, due to
86 higher susceptibility or exposure to parasites than parental species, backcrossing, resulting in
87 introgression of parental genes, should be minimized, leading to reinforcement of species
88 barriers (Mouliia 1999; Grant and Grant 2008; Theodosopoulos et al. 2019). On the other hand, in
89 instances where no other factors work to maintain species barriers, these barriers may be eroded
90 if hybrids have intermediate or greater fitness than parental species, due to reduced susceptibility
91 or exposure to parasitism (Theodosopoulos et al. 2019). Past work has provided support for both
92 parasitic driven maintenance (e.g., Derothe et al. 2001; Parris 2004; Goldberg et al. 2005;
93 Brucker and Bordenstein 2013; González et al. 2014) and erosion (e.g., Hedrick et al. 2006;
94 Guttel and Ben-Ami 2014; Maynard et al. 2016; Eastwood et al. 2017; Liang et al. 2018) of
95 species barriers, across a range of animal taxa, with ca. 37% of studies examined in a 2019
96 review suggesting that hybrids are more negatively affected by parasites, and ca. 41% of studies
97 suggesting that hybrids are less negatively affected by parasites, compared to parental species
98 (Theodosopoulos et al. 2019).

99 Differences in parasite susceptibility, exposure, or fitness costs may arise between
100 hybrids and their parental species due to physiological and/or ecological factors (Le Brun et al.
101 1992; Baack and Rieseberg 2007; Nadachowska-Brzyska et al. 2012; Grossen et al. 2014; Guttel
102 and Ben-Ami 2014; Delmore et al. 2016; Wyman et al. 2016; Zhang et al. 2017; Theodosopoulos

103et al. 2019). In terms of physiology, hybrids may be better or worse equipped than parental
104species to fight off infection. On the one hand, hybrid vigor may exist with respect to immune
105function, and thus parasite resistance, due to the admixture of locally adapted alleles conferring
106resistance from both parental species, the generation of transgressive phenotypes for resistance
107(i.e., hybrids may possess phenotypes that are extreme compared to parental phenotypes),
108interactions between hybrid immune systems and other transgressive traits such as body size,
109and/or higher MHC diversity (Rieseberg et al. 1999; Baack and Rieseberg 2007; Nadachowska-
110Brzyska et al. 2012; Grossen et al. 2014; Guttel and Ben-Ami 2014; Zhang et al. 2017;
111Theodosopoulos et al. 2019; although it is important to note that increased MHC diversity does
112not always result in heightened immune response; Sommer 2005; Sommer et al. 2014). On the
113other hand, hybrids may have decreased immune responses, or it may be more costly for hybrids
114to mount an immune response, compared to parental species, due to a higher stress response,
115fewer resources, and/or because genetic mixing may lead to debilitated immune function and/or
116metabolic processes (Dupont and Crivelli 1988; Moulia 1999; Theodosopoulos et al. 2019).
117Hybrids may also have immune functions intermediate to that of their parental species if they
118possess intermediate MHC diversity or inherit a combination of alleles related to immunity
119(Theodosopoulos et al. 2019). In addition to differences in host-parasite interactions arising from
120physiological factors, hybrids may have an ecology that is distinct from their parental types, such
121as different intra- and interspecific social relationships, food resources, and habitat use, which
122lead to higher or lower parasite transmission or intensity of infection in hybrids, compared with
123parental species (Le Brun et al. 1992; Guttel and Ben-Ami 2014; Delmore et al. 2016; Wyman et
124al. 2016; Theodosopoulos et al. 2019).

125 Using four years of data, we explored infection patterns of *Haemoproteus lophortyx*, an
126 intraerythrocytic parasite, at a site in the northern region of the contact zone between California
127 quail, *Callipepla californica*; Shaw, 1798 and Gambel's quail, *Callipepla gambelii*; Gambel,
128 1843. The hybrid zone is located along an ecological transition between the relatively the xeric
129 habitat of Gambel's quail to the relatively mesic habitat of California quail, and local gene
130 exchange occurs frequently in disjunct patches of species overlap. (Gee 2003, 2004). We
131 compared the status and intensity of *H. lophortyx* infection in California quail, Gambel's quail,
132 and their hybrids. We also considered whether year and sex might influence infection status and
133 intensity.

134

135 **Methods**

136 *Study System and field methods*

137 California quail and Gambel's quail are sister species that are medium-sized (ca. 150-
138 200g), highly social, sexually dichromatic, nonmigratory New World quail (Odontophoridae;
139 Leopold 1977; Zink and Blackwell 1998; Hosner et al. 2015; Gee 2003; 2004). California quail
140 are native to the western United States and Baja California, preferring chaparral and semiarid
141 scrub, while the natural ranges of Gambel's quail span the Mojave and Sonoran Deserts, with
142 these quail preferring more arid environments (Leopold 1977). These sister taxa hybridize
143 readily under captive and natural conditions, and there is no evidence for assortative mating in
144 the hybrid zone (Johnsgard 1971, Gee 2003; 2004; 2005). The hybrid zone straddles a narrow
145 ecotone (roughly 20-30 km), and even across that distance, habitat differences between the
146 species are striking. Hybrids of all classes are present in the quail hybrid zone and hybrids can

147easily comprise at least 20% of the population, depending upon the ecological conditions (Gee
1482004).

149 The genus *Haemoproteus* is a diverse group which parasitizes a range of avian species
150(e.g., Ricklefs et al. 2005; Valkiūnas et al. 2007; 2010; 2013; Iezhova et al. 2011; Levin et al.
1512012; Swanson et al. 2014; Ayadi et al. 2018). Along with parasites of the genus *Plasmodium*,
152*Haemoproteus* can cause avian malaria, however, unlike *Plasmodium*, *Haemoproteus* lineages
153appear to have high host specificity (Atkinson and Van Riper 1991a; Bensch 2000; Clark and
154Clegg 2017; Loiseau et al. 2017; Ayadi et al. 2018). *Haemoproteus lophortyx*; O'Roke, 1929
155may cause anemia, prostration, and death in various quail species, including California quail,
156Gambel's quail, and bobwhite quail, *Colinus virginianus*; Linnaeus, 1758 (O'Roke 1928; 1930;
1571932; Herman et al. 1942; Tarshis 1955; 1958; Gullion 1957; Cardona et al. 2002; Mullens et al.
1582006; Samour 2016). There exists no evidence for sex differences in infection in California quail
159(Herman and Glading 1942). Past work has demonstrated that, in quail, *H. lophortyx* may be
160spread by several vectors including hippoboscids flies (*Lynchia hirsuta*; Ferris 1927 and
161*Stilbometopa impressa*; Bigot, 1885; both of which are obligate ectoparasites) and biting midges
162(*Culicoides* spp. – especially *C. bottimeri*; Wirth, 1955 (Tarshis 1955; 1958; Mullens et al. 2006;
163Samour 2016).

164 JMG and student assistants trapped adult quail using seed-baited, walk-in funnel traps
165from January-September in 1998-2001 at a site called Royal Carrizo (33.6410° N, 116.4253° W),
166which consists of pinyon-juniper woodland habitat at ca. 3000' in Southern California. Hybrid
167reproductive success at this sympatric study site is moderate, based on comparisons of clutch
168size and hatching success with the two parental species in allopatry (Gee 2003). We assigned
169birds as California quail, Gambel's quail, or hybrids, based on morphological features which are

170tightly correlated with genotype (Figure 1; Gee 2004). At the time of first capture, we ringed
171birds for individual identification and collected blood samples to test for *H. lophortyx* infection.
172We collected blood from the left brachial vein into a microcapillary tube. We smeared the blood
173onto a glass slide, which we air-dried and fixed in 95% ethanol before Giemsa-staining. This
174work was conducted under California State Fish and Game permit SC 949 and was approved by
175Princeton University's Institutional Animal Care and Use Committee.

176

177*Parasite quantification*

178 Blood smears consisted of ca. 30,000 erythrocytes per slide. Criteria for *H. lophortyx*
179identification were based on the morphological descriptions given by O'Roke (1928) and
180Atkinson and Van Riper (1991b). We examined a minimum of 30 fields of view at 100x
181magnification, using an oil immersion lens. At this magnification, a mean \pm SD of 184 ± 73
182erythrocytes per field of view per sample was examined. We obscured all information about the
183sample before parasite quantification. We recorded the number of infected and uninfected
184erythrocytes in each sample.

185 We collected 208 blood samples across 4 years (1998: N = 61; 1999: N = 32; 2000: N =
186102; 2001: N = 13; see Table S1). *H. lophortyx* was present in 69 of these blood samples.

187

188*Statistical methods*

189 We used R 3.5.2 (R Core Team 2018) for all analyses and examined the factors affecting
190the status (i.e., 1= one or more *H. lophortyx* were found in the sample, 0 = no *H. lophortyx* were
191found in the sample) and intensity (i.e., proportion of infected erythrocytes) of *H. lophortyx*
192infection. To examine infection status, we used a generalized linear mixed model with a binomial

family and the status of infection as the response. To examine infection intensity, we ran a similar model, but instead, included the ratio of infected to uninfected erythrocytes as the response. For this analysis, we only examined the subset of individuals that were infected. For both sets of analyses we included species (i.e., California quail, Gambel's quail, or hybrids), sex, and year as fixed effects, and individual identity and month as random effects. We included month as a random effect given that past work has demonstrated seasonal fluctuations in *H. lophortyx* infection in quail (Tarshis 1955; Cardona et al. 2002). Given that species was a categorical variable, we examined whether the overall effect of species was significant by comparing models with and without species, using likelihood ratio tests. Similarly, given that year and sex were also categorical, we used likelihood ratio tests to compare a model with and without year and to compare a model with and without sex to determine the overall effect of these variables.

205

206 Results

We tested for the presence of *H. lophortyx* in 193 quail (72 California quail, 27 Gambel's quail, and 94 hybrids), 13 of which were sampled twice across years (3 California quail, 3 Gambel's quail, and 7 hybrids), and one California quail which was sampled thrice, for a total of 208 blood smears. Of the 208 blood smears examined, 69 (~33%) showed signs of *H. lophortyx* infection (24 California quail smears (~31%), 16 Gambel's quail smears (~53%), and 29 hybrid smears (~29%). Of the 69 blood smears in which infection was observed, infection intensity ranged from $4.700\text{e-}5$ – 0.013 *H. lophortyx*/cell with a mean \pm SD of 0.002 ± 0.002 *H. lophortyx*/cell (California quail: range = $5.710\text{e-}5$ – 0.013 *H. lophortyx*/cell, mean \pm SD = 0.003 ± 0.003 *H. lophortyx*/cell; Gambel's quail: range = $6.000\text{e-}5$ – 0.001 *H. lophortyx*/cell, mean \pm SD = 3.686e-

2164 \pm 2.689 e-4 *H. lophortyx*/cell; hybrids: range = 4.700e-5 – 0.009 *H. lophortyx*/cell, mean \pm SD
217= 0.001 \pm 0.002 *H. lophortyx*/cell).

218 For the analysis examining infection status, we found that significantly more Gambel's
219quail were infected than either California or hybrid quail, but we found no significant difference
220between hybrids and California quail (i.e., Gambel's quail > hybrid quail = California quail;
221Table 1; Figure 2a). In contrast, for the analysis examining the intensity of infection in the subset
222of individuals that were infected, California and hybrid quail had significantly higher proportions
223of *H. lophortyx*/cell than Gambel's quail (Table 2; Figure 3a). Again, there was no significant
224difference between hybrids and California quail (i.e., Gambel's quail < hybrid quail = California
225quail, Table 2; Figure 3a). There was a significant overall effect of species on *H. lophortyx*
226infection status (Chi-squared = 8.997, df = 2, p = 0.011) and intensity (Chi-squared = 8.162, df =
2272, p = 0.017). Furthermore, there was a significant overall effect of year on infection status (Chi-
228squared = 21.560, df = 3, p < 0.001), with significantly less individuals infected in 1999 than in
2291998 or 2000 (Table S2; Figure 2b). Although we also found a significant overall effect of year
230on infection intensity (Chi-squared = 17.367, df = 3, p = 0.001), with a significantly higher
231intensity of infection in 1999 compared to 2000 (Table S3; Figure 3b), it is important to note that
232we had a limited sample size for both 1999 and 2001, as we only used the subset of data with
233infected individuals for this analysis. Given this, these results should be taken with caution. We
234found no significant overall effect of sex on infection status (Chi-squared = 0.225, df = 1, p =
2350.635) or intensity (Chi-squared = 0.491, df = 1, p = 0.484).

236 In order to check the robustness of our results, we reran analyses using a subset of data
237where at least 10,000 erythrocytes were examined for each sample (as opposed to examining 30
238fields of view, which sometimes resulted the examination of <10,000 erythrocytes; Rätti et al.

2391993; Staats and Schall 1996; Kelly et al. 2016; Shurulinkov et al. 2018). This reduced our
240overall sample size from 208 to 189 blood smears and our infected sample size from 69 to 63
241blood smears. Our results for these follow-up analyses were qualitatively similar to our original
242results (Appendix 1; Tables S6 and S7), suggesting that our findings were robust to different
243sampling methods.

244

245Discussion

246 The hybridization of two host species can have complex effects on host-parasite
247interactions. We found that while there was no significant difference between hybrid California ×
248Gambel's quail and California quail in *Haemoproteus lophortyx* infection status and intensity,
249both species had lower infection prevalence but higher infection intensity than Gambel's quail.
250Our results suggest that, in this Galliform host system, infection by the blood parasite *H.*
251*lophortyx* has the potential to impact species barrier dynamics.

252 We found that there were fewer *H. lophortyx* infected California and hybrid quail than
253Gambel's quail, but upon infection, California and hybrid quail had higher infection intensities
254compared to Gambel's quail. This may appear counterintuitive, as one might expect relationships
255involving infection status and intensity to be similar in directionality. For example, a study
256conducted on three species of wild doves found that species which had higher likelihoods of
257*Haemoproteus columbae* infection had higher, rather than lower, infection intensities (Adriano &
258Cordeiro 2001). There are two possible scenarios that may lead to the patterns observed in our
259study (see Table S4 for a complete summary of these scenarios and their potential implications).

260 First, it is possible that host behavior or vector preference causes Gambel's quail to
261experience higher rates of exposure to *H. lophortyx*, compared to California quail, leading to

262higher infection prevalence in the former. For example, the frequency, type, network structure, or
263duration of social interactions in Gambel's quail may be more amenable to the direct horizontal
264transmission of *H. lophortyx* carrying hippoboscids flies, compared to the social interactions of
265California quail. Furthermore, biting midges have been implicated in the transmission of *H.*
266*lophortyx* in bobwhite quail and may also transmit *H. lophortyx* in the quail species examined in
267our study (Mullens et al. 2006). Gambel's quail could be more likely than California quail to use
268habitats where biting midges are more prevalent, leading Gambel's quail to experience greater
269exposure to *H. lophortyx*. If Gambel's quail experience higher rates of *H. lophortyx*, compared to
270California quail, due to their ecology, this may indicate a longer coevolutionary history between
271Gambel's quail and *H. lophortyx* than between California quail and *H. lophortyx*, leading
272Gambel's quail to have evolved higher resistance to *H. lophortyx* infection (i.e., they are better
273able to fight off infection), explaining their relatively low infection intensities. From the
274parasite's perspective, if Gambel's quail have a longer coevolutionary history with *H. lophortyx*
275than California quail, it is possible that the hybrid zone may have acted as a bridge for *H.*
276*lophortyx* to expand its range from Gambel's quail to California quail.

277 Hybrid quail may have inherited behavioral traits (e.g., habitat use and/or social
278behavior) more similar to parental California quail, making them less likely to acquire the
279parasite than Gambel's quail. Furthermore, hybrid quail may not have inherited resistance alleles
280from parental Gambel's quail. If Gambel's quail have lower infection intensities than California
281and hybrid quail, due to higher resistance to infection, and if hybrid quail have reduced fitness
282compared to Gambel's quail, as a result of higher infection intensities, we would expect species
283barriers to be maintained. Parasitism has been demonstrated to aid in the maintenance of species

284barriers in a number of species (e.g., Derothe et al. 2001; Parris 2004; Goldberg et al. 2005;
285Brucker and Bordenstein 2013; González et al. 2014; reviewed in Theodosopoulos et al. 2019).

286 Second, Gambel's quail may actually have *lower* resistance to *H. lophortyx* than
287California quail, causing them to exhibit a higher prevalence, but lower intensity of infection,
288compared to California quail. In this scenario, Gambel's quail may experience similar rates of
289exposure to *H. lophortyx* as California quail. However, Gambel's quail may be more likely to
290become infected and less likely to completely clear their system of the parasite, once infected,
291compared to California quail, leading to a relatively high prevalence of low-intensity, chronic
292infections. Because infection intensities are usually highest during the early stages of infection
293(e.g., Ahmed & Mohammed 1978; Cepeda et al. 2019), the few individuals with relatively high
294infection intensities seen in California quail may simply reflect a handful of newly infected
295individuals.

296 In this case, hybrid quail may have inherited resistance alleles from parental California
297quail. Chronic *Haemoproteus* infections have been demonstrated to have fitness costs in other
298host systems (Puente et al. 2010; Asghar et al. 2011; 2015; but see Ortego et al. 2008). If chronic
299infections in Gambel's quail lead individuals to have lower fitness compared to hybrid quail,
300hybridization and backcrossing may facilitate the introgression of resistance alleles, and we
301would expect species barriers to be eroded. This aligns with numerous studies that have
302suggested the occurrence of parasite-mediated breakdowns of species barriers (e.g., Hedrick et
303al. 2006; Guttel and Ben-Ami 2014; Maynard et al. 2016; Eastwood et al. 2017; Liang et al.
3042018; reviewed in Theodosopoulos et al. 2019).

305 While our results were robust across various sampling methods, more work is needed to
306tease apart the two mutually exclusive interpretations of our results presented above. Although

past work has shown that *H. lophortyx* can have high fitness costs for quail, even resulting in death (O’Roke 1930; 1932; Cardona et al. 2002; Mullens et al. 2006), we did not examine the relative fitness costs of this parasite for each host species or the potential costs of chronic, low-intensity infections. Given this, while we can infer the potential for *H. lophortyx* to influence species barrier dynamics, we cannot draw any definitive conclusions regarding the parasite-driven maintenance or breakdown of species barriers, and future work should assess the relative fitness costs and course of infection of *H. lophortyx* across this hybrid zone. Furthermore, our study possesses several other limitations that future studies may work to address. For example, we scored individuals as either hybrids or parental species (Figure 1), however, because hybrids are not truly a single class, differences may exist between hybrids in parasite susceptibility or fitness costs, thereby affecting the influence of parasitism on gene flow (Derothe et al. 2004; Goldberg et al. 2005; Theodosopoulos et al. 2019). For example, if most of the hybrids sampled in our study were highly backcrossed to California quail, this could explain the observed similarities between hybrid and California quail in prevalence and intensity of *H. lophortyx* infection. Furthermore, because interactions between different parasites may not be additive, coinfection dynamics can have drastic consequences for the effects of parasitism on gene flow, and studies which examine a single parasite may only capture a portion of the story (Telfer et al. 2010; Bordes and Morand 2011; Johnson and Hoverman 2012; Rynkiewicz et al. 2015; Vaumourin et al. 2015; Theodosopoulos et al. 2019). Lastly, we assumed California quail, Gambel’s quail, and their hybrids were all infected by a single lineage of *Haemoproteus* in this study. However, a drawback of blood smears is that different lineages may appear morphologically identical, while remaining genetically distinct (Bensch et al. 2000). This is especially relevant given the high host specificity of *Haemoproteus* lineages, and it is possible

330that different lineages may have disparate pathologies (Atkinson and Van Riper 1991a; Bensch
3312000; Clark and Clegg 2017; Loiseau et al. 2017; Ayadi et al. 2018).

332 Lastly, we found that patterns of infection varied across years; however, the cause(s) of
333these differences remains unclear. Increases in precipitation have been found to either increase
334host exposure rates (O'Connor et al., 2007, 2008; Bohrer et al., 2014; Shearer and Ezenwa 2020)
335or decrease host susceptibility to parasitism (Ezenwa 2004; Thurber et al. 2011; Masi et al. 2012;
336Shearer and Ezenwa 2020) in other systems. Although El Niño Southern Oscillations caused
337precipitation to fluctuate across years, in our study, it is unlikely that the differences in infection
338status and intensity seen between years were related to variation in precipitation. (See Table S5
339for a summary of how year relates to precipitation, infection status, and infection intensity.)
340Nevertheless, because hybridization has been shown to increase with increased precipitation
341(Gee 2004), precipitation may work in tandem with *H. lophortyx* to mediate species barrier
342dynamics.

343 In summary, we found that infection prevalence was higher, while infection intensity was
344lower, in Gambel's quail, compared to hybrid and California quail, suggesting that *H. lophortyx*
345infection has the potential to influence species barrier dynamics in this system. Future work
346should focus on examining the fitness consequences and course of infection of *H. lophortyx*, as
347well as the diversity of *Haemoproteus* spp. and their distribution across this quail hybrid zone.
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349**Data Accessibility Statement**

350The data associated with this manuscript are available on Figshare
351(<https://doi.org/10.6084/m9.figshare.12217958>)

352Literature Cited

353

354Adriano, E. A., & Cordeiro, N. S. (2001). Prevalence and intensity of *Haemoproteus columbae*
355in three species of wild doves from Brazil. *Memorias do Instituto Oswaldo Cruz*, 96, 175-178.

356

357Ahmed, F. E., & Mohammed, A. H. H. (1978). *Haemoproteus columbae*: course of infection,
358relapse and immunity to reinfection in the pigeon. *Zeitschrift für Parasitenkunde*, 57(3), 229-236.

359

360Asghar, M., Hasselquist, D., & Bensch, S. (2011). Are chronic avian haemosporidian infections
361costly in wild birds?. *J. Avian Biol.*, 42(6), 530-537.

362

363Asghar, M., Hasselquist, D., Hansson, B., Zehtindjiev, P., Westerdahl, H., & Bensch, S. (2015).
364Hidden costs of infection: chronic malaria accelerates telomere degradation and senescence in
365wild birds. *Science*, 347(6220), 436-438.

366

367Atkinson, C. T., and Van Riper III, C. (1991a). Vectors, epizootiology, and pathogenicity of
368avian species of *Haemoproteus* (Haemosporina: Haemoproteidae). *Bulletin of the Society for*
369*Vector Ecology*, 16, 109-126.

370

371Atkinson, C. T. and Van Riper III, C. (1991b). Pathogenicity and epizootiology of avian
372haematozoa: *Plasmodium*, *Haemoproteus*, and *Leucocytozoon*. In *Bird-parasite interactions* (Ed.
373J. E. Loye and M. Zuk). pp. 19-48. Oxford University Press.

374

375Ayadi, T., Selmi, S., Hammouda, A., Reis, S., Boulinier, T., and Loiseau, C. (2018). Diversity,
376prevalence and host specificity of avian parasites in southern Tunisian oases. *Parasitology*,
377145(7), 971-978.

378

379Baack, E. J., and Rieseberg, L. H. (2007). A genomic view of introgression and hybrid
380speciation. *Curr. Opin. Genet. Dev.*, 17(6), 513-518.

381

382Bensch, S., Stjernman, M., Hasselquist, D., Örjan, Ö., Hansson, B., Westerdahl, H., and
383Pinheiro, R. T. (2000). Host specificity in avian blood parasites: a study of *Plasmodium* and
384Haemoproteus mitochondrial DNA amplified from birds. *Proc. R. Soc. Lond., B, Biol. Sci.*,
385267(1452), 1583-1589.

386

387Bohrer, G., Beck, P. S., Ngene, S. M., Skidmore, A. K., and Douglas-Hamilton, I. (2014).
388Elephant movement closely tracks precipitation-driven vegetation dynamics in a Kenyan forest-
389savanna landscape. *Mov. Ecol.*, 2(1), 2.

390

391Bordes, F., and Morand, S. (2011). The impact of multiple infections on wild animal hosts: a
392review. *Infect. Ecol. Epidemiol.*, 1(1), 7346.

393

394Brucker, R. M., and Bordenstein, S. R. (2013). The hologenomic basis of speciation: gut bacteria
395cause hybrid lethality in the genus *Nasonia*. *Science*, 341(6146), 667-669.

396

397Cardona, C. J., Ihejirika, A., and McClellan, L. (2002). *Haemoproteus lophortyx* infection in
398bobwhite quail. *Avian Dis.*, 46(1), 249-255.

399

400Cepeda, A. S., Lotta-Arévalo, I. A., Pinto-Osorio, D. F., Macías-Zacipa, J., Valkiūnas, G.,
401Barato, P., & Matta, N. E. (2019). Experimental characterization of the complete life cycle of
402*Haemoproteus columbae*, with a description of a natural host-parasite system used to study this
403infection. *Int. J. Parasitol.*, 49(12), 975-984.

404

405Clark, N. J., and Clegg, S. M. (2017). Integrating phylogenetic and ecological distances reveals
406new insights into parasite host specificity. *Mol. Ecol.*, 26(11), 3074-3086.

407

408Delmore, K. E., Toews, D. P., Germain, R. R., Owens, G. L., and Irwin, D. E. (2016). The
409genetics of seasonal migration and plumage color. *Curr. Biol.*, 26(16), 2167-2173.

410

411Derothe, J. M., Le Brun, N., Loubes, C., Perriat-Sanguinet, M., and Moulia, C. (2001).
412Susceptibility of natural hybrids between house mouse subspecies to *Sarcocystis muris*. *Int. J.*
413*Parasitol.*, 31(1), 15-19.

414

415Derothe, J. M., Porcherie, A., Perriat-Sanguinet, M., Loubès, C., and Moulia, C. (2004).
416Recombination does not generate pinworm susceptibility during experimental crosses between
417two mouse subspecies. *Parasitol. Res.*, 93(5), 356-363.

418

419Dupont, F., and Crivelli, A. J. (1988). Do parasites confer a disadvantage to hybrids?. *Oecologia*,
42075(4), 587-592.

421

422Eastwood, J. R., Ribot, R. F., Rollins, L. A., Buchanan, K. L., Walder, K., Bennett, A. T., and
423Berg, M. L. (2017). Host heterozygosity and genotype rarity affect viral dynamics in an avian
424subspecies complex. *Sci. Rep.*, 7(1), 13310.

425

426Ezenwa, V. O. (2004). Interactions among host diet, nutritional status and gastrointestinal
427parasite infection in wild bovids. *Int. J. Parasitol.*, 34(4), 535-542.

428

429Floate, K. D., and Whitham, T. G. (1993). The "hybrid bridge" hypothesis: host shifting via plant
430hybrid swarms. *Am. Nat.*, 141(4), 651-662.

431

432Gee, J. M. (2003). How a hybrid zone is maintained: behavioral mechanisms of interbreeding
433between California and Gambel's quail (*Callipepla californica* and *C. gambelii*). *Evolution*,
43457(10), 2407-2415.

435

436Gee, J. M. (2004). Gene flow across a climatic barrier between hybridizing avian species,
437California and Gambel's quail (*Callipepla californica* and *C. gambelii*). *Evolution*, 58(5), 1108-
4381121.

439

440Gee, J. M. (2005). No species barrier by call in an avian hybrid zone between California and
441Gambel's quail (*Callipepla californica* and *C. gambelii*). *Biol. J. Linnean Soc.*, 86(2), 253-264.

442

443Goldberg, T. L., Grant, E. C., Inendino, K. R., Kassler, T. W., Claussen, J. E., and Philipp, D. P.

444(2005). Increased infectious disease susceptibility resulting from outbreeding depression.

445Conserv. Biol., 19(2), 455-462.

446

447González, R., Lohrmann, K. B., Pizarro, J., and Brokordt, K. (2014). Differential susceptibility to

448the Withering Syndrome agent and renal coccidia in juvenile *Haliotis rufescens*, *Haliotis discus*

449*hannai* and the interspecific hybrid. J. Invertebr. Pathol., 116, 13-17.

450

451Grant, P., and Grant, B.R. (2008). How and why species multiply: The Radiation of Darwin's

452Finches. Princeton University Press. Princeton, New Jersey, USA.

453

454Grossen, C., Keller, L., Biebach, I., Croll, D., and International Goat Genome Consortium.

455(2014). Introgression from domestic goat generated variation at the major histocompatibility

456complex of alpine ibex. PLoS Genet., 10(6), e1004438.

457

458Gullion, G. W. (1957). Gambel quail disease and parasite investigations in Nevada. Am. Midl.

459Nat., 414-420.

460

461Guttel, Y., and Ben-Ami, F. (2014). The maintenance of hybrids by parasitism in a freshwater

462snail. Int. J. Parasitol., 44(13), 1001-1008.

463

464Hafner, M. S., Demastes, J. W., Hafner, D. J., Spradling, T. A., Sudman, P. D., and Nadler, S. A.
465(1998). Age and movement of a hybrid zone: implications for dispersal distance in pocket
466gophers and their chewing lice. *Evolution*, 52(1), 278-282.

467

468Harrison, R. G., and Larson, E. L. (2014). Hybridization, introgression, and the nature of species
469boundaries. *J. Hered.*, 105(S2), 795-809.

470

471Hedrick, R. P., Waltzek, T. B., and McDowell, T. S. (2006). Susceptibility of koi carp, common
472carp, goldfish, and goldfish× common carp hybrids to cyprinid herpesvirus-2 and herpesvirus-3.
473*J. Aquat. Anim. Health*, 18(1), 26-34.

474

475Herman, C. M., and Glading, B. (1942). The protozoan blood parasite *Haemoproteus lophortyx*
476O'Roke in the quail at the San Joaquin experimental range, California. *Calif. Fish Game*, 28,
477150-153.

478

479Hosner, P. A., Braun, E. L., and Kimball, R. T. (2015). Land connectivity changes and global
480cooling shaped the colonization history and diversification of New World quail (Aves:
481Galliformes: Odontophoridae). *J. Biogeogr.*, 42(10), 1883-1895.

482

483Iezhova, T. A., Dodge, M., Sehgal, R. N., Smith, T. B., and Valkiūnas, G. (2011). New avian
484*Haemoproteus* species (Haemosporida: Haemoproteidae) from African birds, with a critique of
485the use of host taxonomic information in hemoproteid classification. *J. Parasitol.*, 97(4), 682-694.

486

487Johnsgard, P. A. (1971). Experimental hybridization of the New World quail (Odontophorinae).
488Auk, 88(2), 264-275.

489

490Johnson, P. T., and Hoverman, J. T. (2012). Parasite diversity and coinfection determine
491pathogen infection success and host fitness. Proc. Natl. Acad. Sci. U.S.A., 109(23), 9006-9011.

492

493Kelly, T. R., MacGillivray, H. L., Sarquis-Adamson, Y., Watson, M. J., Hobson, K. A., and
494MacDougall-Shackleton, E. A. (2016). Seasonal migration distance varies with natal dispersal
495and predicts parasitic infection in song sparrows. Behav. Ecol. Sociobiol., 70(11), 1857-1866.

496

497Kenney, A. M., and Sweigart, A. L. (2016). Reproductive isolation and introgression between
498sympatric *Mimulus* species. Mol. Ecol., 25(11), 2499-2517.

499

500Le Brun, N. L., Renaud, F., Berrebi, P., and Lambert, A. (1992). Hybrid zones and host-parasite
501relationships: effect on the evolution of parasitic specificity. Evolution, 46(1), 56-61.

502

503Leopold, A. S. 1977. The California quail. University of California Press, Berkeley.

504

505Levin, I. I., Valkiūnas, G., Iezhova, T. A., O'Brien, S. L., and Parker, P. G. (2012). Novel
506Haemoproteus species (Haemosporida: Haemoproteidae) from the swallow-tailed gull (Lariidae),
507with remarks on the host range of hippoboscoid-transmitted avian hemoproteids. J. Parasitol.,
50898(4), 847-854.

509

510Liang, S., Luo, X., You, W., and Ke, C. (2018). Hybridization improved bacteria resistance in
511abalone: evidence from physiological and molecular responses. *Fish Shellfish Immunol.*, 72,
512679-689.

513

514Loiseau, C., Melo, M., Lobato, E., Beadell, J. S., Fleischer, R. C., Reis, S., ... and Covas, R.
515(2017). Insularity effects on the assemblage of the blood parasite community of the birds from
516the Gulf of Guinea. *J. Biogeogr.*, 44(11), 2607-2617.

517

518Masi, S., Chauffour, S., Bain, O., Todd, A., Guillot, J., and Krief, S. (2012). Seasonal effects on
519great ape health: a case study of wild chimpanzees and western gorillas. *PLoS One*, 7(12).

520

521Maynard, B. T., Taylor, R. S., Kube, P. D., Cook, M. T., and Elliott, N. G. (2016). Salmonid
522heterosis for resistance to amoebic gill disease (AGD). *Aquaculture*, 451, 106-112.

523

524Mořkovský, L., Janoušek, V., Reif, J., Řídl, J., Pačes, J., Choleva, L., Janko, J., Nachman, M.
525W., and Reifová, R. (2018). Genomic islands of differentiation in two songbird species reveal
526candidate genes for hybrid female sterility. *Mol. Ecol.*, 27(4), 949-958.

527

528Mouliá, C. (1999). Parasitism of plant and animal hybrids: are facts and fates the same?.

529*Ecology*, 80(2), 392-406.

530

531Mullens, B. A., Cardona, C. J., McClellan, L., Szijj, C. E., and Owen, J. P. (2006). *Culicoides*
532**bottimeri** as a vector of *Haemoproteus lophortyx* to quail in California, USA. Vet. Parasitol.,
533140(1-2), 35-43.

534

535Nadachowska-Brzyska, K., Zieliński, P., Radwan, J., and Babik, W. (2012). Interspecific
536hybridization increases MHC class II diversity in two sister species of newts. Mol. Ecol., 21(4),
537887-906.

538

539Ortego, J., Cordero, P. J., Aparicio, J. M., & Calabuig, G. (2008). Consequences of chronic
540infections with three different avian malaria lineages on reproductive performance of Lesser
541Kestrels (*Falco naumanni*). J. Ornithol., 149(3), 337-343.

542

543O'Roke, E. C. (1928). Parasites and parasitic diseases in the California valley quail. Calif. Fish
544Game, 14, 193-198.

545

546O'Roke, E. C. (1930). The morphology, transmission and life-history of *Haemoproteus*
547**lophortyx** O'Roke, a blood parasite of the California valley quail. Univ. Calif. Publ. Zool. 36:1–
54850.

549

550O'Roke, E. C. (1932). Parasitism of the California valley quail by *Haemoproteus lophortyx*, a
551protozoan blood parasite. Calif. Fish Game 18:223–238.

552

553Parris, M. J. (2004). Hybrid response to pathogen infection in interspecific crosses between two
554amphibian species (Anura: Ranidae). *Evol. Ecol. Res.*, 6(3), 457-471.

555

556Puente, J. M. D. L., Merino, S., Tomás, G., Moreno, J., Morales, J., Lobato, E., ... & Belda, E. J.
557(2010). The blood parasite *Haemoproteus* reduces survival in a wild bird: a medication
558experiment. *Biol. Lett.*, 6(5), 663-665.

559

560R Core Team (2018). R: A language and environment for statistical computing. R Foundation for
561Statistical Computing, Vienna, Austria. URL <https://www.R-project.org/>.

562

563Rätti, O., Dufva, R., and Alatalo, R. V. (1993). Blood parasites and male fitness in the pied
564flycatcher. *Oecologia*, 96(3), 410-414.

565

566Reullier, J., J. Perez-Tris, S. Bensch, and J. Secondi. (2005). Diversity, distribution and exchange
567of blood parasites meeting at an avian moving contact zone. *Mol. Ecol.* 0:1-11.

568

569Ricklefs, R. E., and Fallon, S. M. (2002). Diversification and host switching in avian malaria
570parasites. *Proc. R. Soc. Lond., B, Biol. Sci.*, 269(1494), 885-892.

571

572Ricklefs, R. E., Fallon, S. M., and Bermingham, E. (2004). Evolutionary relationships,
573cospeciation, and host switching in avian malaria parasites. *Syst. Biol.*, 53(1), 111-119.

574

575Ricklefs, R. E., Swanson, B. L., Fallon, S. M., Martínez-Abraín, A., Scheuerlein, A., Gray, J.,
576and Latta, S. C. (2005). Community relationships of avian malaria parasites in southern
577Missouri. *Ecol. Monogr.*, 75(4), 543-559.

578

579Rieseberg, L. H., Archer, M. A., and Wayne, R. K. (1999). Transgressive segregation, adaptation
580and speciation. *Heredity*, 83(4), 363-372.

581

582Rynkiewicz, E. C., Pedersen, A. B., and Fenton, A. (2015). An ecosystem approach to
583understanding and managing within-host parasite community dynamics. *Trends Parasitol.*, 31(5),
584212-221.

585

586Samour, J. (2016). *Avian medicine* (No. Ed. 3). Mosby International Ltd.

587

588Shearer, C. L., & Ezenwa, V. O. (2020). Rainfall as a driver of seasonality in parasitism. *Int. J.*
589*Parasitol. Parasites Wildl.*

590

591Shurulinkov, P., Spasov, L., Stoyanov, G., and Chakarov, N. (2018). Blood parasite infections in
592a wild population of ravens (*Corvus corax*) in Bulgaria. *Malar. J.*, 17(1), 33.

593

594Sommer, S. (2005). The importance of immune gene variability (MHC) in evolutionary ecology
595and conservation. *Front. Zool.*, 2(1), 16.

596

597Sommer, S., Rakotondranary, S. J., and Ganzhorn, J. U. (2014). Maintaining microendemic
598primate species along an environmental gradient—parasites as drivers for species differentiation.
599Ecol. Evol., 4(24), 4751-4765.

600

601Staats, C. M., and Schall, J. J. (1996). Distribution and abundance of two malarial parasites of
602the endemic *Anolis* lizard of Saba island, Netherlands Antilles. J. Parasitol., 409-413.

603

604Strauss, S. Y. (1994). Levels of herbivory and parasitism in host hybrid zones. Trends Ecol.
605Evol., 9(6), 209-214.

606

607Swanson, B. L., Lyons, A. C., and Bouzat, J. L. (2014). Distribution, prevalence and host
608specificity of avian malaria parasites across the breeding range of the migratory lark sparrow
609(*Chondestes grammacus*). Genetica, 142(3), 235-249.

610

611Tarshis, I. B. (1955). Transmission of *Haemoproteus lophortyx* O'Roke of the California quail by
612hippoboscids of the species *Stilbometopa impressa* (Bigot) and *Lynchia hirsuta* Ferris. Exp.
613Parasitol., 4(5), 464-492.

614

615Tarshis, I. B. (1958). New data on the biology of *Stilbometopa impressa* (Bigot) and *Lynchia*
616*hirsuta* Ferris (Diptera: Hippoboscidae). Ann. Entomol. Soc. Am., 51(1), 95-105.

617

618Telfer, S., Lambin, X., Birtles, R., Beldomenico, P., Burthe, S., Paterson, S., and Begon, M.
619(2010). Species interactions in a parasite community drive infection risk in a wildlife population.
620Science, 330(6001), 243-246.

621

622Theodosopoulos, A. N., Hund, A. K., and Taylor, S. A. (2019). Parasites and host species
623barriers in animal hybrid zones. Trends Ecol. Evol., 34(1), 19-30.

624

625Thurber, M. I., O'Connell-Rodwell, C. E., Turner, W. C., Nambandi, K., Kinzley, C., Rodwell,
626T. C., ... and Bouley, D. M. (2011). Effects of rainfall, host demography, and musth on strongyle
627fecal egg counts in African elephants (*Loxodonta africana*) in Namibia. J. Wildl. Dis., 47(1),
628172-181.

629

630Tompkins, D. M., White, A. R., and Boots, M. (2003). Ecological replacement of native red
631squirrels by invasive greys driven by disease. Ecol. Lett., 6(3), 189-196.

632

633Vaumourin, E., Vourc'h, G., Gasqui, P., and Vayssier-Taussat, M. (2015). The importance of
634multiparasitism: examining the consequences of co-infections for human and animal health.
635Parasit. Vectors, 8(1), 545.

636

637Valkiūnas, G., Križanauskienė, A., Iezhova, T. A., Hellgren, O., and Bensch, S. (2007).
638Molecular phylogenetic analysis of circumnuclear hemoproteids (Haemosporida:
639Haemoproteidae) of sylviid birds, with a description of *Haemoproteus parabelopolskyi* sp. nov.
640J. Parasitol., 93(3), 680-687.

641

642Valkiūnas, G., Santiago-Alarcon, D., Levin, I. I., Iezhova, T. A., and Parker, P. G. (2010). A new
643Haemoproteus species (Haemosporida: Haemoproteidae) from the endemic Galapagos dove
644*Zenaida galapagoensis*, with remarks on the parasite distribution, vectors, and molecular
645diagnostics. J. Parasitol., 96(4), 783-792.

646

647Valkiūnas, G., Iezhova, T. A., Evans, E., Carlson, J. S., Martínez-Gómez, J. E., and Sehgal, R.
648N. (2013). Two new Haemoproteus species (Haemosporida: Haemoproteidae) from columbiform
649birds. J. Parasitol., 99(3), 513-521.

650

651Whitham, T. G. (1989). Plant hybrid zones as sinks for pests. Science, 1490-1493.

652

653Wyman, M. T., Locatelli, Y., Charlton, B. D., and Reby, D. (2016). Female sexual preferences
654toward conspecific and hybrid male mating calls in two species of polygynous deer, *Cervus*
655*elaphus* and *C. nippon*. Evol. Biol., 43(2), 227-241.

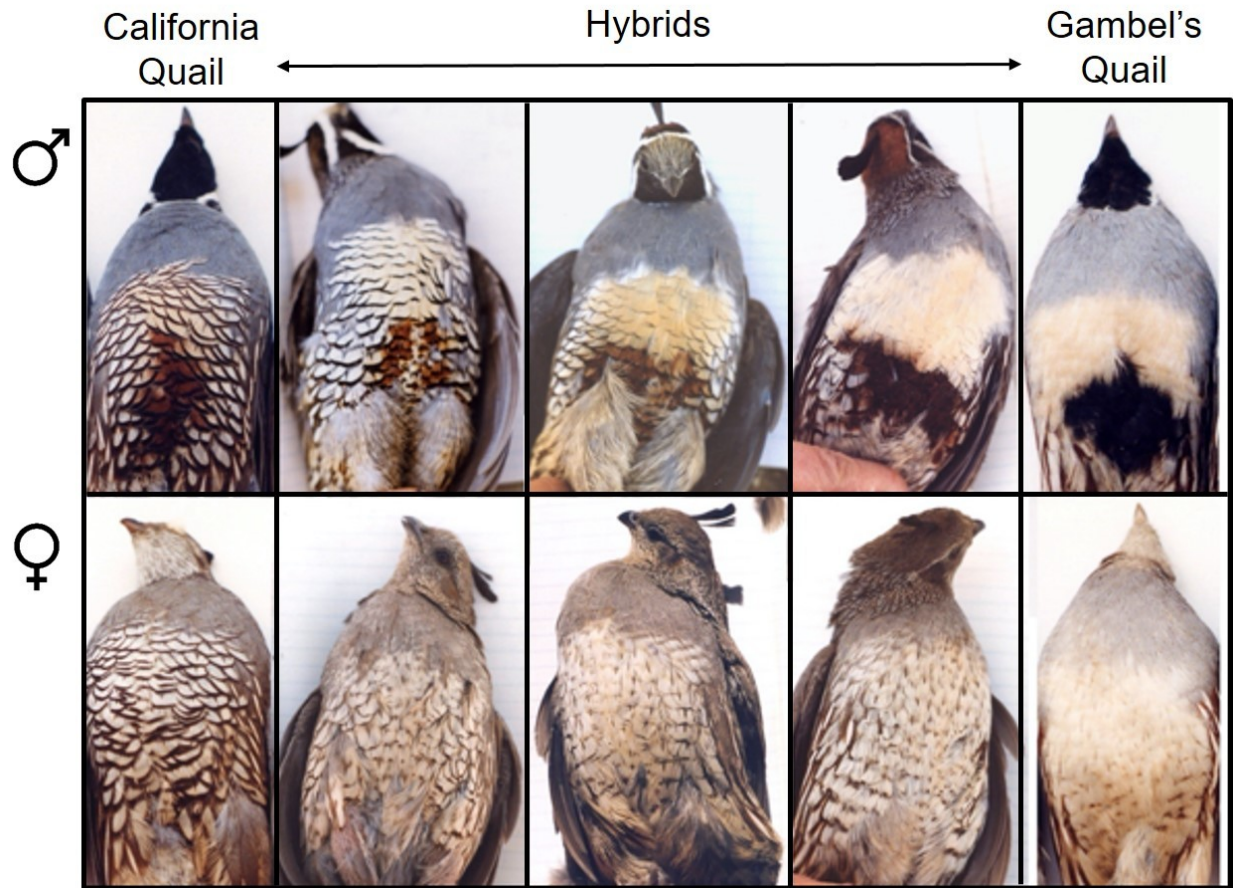
656

657Zhang, H., Xu, X., He, Z., Zheng, T., and Shao, J. (2017). De novo transcriptome analysis
658reveals insights into different mechanisms of growth and immunity in a Chinese soft-shelled
659turtle hybrid and the parental varieties. Gene, 605, 54-62.

660

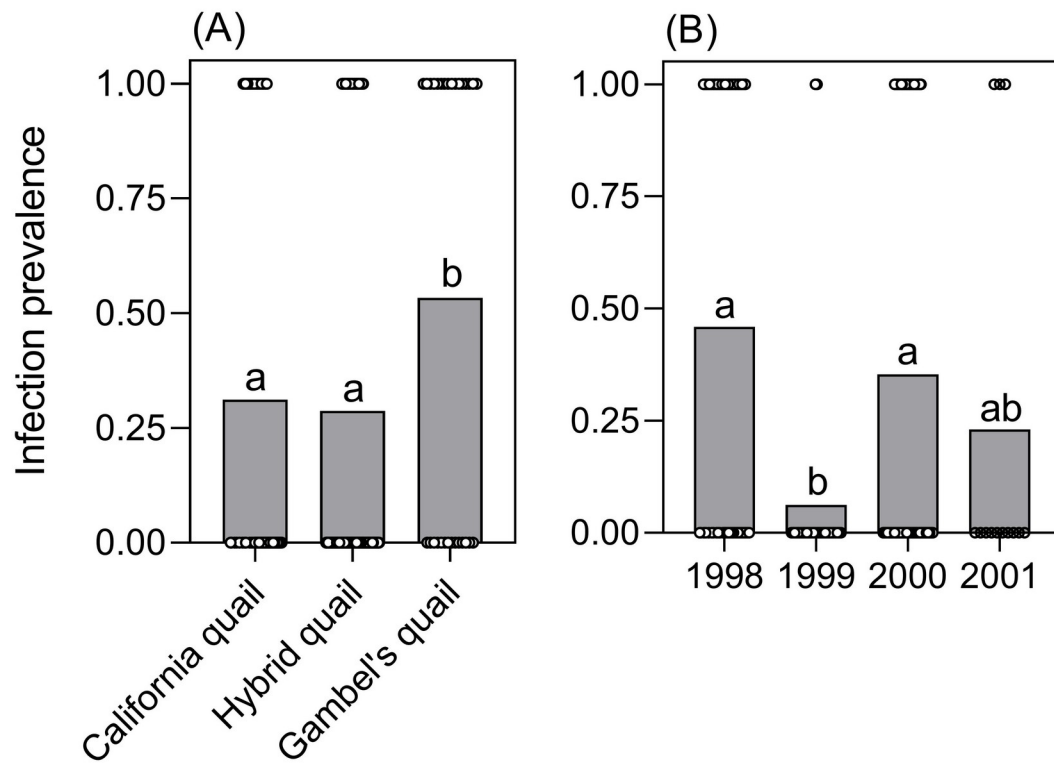
661Zink, R. M., and Blackwell, R. C. (1998). Molecular systematics of the scaled quail complex
662(genus *Callipepla*). Auk, 115(2), 394-403.

663

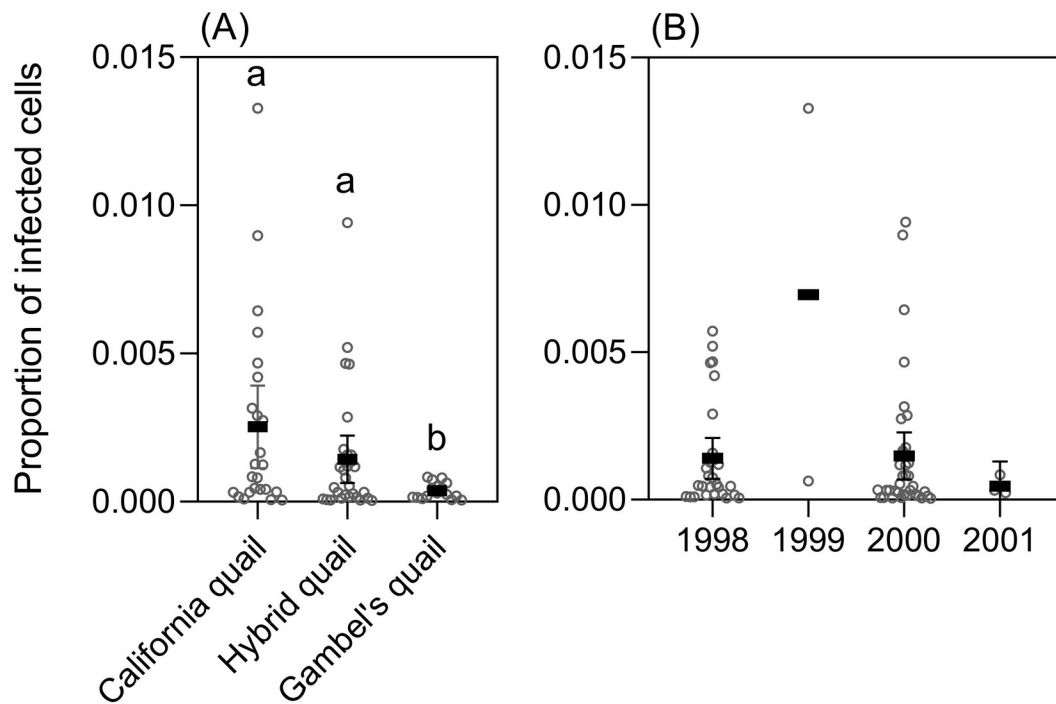


664

665**Figure 1.** The spectrum of morphological traits we examined when characterizing male (top
666row) and female (bottom row) birds as California quail (leftmost vertical panel), hybrid quail (3
667middle vertical panels), or Gambel's quail (rightmost vertical panel). Moving from California
668quail to Gambel's quail, the following patterns are observed: scaled breast, brown abdominal
669patch, overall blue body, chestnut colored cap and flanks, gray forehead and shorter plume
670versus the buffy clear breast, black belly patch, overall tan body, rust colored cap and flanks,
671blackish forehead and longer plume. Photos by JMG.



672
673 **Figure 2.** Lowercase letters denote statistical differences. **a)** Infection status of *Haemoproteus*
674 *lophortyx* in California quail, hybrid quail, and Gambel's quail when infection was determined
675 by examining at least 30 fields of view at 100x magnification. **b)** Infection status of
676 *Haemoproteus lophortyx* in 1998, 1999, 2000, and 2001 when infection was determined by
677 examining at least 30 fields of view at 100x magnification.



678
679 **Figure 3.** Lowercase letters denote statistical differences. a) Infection intensities of
680 *Haemoproteus lophortyx* in California quail, hybrid quail, and Gambel's quail when infection
681 was determined by examining at least 30 fields of view at 100x magnification. b) Infection
682 intensities of *Haemoproteus lophortyx* in 1998, 1999, 2000, and 2001 when infection was
683 determined by examining at least 30 fields of view at 100x magnification.

Table 1. Effects of species (i.e., California quail, Gambel's quail, California × Gambel's quail hybrid), year (1998, 1999, 2000, 2001), and sex on *Haemoproteus lophortyx* infection status, when infection was determined by examining at least 30 fields of view at 100x magnification (N = 208).

Predictor	Est ± SE	Z	p
Reference Class: California Quail			
Species: Hybrid Quail	-0.012 ± 0.355	-0.033	0.973
Species: Gambel's Quail	1.394 ± 0.519	2.685	0.007
Year: 1999	-2.734 ± 0.817	-3.346	0.001
Year: 2000	-0.098 ± 0.374	-0.261	0.794
Year: 2001	-0.972 ± 0.764	-1.272	0.204
Sex: Male	0.173 ± 0.341	0.507	0.612
Reference Class: Hybrid Quail			
Species: California Quail	0.012 ± 0.355	0.033	0.973
Species: Gambel's Quail	1.406 ± 0.504	2.788	0.005
Year: 1999	-2.734 ± 0.817	-3.346	0.001
Year: 2000	-0.098 ± 0.374	-0.261	0.794
Year: 2001	-0.972 ± 0.764	-1.272	0.204
Sex: Male	0.173 ± 0.341	0.507	0.612
Reference Class: Gambel's Quail			
Species: California Quail	-1.394 ± 0.519	-2.685	0.007
Species: Hybrid Quail	-1.406 ± 0.504	-2.788	0.005
Year: 1999	-2.734 ± 0.817	-3.346	0.001
Year: 2000	-0.098 ± 0.374	-0.261	0.794
Year: 2001	-0.972 ± 0.764	-1.272	0.204
Sex: Male	0.173 ± 0.341	0.507	0.612

The results presented in each subsection of this table represent the same model with different species coded as the reference class. Presenting the same model with each species coded as the reference class allows for a comparison in *H. lophortyx* infection status between each pair of species. For year, the reference class is 1998 (see Table S2 for a comparison between years). For sex, the reference class is female.

Table 2. Effects of species (i.e., California quail, Gambel's quail, California × Gambel's quail hybrid), year (1998, 1999, 2000, 2001), and sex on the intensity of *Haemoproteus lophortyx* infection (i.e., proportion of infected erythrocytes), when infection was determined by examining at least 30 fields of view at 100x magnification (N = 69).

Predictor	Est ± SE	Z	p
Reference Class: California Quail			
Species: Hybrid Quail	-0.474 ± 0.374	-1.269	0.204
Species: Gambel's Quail	-1.416 ± 0.486	-2.913	0.004
Year: 1999	0.319 ± 0.305	1.045	0.296
Year: 2000	-0.088 ± 0.290	-0.304	0.761
Year: 2001	-0.770 ± 0.827	-0.932	0.351
Sex: Male	-0.244 ± 0.356	-0.686	0.493
Reference Class: Hybrid Quail			
Species: California Quail	0.474 ± 0.374	1.269	0.204
Species: Gambel's Quail	-0.942 ± 0.469	-2.010	0.044
Year: 1999	0.319 ± 0.305	1.045	0.296
Year: 2000	-0.088 ± 0.290	-0.304	0.761
Year: 2001	-0.770 ± 0.827	-0.932	0.351
Sex: Male	-0.244 ± 0.356	-0.686	0.493
Reference Class: Gambel's Quail			
Species: California Quail	1.416 ± 0.486	2.913	0.004
Species: Hybrid Quail	0.942 ± 0.469	2.010	0.044
Year: 1999	0.319 ± 0.305	1.045	0.296
Year: 2000	-0.088 ± 0.290	-0.304	0.761
Year: 2001	-0.770 ± 0.827	-0.932	0.351
Sex: Male	-0.244 ± 0.356	-0.686	0.493

The results presented in each subsection of this table represent the same model with different species coded as the reference class. Presenting the same model with each species coded as the reference class allows for a comparison in *H. lophortyx* infection intensity between each pair of species. For year, the reference class is 1998 (see Table S3 for a comparison between years). For sex, the reference class is female.

702APPENDIX 1

703When rerunning analyses using the subset of data where at least 10,000 erythrocytes were
704examined for each sample, our results were identical to the results from models where we
705examined at least 30 fields of view at 100x magnification, with one small exception (Tables S6 –
706S9). In the model that examined at least 30 fields of view, Gambel's and hybrid quail differed
707significantly in intensity of infection. In contrast, when we limited data to 10,000 or more
708erythrocytes, this difference was marginally nonsignificant ($p = 0.050$; Table S7). As in the
709models that used data where at least 30 fields of view were examined, when at least 10,000
710erythrocytes were examined, there was an overall effect of species on infection status (Chi-
711squared = 7.778, $df = 2$, $p = 0.020$) and intensity (Chi-squared = 7.361, $df = 2$, $p = 0.025$), as well
712as an overall effect of year on infection status (Chi-squared = 18.778, $df = 3$, $p < 0.001$) and
713intensity (Chi-squared = 17.330, $df = 3$, $p = 0.001$). Sex did not appear to have an overall effect
714on either infection status (Chi-squared = 0.045, $df = 1$, $p = 0.832$) or intensity (Chi-squared =
7151.087, $df = 1$, $p = 0.297$).

716**Table S1.** Breakdown of the number of samples obtained for each species in each year.

Predictor	California	Hybrid	Gambel's
1998	25	23	13
1999	6	19	7
2000	43	51	8
2001	3	8	2

717

Table S2. Effects of species (i.e., California quail, Gambel's quail, California × Gambel's quail hybrid), year (1998, 1999, 2000, 2001), and sex on *Haemoproteus lophortyx* infection status, when infection was determined by examining at least 30 fields of view at 100x magnification (N = 208).

Predictor	Est ± SE	Z	p
Reference Class: 1998			
Species: California Quail	-1.394 ± 0.519	-2.685	0.007
Species: Hybrid Quail	-1.406 ± 0.504	-2.788	0.005
Year: 1999	-2.734 ± 0.817	-3.346	0.001
Year: 2000	-0.098 ± 0.374	-0.261	0.794
Year: 2001	-0.972 ± 0.764	-1.272	0.204
Sex: Male	0.173 ± 0.341	0.507	0.612
Reference Class: 1999			
Species: California Quail	-1.394 ± 0.519	-2.685	0.007
Species: Hybrid Quail	-1.406 ± 0.504	-2.788	0.005
Year: 1998	2.734 ± 0.817	3.346	0.001
Year: 2000	2.637 ± 0.805	3.276	0.001
Year: 2001	1.762 ± 1.050	1.679	0.093
Sex: Male	0.173 ± 0.341	0.507	0.612
Reference Class: 2000			
Species: California Quail	-1.394 ± 0.519	-2.685	0.007
Species: Hybrid Quail	-1.406 ± 0.504	-2.788	0.005
Year: 1998	0.098 ± 0.374	0.261	0.794
Year: 1999	-2.637 ± 0.805	-3.276	0.001
Year: 2001	-0.874 ± 0.765	-1.142	0.253
Sex: Male	0.173 ± 0.341	0.507	0.612
Reference Class: 2001			
Species: California Quail	-1.394 ± 0.519	-2.685	0.007
Species: Hybrid Quail	-1.406 ± 0.504	-2.788	0.005
Year: 1998	0.972 ± 0.764	1.272	0.204
Year: 1999	-1.762 ± 1.050	-1.679	0.093
Year: 2000	0.874 ± 0.765	1.142	0.253
Sex: Male	0.173 ± 0.341	0.507	0.612

The results presented in each subsection of this table represent the same model with different years coded as the reference class. Presenting the same model with each year coded as the reference class allows for a comparison between each pair of years in *H. lophortyx* infection status. The reference class for species is Gambel's quail, and the reference class for sex is female.

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Table S3. Effects of species (i.e., California quail, Gambel's quail, California × Gambel's quail hybrid), year (1998, 1999, 2000, 2001), and sex on the intensity of *Haemoproteus lophortyx* infection (i.e., proportion of infected erythrocytes), when infection was determined by examining at least 30 fields of view at 100x magnification (N = 69).

Predictor	Est ± SE	Z	p
Reference Class: 1998			
Species: California Quail	1.416 ± 0.486	2.913	0.004
Species: Hybrid Quail	0.942 ± 0.469	2.010	0.044
Year: 1999	0.319 ± 0.305	1.045	0.296
Year: 2000	-0.088 ± 0.290	-0.304	0.761
Year: 2001	-0.770 ± 0.827	-0.932	0.351
Sex: Male	-0.244 ± 0.356	-0.686	0.493
Reference Class: 1999			
Species: California Quail	1.416 ± 0.486	2.913	0.004
Species: Hybrid Quail	0.942 ± 0.469	2.010	0.044
Year: 1998	-0.319 ± 0.305	-1.045	0.296
Year: 2000	-0.407 ± 0.101	-4.048	<0.001
Year: 2001	-1.089 ± 0.811	-1.343	0.179
Sex: Male	-0.244 ± 0.356	-0.686	0.493
Reference Class: 2000			
Species: California Quail	1.416 ± 0.486	2.913	0.004
Species: Hybrid Quail	0.942 ± 0.469	2.010	0.044
Year: 1998	0.088 ± 0.290	0.304	0.761
Year: 1999	0.407 ± 0.101	4.048	<0.001
Year: 2001	-0.682 ± 0.805	-0.848	0.397
Sex: Male	-0.244 ± 0.356	-0.686	0.493
Reference Class: 2001			
Species: California Quail	1.416 ± 0.486	2.913	0.004
Species: Hybrid Quail	0.942 ± 0.469	2.010	0.044
Year: 1998	0.770 ± 0.827	0.932	0.351
Year: 1999	1.089 ± 0.811	1.343	0.179
Year: 2000	0.682 ± 0.805	0.848	0.397
Sex: Male	-0.244 ± 0.356	-0.686	0.493

The results presented in each subsection of this table represent the same model with different years coded as the reference class. Presenting the same model with each year coded as the reference class allows for a comparison between each pair of years in *H. lophortyx* infection intensity. The reference class for species is Gambel's quail, and the reference class for sex is female.

Table S4. Summary of the two mutually exclusive explanations for our observed results and how each scenario is expected to affect species barrier dynamics, as well as the potential range expansion of *H. lophortyx*.

Scenario	Species		Species Barrier Maintenance or Breakdown	Hybrid zone acts as a bridge for <i>H. lophortyx</i> to expand its range?
	Gambel's Quail	California/Hybrid Quail		
1	<p>*Host behavior/vector preference leads to higher exposure to <i>H. lophortyx</i>, which leads to higher infection prevalence</p> <p>*Have <u>higher resistance</u> to <i>H. lophortyx</i>, due to longer coevolution with the parasite, which leads to lower intensities of infection (i.e., they are better at fighting off infection once infected)</p>	<p>*Host behavior/vector preference leads to lower exposure to <i>H. lophortyx</i>, which leads to lower infection prevalence</p> <p>*Have <u>lower resistance</u> to <i>H. lophortyx</i>, due to shorter coevolution with the parasite, which leads to higher intensities of infection (i.e., they are worse at fighting off infection once infected)</p>	Species barrier maintenance is expected if the higher intensity of infection in hybrid quail leads them to have lower fitness, compared to Gambel's quail	If Gambel's quail have a longer co-evolution with <i>H. lophortyx</i> than California quail, hybrid zones may act as a bridge for the parasite to colonize California quail
2	<p>*Have similar rates of exposure to <i>H. lophortyx</i> as California/hybrid quail, but have <u>lower resistance</u>, and are therefore more likely to succumb to initial infection and are less able to completely clear infections, once infected</p> <p>*This leads to a high prevalence of low-intensity, chronic infections</p>	<p>*Have similar rates of exposure to <i>H. lophortyx</i> as Gambel's quail, but have <u>higher resistance</u>, and are therefore better at resisting initial infection and clearing an infection, once infected</p> <p>*This leads to a low prevalence of high intensity infections, given that infection intensities are highest during the initial stages of infection</p>	Species barrier breakdown is expected if chronic infection in Gambel's quail leads them to have lower fitness, compared to hybrid quail	n/a because the ecology of each quail species leads to similar rates of exposure across species

Table S5. Summary of winter precipitation across our four study years and how infection prevalence and intensity differed across years, relative to one another. We found that significantly fewer individuals were infected in 1999 than in 1998 or 2000, and, once infected, individuals expressed higher infection intensities in 1999 compared to 2000. Because 1) there was a significant difference in the proportion of infected individuals between the first and second and between the first and fourth driest years, but not between the first and third driest years and 2) the only significant difference in infection intensity occurred between 1999 and 2000, which were the two study years with the lowest levels of precipitation, it is unlikely that interannual variation in precipitation drove the differences in infection status and intensity seen between years in our study. Furthermore, given that we had a very limited sample size in 1999 and 2001, for the analysis examining the intensity of infection, both the results and any interpretations of the underlying mechanisms must be taken with caution.

Year	Precipitation (mm)	Infection Prevalence	Infection Intensity
1998	149.2	High	-
1999	28.3	Low	High
2000	59.3	High	Low
2001	81.1	-	-

We calculated winter precipitation from December 1 (of the preceding year) – February 28 for each year of our study using data acquired from the University of California, Boyd Deep Canyon Desert Research Center (<http://deepcanyon.ucnrs.org>).

Table S6. Effects of species (i.e., California quail, Gambel's quail, California × Gambel's quail hybrid), year (1998, 1999, 2000, 2001), and sex on *Haemoproteus lophortyx* infection status, when infection was determined by examining at least 10,000 erythrocytes (N = 189).

Predictor	Est ± SE	Z	p
Reference Class: California Quail			
Species: Hybrid Quail	0.009 ± 0.369	0.024	0.981
Species: Gambel's Quail	1.370 ± 0.546	2.509	0.012
Year: 1999	-2.702 ± 0.824	-3.278	0.001
Year: 2000	-0.272 ± 0.391	-0.696	0.486
Year: 2001	-1.044 ± 0.767	-1.362	0.173
Sex: Male	0.085 ± 0.356	0.239	0.811
Reference Class: Hybrid Quail			
Species: California Quail	-0.009 ± 0.369	-0.024	0.981
Species: Gambel's Quail	1.361 ± 0.523	-2.602	0.009
Year: 1999	-2.702 ± 0.824	-3.278	0.001
Year: 2000	-0.272 ± 0.391	-0.696	0.486
Year: 2001	-1.044 ± 0.767	-1.362	0.173
Sex: Male	0.085 ± 0.356	0.239	0.811
Reference Class: Gambel's Quail			
Species: California Quail	-1.370 ± 0.546	-2.509	0.012
Species: Hybrid Quail	-1.361 ± 0.523	-2.602	0.009
Year: 1999	-2.702 ± 0.824	-3.278	0.001
Year: 2000	-0.272 ± 0.391	-0.696	0.486
Year: 2001	-1.044 ± 0.767	-1.362	0.173
Sex: Male	0.085 ± 0.356	0.239	0.811

The results presented in each subsection of this table represent the same model with different species coded as the reference class. Presenting the same model with each species coded as the reference class allows for a comparison in *H. lophortyx* infection status between each pair of species. For year, the reference class is 1998 (see Table S8 for a comparison between years). For sex, the reference class is female.

Table S7. Effects of species (i.e., California quail, Gambel's quail, California × Gambel's quail hybrid), year (1998, 1999, 2000, 2001), and sex on the intensity of *Haemoproteus lophortyx* infection (i.e., proportion of infected erythrocytes), when infection was determined by examining at least 10,000 erythrocytes (N = 63). Fixed effects that differ in significance between models using data where at least 10,000 erythrocytes were examined and models using data where at least 30 fields of view at 100x magnification were examined are shaded in grey.

Predictor	Est ± SE	Z	p
Reference Class: California Quail			
Species: Hybrid Quail	-0.494 ± 0.407	-1.216	0.224
Species: Gambel's Quail	-1.482 ± 0.537	-2.762	0.006
Year: 1999	0.370 ± 0.319	1.160	0.246
Year: 2000	-0.037 ± 0.304	-0.123	0.903
Year: 2001	-0.753 ± 0.857	-0.879	0.380
Sex: Male	-0.399 ± 0.391	-1.020	0.308
Reference Class: Hybrid Quail			
Species: California Quail	0.494 ± 0.407	1.216	0.224
Species: Gambel's Quail	-0.988 ± 0.504	-1.959	0.050
Year: 1999	0.370 ± 0.319	1.160	0.246
Year: 2000	-0.037 ± 0.304	-0.123	0.903
Year: 2001	-0.753 ± 0.857	-0.879	0.380
Sex: Male	-0.399 ± 0.391	-1.020	0.308
Reference Class: Gambel's Quail			
Species: California Quail	1.482 ± 0.537	2.762	0.006
Species: Hybrid Quail	0.988 ± 0.504	1.959	0.050
Year: 1999	0.370 ± 0.319	1.160	0.246
Year: 2000	-0.037 ± 0.304	-0.123	0.903
Year: 2001	-0.753 ± 0.857	-0.879	0.380
Sex: Male	-0.399 ± 0.391	-1.020	0.308

The results presented in each subsection of this table represent the same model with different species coded as the reference class. Presenting the same model with each species coded as the reference class allows for a comparison in *H. lophortyx* infection intensity between each pair of species. For year, the reference class is 1998 (see Table S9 for a comparison between years). For sex, the reference class is female.

Table S8. Effects of species (i.e., California quail, Gambel's quail, California × Gambel's quail hybrid), year (1998, 1999, 2000, 2001), and sex on *Haemoproteus lophortyx* infection status, when infection was determined by examining at least 10,000 erythrocytes (N = 189).

Predictor	Est ± SE	Z	p
Reference Class: 1998			
Species: California Quail	-1.370 ± 0.546	-2.509	0.012
Species: Hybrid Quail	-1.361 ± 0.523	-2.602	0.009
Year: 1999	-2.702 ± 0.824	-3.278	0.001
Year: 2000	-0.272 ± 0.391	-0.696	0.486
Year: 2001	-1.044 ± 0.767	-1.362	0.173
Sex: Male	0.085 ± 0.356	0.239	0.811
Reference Class: 1999			
Species: California Quail	-1.370 ± 0.546	-2.509	0.012
Species: Hybrid Quail	-1.361 ± 0.523	-2.602	0.009
Year: 1998	2.702 ± 0.824	3.278	0.001
Year: 2000	2.430 ± 0.807	3.012	0.003
Year: 2001	1.658 ± 1.044	1.587	0.112
Sex: Male	0.085 ± 0.356	0.239	0.811
Reference Class: 2000			
Species: California Quail	-1.370 ± 0.546	-2.509	0.012
Species: Hybrid Quail	-1.361 ± 0.523	-2.602	0.009
Year: 1998	0.272 ± 0.391	0.696	0.486
Year: 1999	-2.430 ± 0.807	-3.012	0.003
Year: 2001	-0.772 ± 0.755	-1.023	0.306
Sex: Male	0.085 ± 0.356	0.239	0.811
Reference Class: 2001			
Species: California Quail	-1.370 ± 0.546	-2.509	0.012
Species: Hybrid Quail	-1.361 ± 0.523	-2.602	0.009
Year: 1998	1.044 ± 0.767	1.362	0.173
Year: 1999	-1.658 ± 1.044	-1.587	0.112
Year: 2000	0.772 ± 0.755	1.023	0.306
Sex: Male	0.085 ± 0.356	0.239	0.811

The results presented in each subsection of this table represent the same model with different years coded as the reference class. Presenting the same model with each year coded as the reference class allows for a comparison between each pair of years in *H. lophortyx* infection status. The reference class for species is Gambel's quail, and the reference class for sex is female.

Table S9. Effects of species (i.e., California quail, Gambel's quail, California × Gambel's quail hybrid), year (1998, 1999, 2000, 2001), and sex on the intensity of *Haemoproteus lophortyx* infection (i.e., proportion of infected erythrocytes), when infection was determined by examining at least 10,000 erythrocytes (N = 63). Fixed effects that differ in significance between models using data where at least 10,000 erythrocytes were examined and models using data where at least 30 fields of view at 100x magnification were examined are shaded in grey.

Predictor	Est ± SE	Z	p
Reference Class: 1998			
Species: California Quail	1.482 ± 0.537	2.762	0.006
Species: Hybrid Quail	0.988 ± 0.504	1.959	0.050
Year: 1999	0.370 ± 0.319	1.160	0.246
Year: 2000	-0.037 ± 0.304	-0.123	0.903
Year: 2001	-0.753 ± 0.857	-0.879	0.380
Sex: Male	-0.399 ± 0.391	-1.020	0.308
Reference Class: 1999			
Species: California Quail	1.482 ± 0.537	2.762	0.006
Species: Hybrid Quail	0.988 ± 0.504	1.959	0.050
Year: 1998	-0.370 ± 0.319	-1.160	0.246
Year: 2000	-0.407 ± 0.101	-4.050	<0.001
Year: 2001	-1.123 ± 0.837	-1.342	0.180
Sex: Male	-0.399 ± 0.391	-1.020	0.308
Reference Class: 2000			
Species: California Quail	1.482 ± 0.537	2.762	0.006
Species: Hybrid Quail	0.988 ± 0.504	1.959	0.050
Year: 1998	0.037 ± 0.304	0.123	0.903
Year: 1999	0.407 ± 0.101	4.050	<0.001
Year: 2001	-0.716 ± 0.831	-0.862	0.389
Sex: Male	-0.399 ± 0.391	-1.020	0.308
Reference Class: 2001			
Species: California Quail	1.482 ± 0.537	2.762	0.006
Species: Hybrid Quail	0.988 ± 0.504	1.959	0.050
Year: 1998	0.753 ± 0.857	0.879	0.380
Year: 1999	1.123 ± 0.837	1.342	0.180
Year: 2000	0.716 ± 0.831	0.862	0.389
Sex: Male	-0.399 ± 0.391	-1.020	0.308

The results presented in each subsection of this table represent the same model with different years coded as the reference class. Presenting the same model with each year coded as the reference class allows for a comparison between each pair of years in *H. lophortyx* infection intensity. The reference class for species is Gambel's quail, and the reference class for sex is female.