

Elucidation of ligand binding and dimerization of NADPH:protochlorophyllide (Pchl_{id}) oxidoreductase (POR) from pea (*Pisum sativum* L.) by structural analysis and simulations

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Abstract

NADPH:protochlorophyllide (Pchl_{id}) oxidoreductase (POR) is a key enzyme of chlorophyll biosynthesis in angiosperms. It is one of few known photoenzymes, which catalyzes the light-activated trans-reduction of the C17-C18 double bond of Pchl_{id}'s porphyrin ring. Due to the light requirement, dark-grown angiosperms cannot synthesize chlorophyll. No crystal structure of POR is available, so to improve understanding of the protein's three-dimensional structure, its dimerization, and binding of ligands (both the cofactor NADPH and substrate Pchl_{id}), we computationally investigated the sequence and structural relationships among homologous proteins identified through database searches. The results indicate that $\alpha 4$ and $\alpha 7$ helices of monomers form the interface of POR dimers. On the basis of conserved residues, we predicted 11 functionally important amino acids that play important roles in POR binding to NADPH. Structural comparison of available crystal structures revealed that they participate in formation of binding pockets that accommodate the Pchl_{id} ligand, and that five atoms of the closed tetrapyrrole are involved in non-bonding interactions. However, we detected no clear pattern in the physico-chemical characteristics of the amino acids they interact with. Thus, we hypothesize that interactions of these atoms in the Pchl_{id} porphyrin ring are important to hold the ligand within the POR binding site. Analysis of Pchl_{id} binding in POR by molecular docking and PELE simulations revealed that the orientation of the nicotinamide group is important for Pchl_{id} binding. These findings highlight the complexity of interactions of porphyrin-containing ligands with proteins, and we suggest that fit-inducing processes play important roles in POR-Pchl_{id} interactions.

Introduction

Chlorophyll biosynthesis is crucial for photosynthesis, and thus photoautotrophic growth of land plants. It is subject to complex regulation, involving various anterograde and retrograde

signaling pathways and several feedback mechanisms (Solymosi and Schoefs 2010, Liu et al. 2017). One of the key regulatory steps is the penultimate step in chlorophyll formation, the reduction of protochlorophyllide (Pchl_{id}) to chlorophyllide (Chl_{id}). Two nonhomologous enzymes with completely different structures and reaction mechanisms have evolved that catalyze this process (Vedalankar and Tripathy 2019). Dark operative (or light-independent) Pchl_{id} oxidoreductase (DPOR) putatively evolved from a nitrogenase-like enzyme of methanogenic archaea under anoxygenic conditions (Vedalankar and Tripathy 2019). This is a multimeric, oxygen-sensitive enzyme composed of three different subunits encoded by the plastid genome. The other enzyme is nuclear-encoded and oxygen-insensitive, but requires light for its activity and is thus called light-dependent NADPH:Pchl_{id} oxidoreductase (LPOR, hereafter simply POR, for convenience) (Solymosi and Schoefs 2010, Gabruk and Mysliwa-Kurdziel 2015). Anoxygenic photosynthetic bacteria contain only DPOR, many organisms (most green algae, bryophytes, pteridophytes and gymnosperms) contain both types of enzymes, but angiosperms and some other organisms have lost DPOR during evolution and thus the ability to produce chlorophyll in the dark (Schoefs and Franck 2003, Vedalankar and Tripathy 2019).

Without light-induced developmental signals and chlorophyll, chloroplast formation is also inhibited in dark-germinated angiosperm seedlings. Instead, proplastids become etioplasts, containing three-dimensional networks of tubular membrane structures called prolamellar bodies (PLBs) (Solymosi and Schoefs 2008, 2010). PLBs are also found in plants grown under natural light conditions, e.g. in fruit pericarps and leaf buds (Solymosi and Schoefs 2008, 2010, Solymosi and Aronsson 2013). The lipid composition of PLBs favors formation of the highly regular and curved network of PLB membranes, but is not the crucial factor (Selstam 1998, Jarvis et al. 2000, Solymosi and Schoefs 2008). Key identified factors include the presence and supramolecular organization (dimerization or oligomerization) of ternary complexes of POR

with Pchl_a and NADPH (Ryberg and Sundqvist 1988, Böddi et al. 1989, Wiktorsson et al. 1993, Sperling et al. 1997, Ouazzani-Chahdi et al. 1998, Sperling et al. 1998, Frick et al. 2003, Masuda et al. 2003, Solymosi and Schoefs 2010). However, the mechanisms involved in POR's interactions with PLB lipids and carotenoids that stimulate formation of the highly regular network of PLB membranes are poorly understood as no high-resolution structure of POR is available. Information about its structure is limited to relative amounts of secondary structure elements, experimentally determined by circular dichroism (Birve et al. 1996), and predictions of secondary and tertiary structures obtained using various bioinformatic tools (Birve et al. 1996, Dahlin et al. 1999, Townley et al. 2001, Buhr et al. 2008, Gabruk et al. 2012, Menon et al. 2016).

Several *in vivo* and *in vitro* washing experiments and mutagenesis experiments have been performed in attempts to elucidate the strength of POR's membrane association. The results show that its association with thylakoid or etioplast inner membranes requires NADPH and ATP (Dahlin et al. 1995, Engdahl et al. 2001). Furthermore, charged amino acids, cysteine (Cys) residues and the C-terminal are important for membrane association (Dahlin et al. 1999, Aronsson et al. 2001). In addition, various salts (Grevby et al. 1989, Widell-Wigge and Selstam 1990, Dahlin et al. 1995, Engdahl et al. 2001), detergents (Grevby et al. 1989, Selstam and Widell-Wigge 1989) and proteases (Lütz and Tönissen 1984, Dahlin et al. 1995, Engdahl et al. 2001) have been used in washing experiments. These studies have shown that POR is firmly attached to PLBs and PTs, but more loosely attached to thylakoids, indicating that POR's affinity for the membranes may decline during greening (Aronsson et al. 2003).

The difficulties of releasing POR from PLBs and PTs using salts and detergents led to the conclusion that POR must be an integral membrane protein (Grevby et al. 1989, Widell-Wigge and Selstam 1990). Integral membrane proteins are generally classed as integral transmembrane proteins with one or more regions spanning the membrane or integral

monotopic membrane proteins permanently attached to one side of a membrane (Lomize et al. 2007). However, the suggestion that POR is an integral transmembrane protein was subsequently withdrawn (Widell-Wigge 1993) in the light of its deduced amino acid sequence (Schulz et al. 1989, Darrah et al. 1990).

Today, numerous sequences of POR from diverse plant groups have been characterized (Masuda and Takamiya 2004, Gabruk and Mysliwa-Kurdziel 2015, 2020). Two differently regulated POR isoforms, designated PORA and PORB, have been characterized in barley (*Hordeum vulgare*) (Holtorf et al. 1995), *Arabidopsis thaliana* (Armstrong et al. 1995), wheat (*Triticum aestivum*) (Blomqvist et al. 2008) and rice (*Oryza sativa*) (Kwon et al. 2017). PORA accumulates in dark-grown tissues and is active during the first hours of greening while PORB is active throughout the life of the plant. A third isozyme, PORC, has only been found, to date, in light-grown *A. thaliana* (Oosawa et al. 2000) and may be involved in regulation of oxidative stress (Pattanayak and Tripathy 2011). POR sequences generally share high sequence similarity and have high contents of basic and hydrophobic amino acids (Schoefs and Franck 2003). However, no evidence of hydrophobic segments long enough to span a membrane have been detected in hydropathy plots (Benli et al. 1991, Spano et al. 1992, Timko 1993), which have been persistently cited since their publication. However, these plots were based exclusively on searches of sequences for a sufficiently long hydrophobic stretch of residues to traverse a membrane based on the hydrophobicity scale (Kyte and Doolittle 1982). Refinements of the methodology have included consideration of statistical and structural information about known transmembrane helices, such as the presence of specific sequence patterns and motifs, the packing of helix bundles, and properties of inter-helical residue interactions. Methods considering this kind of information are referred to as advanced transmembrane methods and generally perform better than simple hydrophobicity scale-based methods according to (Chen et al. 2002).

Some early experiments involving attempts to isolate and characterize photoactive POR complexes showed that they may be associated with carotenoids, particularly zeaxanthin and violaxanthin (Ouazzani-Chahdi et al. 1998), or carotenoids may play important roles in their membrane association (Denev et al. 2005). *In vitro* reconstitution experiments have also shown the importance of various membrane lipids in the proper organization of the photoactive complexes (Gabruk et al. 2017), which as yet have not been reconstituted without lipids, and thus membranes. Similarly, data on lipid biosynthesis mutants have shown that membrane lipids are crucial for proper POR activity, which requires its oligomerization as well as membrane association (Fujii et al. 2017, 2018).

As already stated, despite its importance in chlorophyll biosynthesis, and thus plant metabolism, as well as its highly interesting photochemical catalytic activity, there is still no available crystal structure of POR and we do not fully understand its structure and oligomerization. Therefore, as reported here, we performed a detailed bioinformatic analysis of POR, focusing on its potential dimerization and binding of Pchl_{ide}. The modelling and simulations indicate that $\alpha 4$ and $\alpha 7$ helices of the POR protein form the dimer interface, and that Pchl_{ide} binds close to the pro-S hydrogen of the nicotinamide ring of NADPH and Tyr191, which is involved in the catalytic mechanism.

Materials and Methods

Sequence and structural analysis

The amino acid sequence of the POR protein of pea (*Pisum sativum* L.) was retrieved from the Uniprot database (Q01289). Its domain pattern was analyzed using the protein families (PFAM) (Finn et al. 2014) and Prosite (Sigrist et al. 2013) databases. PSI-Blast (Oda et al. 2017) was used to search the POR sequence against the PDB database (Burley et al. 2017) to identify

homologous proteins with crystal structures, which were superimposed using the PDBefold server (Krissinel and Henrick 2004). Structure-based sequence alignment was applied to identify regions of these proteins that were structurally conserved, despite low sequence identity. The POR sequence was then aligned with the multiple sequence alignment of these templates using the MAFFT program (Katoh et al. 2017).

Homology model

Homologs of POR were identified by a BLAST search of the PDB database using the retrieved pea sequence as candidate templates for model building. Nine with crystal structures were identified that bind NADPH (PDB_ID: 3WXB, 2JAH, 3P19, 3CSD, 3SJU, 1XU9, 3O26, 1WMA, and 1N5D), and the crystallographically determined structure of chicken carbonyl reductase (3WXB) was selected as a template, as it has high resolution (1.98 Å), bound cognate ligand (NADPH), dimeric form, and sequence identity. An optimized model of POR was built using Modeller v. 9.12 (Webb and Sali 2016) with the sequence alignment described above. Models of both monomeric and dimeric forms of the protein were generated. Ten models were constructed and the best, according to Modeller's atomic statistical potential function, was selected. Multiple templates were used to model the long insertions of amino acids in POR. The model was validated using a Ramachandran plot, VERIFY3D (Eisenberg et al. 1997) and ProSA-web (Wiederstein and Sippl 2007).

Molecular Docking

Molecular docking of NADPH with POR was applied to study its mode of interaction with its POR binding site using GOLD (Genetic Optimisation for Ligand Docking) v. 5.38. This is a widely used protein-ligand docking program (Verdonk et al. 2003), which applies a genetic algorithm for protein-ligand docking that allows full ligand and partial protein flexibility. The

homology model of POR with added hydrogen atoms was used as the receptor, the NADPH ligand model was corrected with the Auto-Edit Ligand option, and 200 conformers were generated using the Conformation Generation module of Mercury Software (Macrae et al. 2008). The NADPH active site was identified and reconfirmed by superposition and multiple structure-based sequence alignments of POR with the nine selected NADPH-bound crystal structures (described above). Based on the alignment, the binding site of NADPH was mapped onto the POR protein and used as input for GOLD calculation. Default genetic algorithm (GA) settings were used for docking, generating 10 poses for each of the 200 conformers. Early termination of the GA runs was allowed when the root mean square deviations (RMSD values) of the top three GA solutions were $< 1.5 \text{ \AA}$. The best pose of the docked ligand was selected based on GOLD score and calculated RMSD between the reference NADPH and crystal structure of 3WXB. The docking was submitted to the Albiorix cluster available at the University of Gothenburg (<http://albiorix.bioenv.gu.se/>).

PELE simulations

While the NADPH binding site can be inferred from homologs, Pchlide binding information is not available. Thus, to map Pchlide's binding to POR we used Protein Energy Landscape Exploration (PELE) Monte Carlo (MC) molecular simulation software (Borrelli et al. 2005). PELE has been developed for analyzing protein-ligand interactions, through functions such as binding site search and refinement, ligand migration, and fragment growth (Gygli et al. 2017, Lecina et al. 2017, Perez et al. 2020). Each PELE MC step consists of a ligand perturbation based on a (random) translation and rotation, followed by a backbone protein perturbation using a low frequency subset of normal modes, side chain prediction and, finally, a global minimization. The resulting structure is accepted or rejected by applying a Metropolis criterion, based on energy calculations generated using a OPLS2005 force field with an implicit solvent,

as explained in detail by (Gilabert et al. 2018). In the applied approach, which we call the AdaptivePELE enhanced sampling method (Lecina et al. 2017), several iterations are run, consisting of a short PELE simulation, then the output is clustered and some of the clusters are selected to start the next epoch. All accepted MC steps are clustered with the leader algorithm using the ligand RMSD (after protein superposition). The following initial structures are then selected using criteria deemed appropriate, for example, the clusters' populations or value of some metric such as solvent-accessible surface area (SASA) or interaction energy. AdaptivePELE can be an order of magnitude faster than PELE simulations, allowing us to map complex unbiased binding mechanisms in less than an hour.

Two PELE simulations were performed. First a global simulation, in which the ligand was allowed to explore the entire protein surface by using large translations and rotations, as well as 20 different initial structures randomly distributed at the enzyme surface. In the second simulation local sampling was applied, with reduced ligand translations and rotations together with application of a steering vector (translation direction) for two consecutive MC steps. The aim of the second simulation was to refine the binding site, by allowing the ligand to enter inner cavities.

Results and Discussion

Sequence search

POR is a member of the short-chain dehydrogenase/reductase (SDR) superfamily of proteins. Accordingly, a Blast sequence homology search against the PDB database showed POR to be highly similar to other members of this family, which have a highly conserved nucleotide-binding Rossmann fold (Kavanagh et al. 2008), despite sharing very low sequence identity (Jörnvall et al. 1995, Kallberg et al. 2002). However, based on the Blast search results, we selected two crystal structures: carbonyl reductase from chicken fatty liver (PDB_ID: 3WXB)

(Fukuda et al. 2015) and salutaridine reductase from *Papaver somniferum* (opium poppy, PDB_ID: 3O26) (Higashi et al. 2011) from the hits, based on total sequence coverage and presence of a bound NADPH cofactor.

Structural alignment of POR-homologous SDR proteins

POR belongs to the SDR family that includes enzymes that utilize both NADPH and NADH as cofactors. However, the enzymes have small structural differences and residue preferences within their cofactor binding motif that may account for the differences in specificities for these two cofactors (Baker 1994, Wilks and Timko 1995). Presence of a basic amino acid in the glycine-rich motif immediately preceding the second conserved glycine has been reported in NADP(H)-specific enzymes (Kallberg et al. 2002). Other studies indicate that POR of a *Synechocystis* species can bind to NADPH, but not NADH (Griffiths 1980, Heyes et al. 2000). Thus, a key task was to identify structural homologs bound with NADPH as a cofactor to find residues that play important roles in its binding in the homologs and, potentially, POR.

Using the selected template, 3WXB, a Blast search against the PDB database (using the DALI server) identified more than 1000 structures with Z-scores between 42.3 and 12. From this set we selected SDR protein structures that met two criteria: possession of the adh_short domain (PFAM ID: PF00106) and binding of the NADPH cofactor (Figure 1A). SDR protein family members have structural differences that depend on whether they bind to NAD(H) or NADP(H) (Kallberg et al. 2002). POR has two conserved regions: a consensus Gly-X-X-X-Gly-X motif associated with cofactor binding (Ghosh et al. 1995) and a region that has been strictly conserved through evolution in the SDR family with a Tyr-X-X-X-Lys sequence that is essential for enzyme activity (Persson et al. 1991). However, in the set of nine structures considered as candidate templates here, Arg, Ser and Lys precede the second conserved Gly. The presence of the Ser residue before the second Gly of the Gly motif confers preference for

NADP(H) binding in the human 17 β -hydroxysteroid dehydrogenase type 1 enzyme (Huang et al. 2001).

The pairwise sequence identity and RMSD of these nine proteins range from 13.2% to 76% and 0.5 Å to 2.8 Å, respectively (Table 1). Structural superposition of these structures shows that they have a well-conserved Rossmann fold, despite its very low sequence identity (Figure 1B). Each of these proteins has a α/β doubly wound structure, in which seven parallel β strands that form a β sheet are sandwiched between two arrays of three α helices. Similarly, the bound conformation of NADPH within pockets of these protein structures was found to have an RMSD less than 0.7 Å (Figure 1C). Thus, the bound conformation of NADPH is well conserved among these proteins, despite low overall sequence identity. Further, we analyzed the interaction between each of the nine protein structures and NADPH. Aligned residues of the proteins at positions corresponding to Asn15, Arg16, Ile18, Arg41, Arg47, Val68, Asn96, Thr178, Lys182, Val211, Thr213 in 3WXB were observed to form hydrogen bonds with the NADPH cofactor in all nine structures (Figure 2A, B). Thus, hydrogen bond interaction involving these residues is conserved in all nine NADPH complexes (Table 2). In addition, 10 atoms of the ligand (O7N, O2D, O3D, O1N, O2N, O3B, O1X, O2X, O3X, and N1A according to standard labelling; reference) form hydrogen bonds in all nine structures (Figure S1). We observed that the adenine group is captured near the loop connecting β 3 and α 3 via interactions with Glu67 and Val68 (in 3WXB and corresponding residues in the other proteins), while the ribose phosphate interacts with Asn15 and Arg41 located on the loop connecting β 1 and α 1, immediately after β 3. The nicotinamide moiety appears to interact with Val211 and Thr213 in the loop after β 6.

We further aligned 14 POR sequences from different organisms with the nine sequences associated with the nine crystal structures, then mapped the NADPH binding site onto the POR sequences (Figure S2). We found that eight of the 24 residues identified in the

binding site of NADPH of co-crystal structures were identical to residues at corresponding positions in the POR enzymes. Of these eight residues, four form hydrogen bonds with ribose groups of the NADPH cofactor. We infer from results of this analysis that these residues play an important role in the geometric alignment of the NADPH cofactor within active sites of these SDR enzymes.

The 3D structure of POR was predicted using Modeller software v. 9.19 and the NADPH bound co-crystal structure of carbonyl reductase from chicken fatty liver (PDB_ID:3WXB) as template. 3WXB was used for this purpose because both POR and 3WXB proteins have the short-chain dehydrogenase domain and the crystal structure of dimeric 3WXB bound with NADPH provides insights into the spatial arrangement of the active site residues and two monomeric subunits. However, the sequence identity between the target and template is low (21.19%) so we applied profile-based alignment to improve the alignment, as follows. First, the nine crystal structures and set of 14 POR sequences were subjected to structure-based sequence alignment using PDBeFold and sequence alignment using Clustal Omega. Both alignments were then combined using the Merge program provided by the MAFFT server, and the merged alignment was then used as input for modeling POR by the Modeller software. The most noticeable differences from the sequence alignment between POR and the template are the two long amino acid insertions, one 38 (between $\beta 5$ and $\alpha 7$) and the other 22 residues long (between $\beta 6$ and $\alpha 8$) were observed in the central region of POR. The most pronounced differences in sequences between POR and the template are two long amino acid insertions, one of 38 residues (between $\beta 5$ and $\alpha 7$) and one of 22 residues (between $\beta 6$ and $\alpha 8$) in the central region of POR.

We modelled POR by first modelling the POR monomer (residues 84-371) then the dimer using both one template (3WXB) and a combination of multiple templates to cover the long amino acid insertions. The quality and accuracy of the model was then assessed using a Ramachandran plot, Verify3D and ProSA. In the Ramachandran plot of the POR monomer

about 85.9, 13 and 0.8% of the residues are in the most favored, generously allowed and disallowed regions, respectively (Figure 3A), according to criteria presented by Reference (Date). Verify 3D indicated that about 77.78% of the modelled structure is compatible with its amino acid sequence and the Z score for the model calculated using ProSA is -5.87. Following this validation, the modelled POR structure was superposed onto the template 3WXB, resulting in an RMSD value of 1.34 Å (Figure 3B). Thus, the overall topology of the modelled structure is similar to the template structure except for the two long insertions. The overall analysis confirms that the model generated is reliable and can be used for further analysis.

Most SDR enzymes reportedly occur predominantly in dimeric and tetrameric forms in nature and the two long α -helices are involved in oligomerization (Jörnvall et al. 1995). Oligomerization of POR in an experimental reaction mixture was recently reported (Gabruk et al. 2015), and dimerization or oligomerization of POR proteins isolated from *in vivo* systems has been observed both with no pretreatment (Ouazzani-Chahdi et al. 1998) and following use of protein cross-linkers (Wiktorsson et al. 1993). The interface between two monomers of a protein in a dimer or oligomer is considered to be conserved if it tends to overlap strongly in similar protein structures (Uguzzoni et al. 2017).

In the 3WXB template, the two monomers interact with each other through α 4 and α 5 helices of both monomers. Most of the residues involved in dimerization of the template are conserved in POR, according to the sequence alignment. Thus, we modelled the dimeric form of POR using 3WXB in its dimeric form as the template (Figure 4A). To further confirm the dimerization of POR, we also docked the two monomers using the MZDOCK server. The top three predictions of MZDOCK had similar dimeric interfaces to that of the modelled POR dimer (Figure 4B), corroborating the inference that α 4 and α 7 helices of POR form the dimer interface between pairs of POR monomers.

Docking of the cofactor NADPH in the POR binding site

Molecular docking was applied to further elucidate the binding of NADPH in the active site of both chains of POR dimers. A large number of conformers were generated for NADPH using CCDC Mercury software to identify the most stable NADPH binding pose, and the identified NADPH-binding residues in POR (Table 2) were used to guide the POR-NADPH docking analysis. The best docking pose of NADPH in both chains of POR was identified based on GOLD scores and RMSD values between the docked pose and template NADPH, following superposition (Figure 5). The GOLD scores for NADPH in the A and B monomeric chains of POR were 389 and 393 kJ/mol respectively. The NADPH docked *in silico* in the POR binding site forms eight hydrogen bonds with Gly10, Ser12, Ser13, Asn88, Val141, Gly142, Ile225, and Thr227. Of these eight H-bonding residues, five (Ser12, Ser13, Asn88, Ile225 and Thr227) are the ones conserved with respect to the 3WXB crystal structure.

Structural similarity of protochlorophyllide (Pchl_{id}) with other molecules

Pchl_{id} is a precursor of chlorophyll, and is structurally similar to chlorophyll but lacks the phytol side chain of chlorophyll and has one more double bond, between C17 and C18 of the porphyrin ring. Pchl_{id} is the major pigment accumulating in dark-grown plants (Solymosi and Schoefs 2008, 2010), and the substrate of POR, which catalyzes reduction of the C17–C18 double bond of Pchl_{id} to form Chl_{id}, an essential regulatory step in chlorophyll biosynthesis (Scrutton et al. 2012). Many studies have shown that the POR-NADPH-Pchl_{id} ternary complex accumulates *in vivo* when plants are germinating in the absence of light (Lebedev and Timko 1998, Raskin and Schwartz 2002, Schoefs and Franck 2003, Masuda and Takamiya 2004).

The availability of numerous structures of protein complexes in the PDB database enables comparison of the binding pockets of proteins with different structures that bind similar

ligands (Gold and Jackson 2006, Kahraman et al. 2007, Minai et al. 2008). Crystal structures of the light-independent (dark-operative) Pchlide oxidoreductase (DPOR) enzymes bound with Pchlide from *Rhodobacter capsulatus* (PDB_ID:3AEK) and *Prochlorococcus marinus* (PDB_ID: 2YNM) have been reported (Muraki et al. 2010, Moser et al. 2013). Pchlide binds between the interface of protein subunits A, B and D in the 3AEK structure, and protein subunits C and D in the 2YNM structure. Superposing the two structures 3AEK (A, B) and 2YNM (C, D) revealed high structural similarity (RMSD 1.6 Å), together with high conservation in the binding site and conformation of Pchlide (Table 3).

To identify the Pchlide binding pocket in POR based on pocket similarity, we performed a chemical structure search using the Pchlide structure as a query against the PDB database with a 90% similarity cut-off. We searched for pockets that accommodate chemically similar or identical ligands, and identified Pchlide (PMR, according to the PDB 3-letter code) bound to the DPOR enzymes and three other chemical structures: chlorophyll b (CHL), chlorophyll a (CLA) and bacteriochlorophyll g (GB0) that share some structural similarity with Pchlide in their closed tetrapyrrole structure (Figure 6A). We picked and compared five crystal structures binding to these four ligands (PMR, CHL, CLA and GB0), and found that most proteins that bind to these structurally similar porphyrin-containing ligands have different folds. The analysis further confirmed that these molecules are capable of binding to pockets with different geometries (Figure 6B), reorienting themselves in different positions within the active sites. Comparison of the binding sites in the five crystal structures showed that four atoms in the porphyrin ring had non-bonded interactions in all the five structures. However, we observed no conservation of the amino acids (Figure S3) involved in interactions with these four atoms (Table 4). Comparing the protein–ligand interactions from five different crystal structures, both 3AEK bound with Pchlide and 5L8R bound with CLA, were observed to have only one

hydrogen bonded interaction with O2A and OBD atoms of the ligand, respectively. The porphyrin binding pockets in all these protein structures are hydrophobic (Figure 6C).

To summarize, analysis of the overall ligand atom-amino acid interactions in the 14 structures show that frequencies of these non-bonded interactions in the five (A-E) rings of the CBB, CBC, CMD, CMA, and O2D ligands were 64, 78, 71, 64, and 57%, respectively, strongly suggesting that the interaction of these atoms of the porphyrin ring within the ligand binding pocket are important, strongly suggesting that the interaction of these atoms of the porphyrin ring within the ligand binding pocket are important. Overall, our analysis suggests that these ligands are capable of binding to proteins with diverse folds and active sites, through both the ligands and proteins undergoing adaptive conformational changes that enable them to accommodate their partners in binding sites with surprisingly different shapes. The findings suggest that we cannot easily predict the Pchl_{ide} binding site in pea POR, which does not share fold similarity with any of these 14 protein structures. Thus, it will be interesting to explore Pchl_{ide}'s mode of binding in the undetermined binding site of POR.

Short-chain dehydrogenase protein bound with substrate

To further elucidate the binding of the Pchl_{ide} substrate with POR, we searched the available X-ray crystal structures of short-chain dehydrogenases/reductases with structural homology to POR and identified the crystal structure of enoyl-ACP-reductase (INHA) protein from *Mycobacterium tuberculosis* (PDB_ID: 1BVR) in complex with NAD⁺ and a C16-fatty-acyl substrate (Rozwarski et al. 1999). Comparison of the crystal structure of INHA and the modelled structure of dimeric POR revealed very high structural similarity between the two proteins (RMSD of C α : 3Å). Comparing the secondary structures close to the substrate-binding region in INHA to that of the POR model, we observed a large insertion and significant structural differences. However, we mapped five amino acids (Pro94, Ser143, Gly229, Leu230

and Arg232) binding to the substrate in INHA to the POR protein based on the POR and INHA structural alignment. In the surroundings of this predicted binding site there are two large structurally flexible exposed loop regions (Gly142-Ala188 and Pro222-Glu256) between β 5- α 5 and β 6- α 7, which we hypothesize favor the adaptation of POR to its Pchlide binding partner and facilitate the binding process.

To explore the Pchlide binding mechanism in POR and corroborate the Pchlide binding site, we performed several PELE simulations in which the ligand was allowed to freely explore (and enter) the POR surface. PELE has proven ability to portray porphyrin diffusion, active site searching and binding (Kopečná et al. 2015). First, we performed a global search in which the substrate was allowed to explore the entire protein surface. As can be seen in Figure 7A, this procedure returned one main surface binding mode, from which we started a finer local search by exploring a space around up to 20 Å from the initial position (which the ligand's center of mass could not leave, Figure 7B). PELE clearly identified a binding pocket where two poses, differing only in the level of pocket penetration, were selected based on population analysis. The second most populated pose (designated A, Figure 7C), shows a slightly more solvent-exposed Pchlide than the most populated pose (B, which was also best in terms of POR-Pchlide interaction energy according to the PELE simulations), in which the substrate is more deeply bound (Figure 7D). According to earlier studies, the mechanism of Pchlide reduction involves several steps, including: light absorption, subsequent hydride transfer from the pro-S face of the nicotinamide ring of NADPH to C17 of the Pchlide molecule, and finally the transfer of a proton from Tyr280 to C18 of Pchlide (Wilks and Timko 1995, Heyes et al. 2015). To further clarify which of the two poses represents the substrate Pchlide binding pocket in POR most accurately, we compared the positioning of Pchlide and NADPH molecules in the POR complex. In the bound conformation of the Pchlide molecule in pose B (Figure 7D) (Figure 10D) the C17-C18 bond of Pchlide is close to the pro-S hydrogen from the nicotinamide ring

of the NADPH cofactor and Tyr191, which plays an important role in proton transfer to C18 of Pchlide in the catalytic mechanism. In contrast, in pose A the pro-S hydrogen of the NADPH cofactor is far from the C16-C17 bond of the Pchlide molecule. Thus, we predict that the deep docking position of Pchlide in the POR structure is closer to the real substrate binding pocket.

Conclusion

In this study, the structure of POR protein from pea was predicted in its monomeric and dimeric form. Alpha helices 4 and 7 form the interface between the two POR monomeric structures. Comparison of the NADPH binding site and its conformation within the active site of other NADPH binding proteins showed that it is well conserved, despite low sequence identity. Analysis of the binding sites of tetrapyrrole-containing molecules revealed high variability in the amino acids involved and active site conformation. In addition, exploration of the Pchlide binding mechanism in PELE simulations yielded two binding poses (designated A and B). In pose B the C17-C18 bond of Pchlide is close to the pro-S hydrogen from the nicotinamide ring of the NADPH and Tyr191, which is involved in proton transfer.

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Author's contributions

Conceived and designed the experiments: SH, HA. Performed the experiments: SH, VG. Analyzed the data: SH, VG, KS, HA. Wrote the paper: SH, VG, KS, HA. All authors read and approved the final manuscript.

Availability of data and materials

The data that support the findings of this study are available from the corresponding author upon reasonable request.

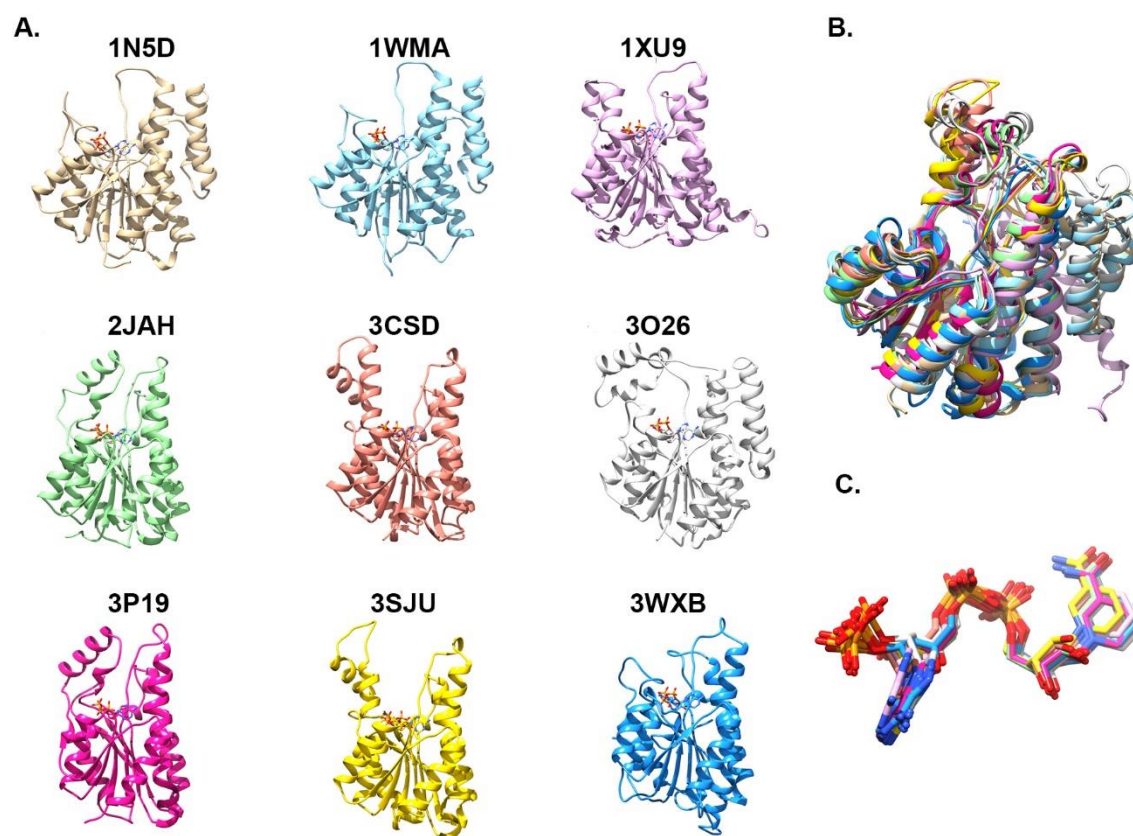


Figure 1. Structural comparison. (A) The nine selected homologous structures of POR that bind NADPH. (B) structural superimposition of all nine structures. (C) Conformations of the bound NADPH molecules in the nine structures.

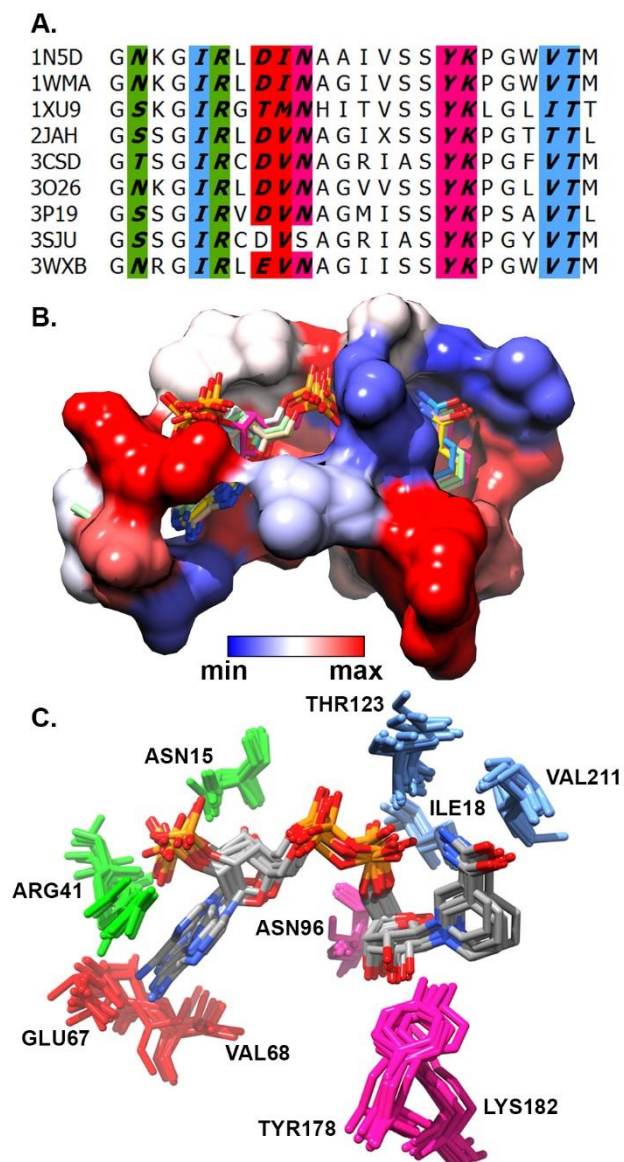


Figure 2. NADPH binding site analysis. (A) Sequence alignment of the nine selected crystal protein structures from PDB. Colored residues are conserved residues that form hydrogen bonds with the ligand. (B) Electrostatic representation of the NADPH binding cavities of the homologous structures. Red and blue colors indicate negatively and positively charged residues, respectively. (C) Binding of the conserved residues to the NADPH molecule. The residues are colored in accordance with the colors in the sequence alignment (A).

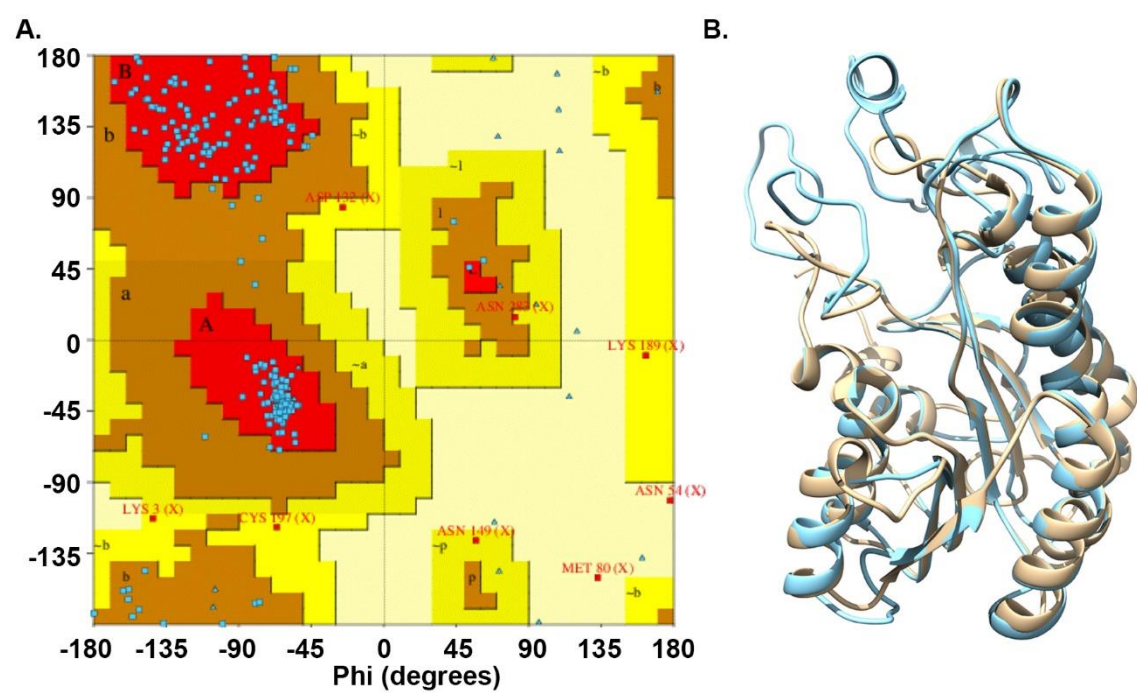
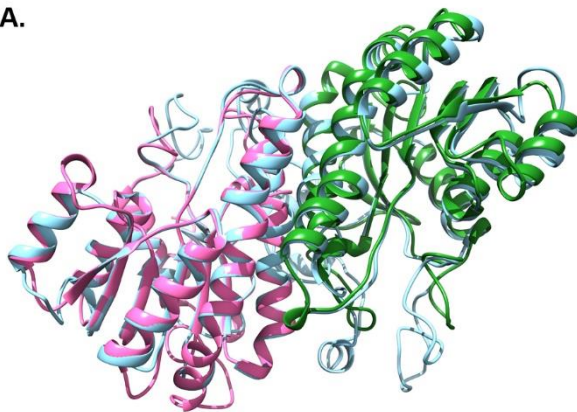


Figure 3. Modelled structure of POR. (A) Ramachandran plot of the modelled POR. (B) Structural superimposition of the modelled POR (in cyan) with the template structure, 3WXB (brown).

A.



B.

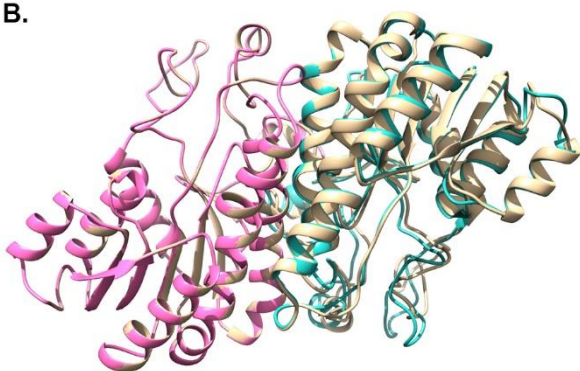


Figure 4. Dimer structure of POR. (A) The modelled structure of POR in dimer form (in cyan) superimposed on the homodimer structure of the template 3WXB (A and B chain in pink and green, respectively). (B) The MZDOCK predicted dimeric form (brown) of POR superimposed on the POR modelled dimer structure (A and B chains, pink and cyan, respectively).

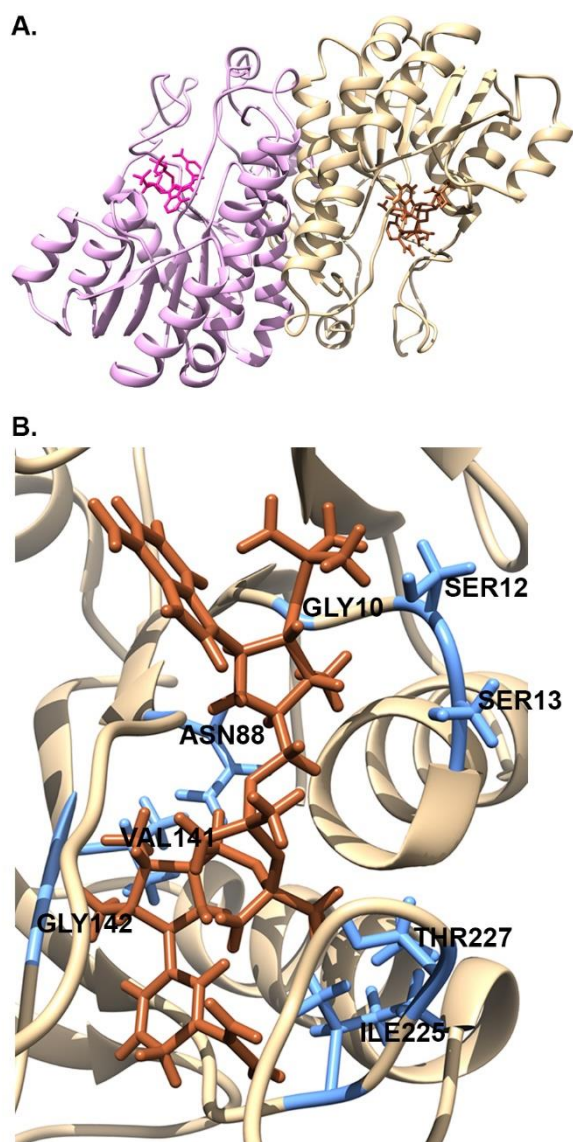


Figure 5. The binding pose of NADPH in the active site of each POR monomer of the POR homodimer obtained by molecular docking. (A) The A and B monomer in purple and light brown, respectively. The docked NADPH in the A and B monomer is shown in dark pink and dark brown, respectively. (B) The inset shows a magnification of the NADPH binding site.

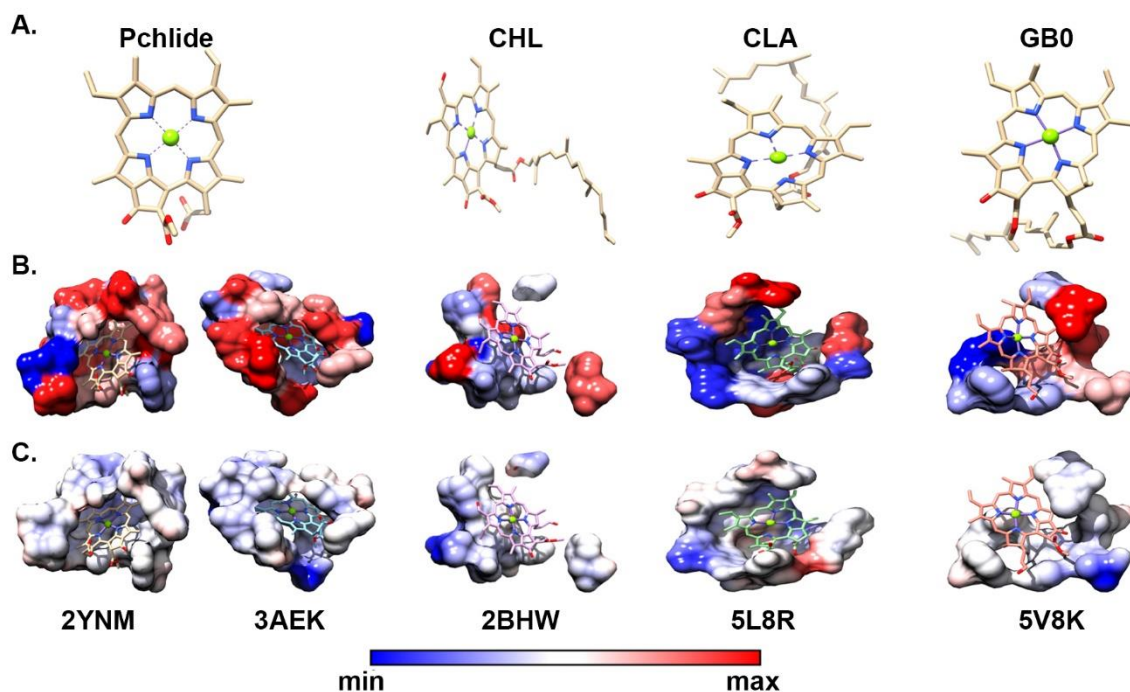


Figure 6. Ligand binding pockets for protochlorophyllide (Pchlde) and its structural analogues. (A) Structures of the four ligands — Pchlde, chlorophyll b (CHL), chlorophyll a (CLA) and bacteriochlorophyll g (GB0) — used for analysis. (B) Surface representation of the binding pocket in the representative structures (PDB_IDs shown at the bottom) colored according to each ligand's electrostatic potential. (C) Surface representation of the binding pocket colored according to hydrophobicity. 2YNM, nitrogenase-like dark-operative protochlorophyllide oxidoreductase complex from *Prochlorococcus marinus*; 3AEK, light-independent protochlorophyllide reductase from *Rhodobacter capsulatus*; 2BHW, chlorophyll a-b binding protein from pea; 5L8R, structure of plant photosystem I supercomplex from pea; 5V8K, structure of a symmetric photosynthetic reaction center-photosystem from *Heliobacterium modesticaldum*.

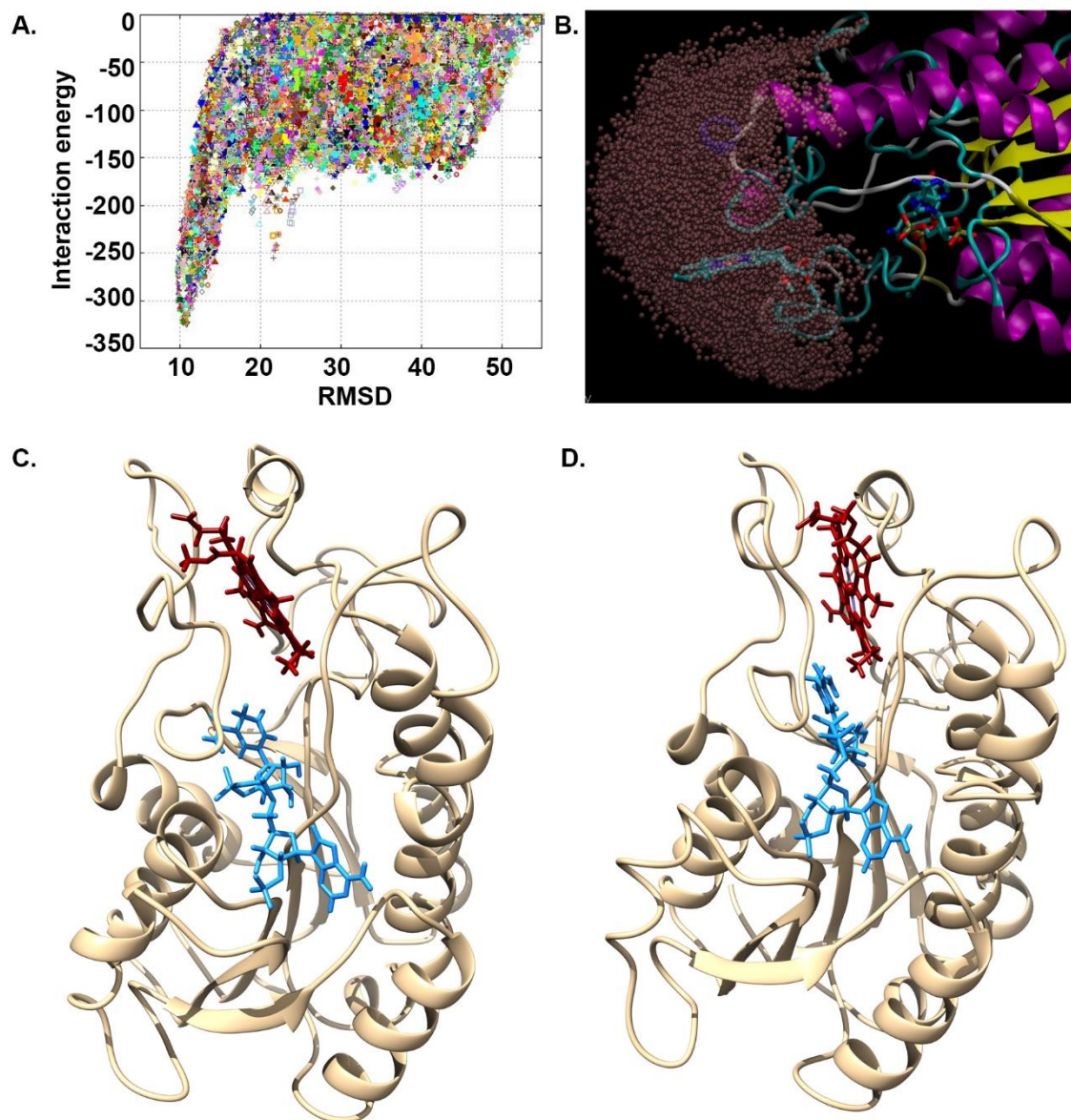


Figure 7. Snapshots of POR and Pchlride binding. (A) PELE's global search for Pchlride binding. The x axis shows the all heavy atom RMSD (in angstroms) to our preferred binding structure, pose B. (B) The exploration space around $\sim 20\text{\AA}$ from the initial position of Pchlride for PELE's local search. (C) Pose A bound structure. (D) Pose B bound structure, deeper in the POR binding pocket.

Table 1. Pairwise structural comparison of NADPH binding proteins. Each cell shows the percentage identity of the indicated pair, with RMSD in brackets.

PDB_ID	3WXB	2JAH	3P19	3CSD	3SJU	1XU9	3O26	1WMA	1N5D
3WXB		18.3 (1.9)	15.8 (2.0)	17.9 (2.1)	17.9 (2.2)	18.0 (2.0)	16.6 (2.4)	17.8 (2.4)	19.0 (2.3)
2JAH	18.3 (1.9)		38.2 (1.1)	28.8 (1.1)	29.5 (1.) (1.4)	25.2 (1.4)	16.4 (1.7)	20.5 (1.9)	19.7 (1.9)
3P19	15.8 (2.0)	38.2 (1.1)		20.4 (1.5)	20.1 (1.5)	19 (1.7) (1.9)	14.8 (1.9)	16 (2.0) (2.0)	15.9 (2.0)
3CSD	17.9 (2.1)	28.8 (1.1)	20.4 (1.5)		62.1 (0.5)	19.4 (1.6)	17.8 (1.8)	20.3 (1.9)	20.2 (1.9)
3SJU	17.9 (2.2)	29.5 (1.1)	20.1 (1.5)	62.1 (0.5)		17.6 (2.1)	18.7 (1.9)	19 (2.0) (2.0)	18.9 (2.0)
1XU9	18.0 (2.0)	25.2 (1.4)	19 (1.7) (1.6)	19.4 (1.6)	17.6 (1.6)		13.2 (2.1)	16.3 (2.1)	15.0 (2.1)
3O26	16.6 (2.4)	16.4 (1.7)	14.8 (1.9)	17.8 (1.8)	18.7 (1.9)	13.2 (2.1)		26.3 (1.6)	28.0 (1.6)
1WMA	17.8 (2.4)	20.5 (1.9)	16 (2.0) (1.9)	20.3 (1.9)	19 (2.0) (2.1)	16.3 (2.1)	26.3 (1.6)		76.0 (1.0)
1N5D	19.0 (2.3)	19.7 (1.9)	15.9 (2.0)	20.2 (1.9)	18.9 (2.0)	15.0 (2.1)	28.0 (1.6)	76.0 (1.0)	

Table 2. Critical amino acids for NADPH binding according to analysis of nine SDR-NADPH ligand complex structures deposited in the PDB database. The residue numbering (in parentheses) is based on 3WXB.

PDB_ID	Asn (15)	Arg (16)	Ile (18)	Arg (41)	Glu (67)	Val (68)	Asn (96)	Tyr (178)	Lys (182)	Val (211)	Thr (213)
3WXB	×	×	×	×	×	×	×	×	×	×	×
2JAH	×	×	×	×	×	×	×	×	×	×	×
3P19	×	×	×	×	×	×	×	×	×	×	×
3CSD	×	×	×	×	×	×	×	×	×	×	×
3SJU	×	×	×	×	-	×	-	×	×	×	×
1XU9	×	-	×	×	×	×	×	×	×	×	×
3O26	×	-	×	×	×	×	×	×	×	×	×
1WMA	×	-	×	×	×	×	×	×	×	×	×
1N5D	×	-	×	×	×	×	×	×	×	×	×

Table 3. Comparison of the protochlorophyllide (Pchl_{id}) binding sites of the PDB structures 3AEK and 2YNM. A.A., amino acids; ACC, hydrogen bond acceptor; ALI, aliphatic hydrophobic property; DAC, hydrogen bond donor and acceptor; DON, hydrogen bond donor; PII, aromatic property (pi contacts); b, amino acid backbone, buried; s, amino acid side chain, exposed; * identical amino acids between the two structures.

3AEK			2YNM			Property	
Chain.ID	A. A.	Type	Chain.ID	A. A.	Type	Source	Same AA
B.274	Asp	ACC	D.45	Met	ACC		
C.25	Phe	PII	C.16	Phe	PII	s	*
C.29	Thr	ALI	C.20	Thr	ALI	s	*
C.33	Trp	PII	C.24	Trp	PII	s	*
C.53	His	DAC	C.48	Ser	DAC		
C.54	Leu	ALI	C.45	Leu	ALI	s	*
C.57	Ala	ACC	C.48	Ser	ACC		
C.58	Ala	ACC	C.49	Ala	ACC	b	*
C.58	Ala	ALI	C.49	Ala	ALI	s	*
C.150	Phe	PII	C.141	Phe	PII	s	*
C.372	Leu	DON	C.363	Met	DON		
C.372	Leu	ALI	C.363	Met	ALI		
C.387	Trp	PII	C.378	Trp	PII	s	*
C.389	Ile	ALI	C.380	Ile	ALI	s	*

D.38	Tyr	PII	D.38	Tyr	PII	s	*
D.41	Leu	PII	D.41	Leu	PII	b	*
D.41	Leu	ALI	D.41	Leu	ALI	s	*
D.42	Leu	ALI	D.42	Leu	ALI	s	*
D.45	Met	ALI	D.45	Met	ALI	s	*
D.164	Leu	ALI	D.177	Leu	ALI	s	*
D.379	Val	ALI	D.395	Val	ALI	s	*

Table 4. Amino acid residues in the five crystal structures that interacted with the four atoms of the porphyrin-containing ligand molecules found to participate in non-bonding interactions.

	2BHW	2YNM	3AEK	5L8R	5V8K
CHB	Leu25A	Met45D His394D	Met45D	Glu1841 Tyr1801	Trp540A His537A
CMD	Tyr44A	Trp378C	Trp387C	Phe1651	Thr518A Leu605A
NB	Leu25A, Tyr24A	Leu41D Ser48C	Ala57C	Glu1841	His537A
NC	Tyr24A	Met363C	Ile389C Ala58C	Glu1841 AegG851	His537A

Table 5. Interacting atoms in rings A-E of Pchlride, chlorophyll b (CHL), chlorophyll a (CLA) and bacteriochlorophyll g (GB0) in the 15 studied crystal structures.

Sub- structure	PDB_ID	C1A	C2A	C3A	C4A	CAA	CBA	CGA	CMA	NA	O1A	O2A
Ring A	1JB0	✓	-	✓	✓	-	✓	✓	✓	✓	✓	✓
	1Q90	-	-	-	-	-	-	-	-	-	✓	-
	2BHW	✓	-	-	✓	✓	✓	-	-	✓	-	-
	2C9E	-	-	-	-	-	-	-	-	-	-	✓
	2DRE	-	-	✓	✓	-	✓	✓	✓	✓	✓	✓
	2YNM	-	✓	✓	-	✓	-	-	✓	-	-	-
	3AEK	-	-	-	-	-	✓	✓	✓	-	-	✓
	3IIS	-	-	-	-	-	-	-	✓	-	-	-
	4OGQ	-	-	-	-	✓	-	-	✓	-	✓	-
	4RI2	-	-	✓	-	-	-	-	✓	-	-	-
	5L8R	-	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓
	5V8K	-	-	✓	✓	-	✓	✓	✓	✓	✓	-
	PDB_ID	C1B	C2B	C3B	C4B	CAB	CBB	CMB	NB			

Ring B	1JB0	✓	-	-	-	-	-	-	✓			
	1PPR	-	-	-	-	-	✓	-	-			
	1Q90	-	✓	✓	-	✓	✓	✓	-			
	2BHW	✓	-	✓	✓	✓	✓	✓	✓			
	2C9E	-	-	-	-	-	✓	-	-			
	2DRE	✓	✓	✓	✓	✓	✓	✓	✓			
	2YNM	✓	✓	✓	✓	✓	✓	✓	✓			
	3AEK	-	✓	✓	✓	✓	✓	✓	✓			
	3IIS	-	-	-	✓	-	-	-	-			
	4OGQ	✓	✓	✓	-	✓	✓	✓	✓			
	4RI2	-	-	-	-	✓	✓	✓	-			
	5L8R	✓	✓	-	✓	✓	✓	✓	✓			
	5V8K	✓	-	-	-	-	-	-	✓			
	PDB_ID	C1C	C2C	C3C	C4C	CAC	CBC	NC				
Ring C	1JB0	-	✓	-	-	✓	✓	✓				
	1PPR	✓	✓	✓	✓	-	✓	✓				

	1Q90	✓	✓	✓	-	-	✓	-				
	2BHW	✓	✓	-	✓	-	✓	✓				
	2C9E	✓	✓	-	✓	-	✓	✓				
	2DRE	-	-	-	✓	✓	✓	✓				
	2X20	✓	✓	✓	✓	-	✓	✓				
	2YNM	✓	✓	-	-	✓	✓	✓				
	3AEK	✓	✓	✓	✓	✓	✓	✓				
	3IIS	✓	✓	✓	✓	-	✓	✓				
	4OGQ	✓	✓	✓	-	-	✓	-				
	4RI2	-	-	-	✓	✓	-	✓				
	5L8R	-	-	-	✓	✓	✓	✓				
	5V8K	-	-	-	-	✓	-	✓				
	PDB_ID	C1D	C2D	C3D	C4D	CMD	ND					
Ring D	1JB0	✓	✓	-	✓	✓	✓					
	1PPR	-	-	-	-	✓	-					
	2BHW	✓	-	-	✓	✓	✓					

	2C9E	✓	-	-	-	✓	-					
	2DRE	✓	✓	✓	✓	✓	✓					
	2X20	✓	-	-	-	✓	-					
	2YNM	-	✓	✓	-	✓	-					
	3AEK	✓	✓	✓	✓	✓	✓					
	3IIS	✓	-	-	-	✓	-					
	4OGQ	✓	✓	✓	✓	-	✓					
	4RI2	-	-	-	-	✓	-					
	5L8R	-	-	-	-	✓	✓					
	5V8K	-	-	-	✓	✓	✓					
	PDB_ID	C2O	C3D	C4D	CAD	CBD	CGD	CHA	O1D	O2D	OAD	
Ring E	1JB0	-	-	✓	✓	-	✓	✓	✓	✓	-	
	1Q90	-	-	-	-	-	-	-	✓	✓	-	
	2BHW	-	-	✓	-	✓	-	-	-	-	-	
	2DRE	-	✓	✓	✓	✓	✓	✓	✓	-	-	
	2YNM	✓	✓	-	✓	-	-	-	-	✓	✓	

	3AEK	✓	✓	✓	✓	✓	-	-	✓	✓	✓	
	4OGQ	-	✓	✓	-	-	-	-	✓	✓	-	
	5L8R	-	-	-	✓	✓	✓	-	✓	✓	-	
	5V8K	-	-	✓	✓	✓	-	-	-	✓	-	

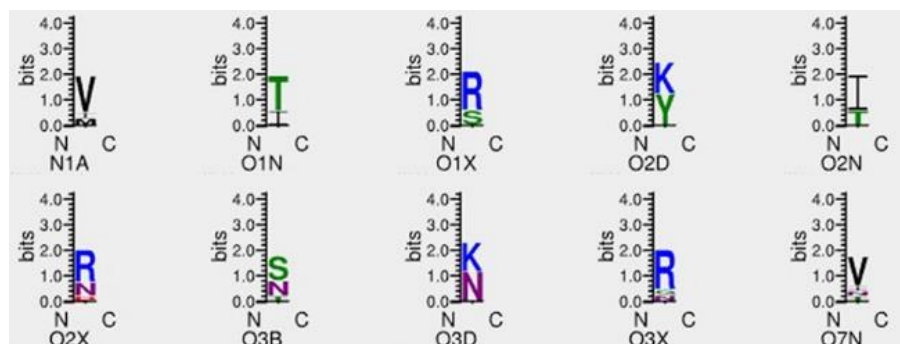


Figure S1. WebLogo representation of the residues identified as forming hydrogen bonds to atoms of the NADPH molecule.

3wxb β1 α1 β2 η1

1 10 20 30 40 50 TT 2222

3wxb S S G L V P R G S H M G E L R V R S V L V T C A N R C I G L G F V C H L L A L S N E . P E W V F A T C R P K G Q R A Q

2jah S A L Q G K V A L I T C A S S C I G E A T A R A L A E . G . A . A V A I A A R V E

3p19 M K K L V V I T C A S S C I G E A I A R F S E E . G . H . P L L L L A R V E

3csd S E V A I V T C A T S C L E I A R R L G R E . G . L . R V E V C A R E E

3sju Q T A F V T C V S S C I G L A V A T L A A R . G . T . A V Y G C A R A K

1xu9 Q P L N E E F R P E M L Q G K K V I V T C A S S C I G R E M A Y H L A K M . G . A . H V V V T A R K E

3o26 R R C A V V T C G N K C I G F E I C K Q L S S N . G . I . M V V L T C R V T

1wma S G I H V A L V T C G N K C I G L A I V R D L C R L F . S . G . G D V V L T A R V T

1n5d S S N T R V A L V T C A N K C I G F A I V R D L C R Q . . F A G . D V V L T A R V A

POR L R K G N V V I T C A S S C I G L A T A K A L A E S . G . K W H V I M A C R Y L

psPOR V V I T C A S S C I G L A T A K A L A E S . G . K W H V I M A C R Y L

atPORB V V V T C A S S C I G L A T A K A L A E T . G . K W N V I M A C R F L

dcPOR V I I T C A S S C I G L A T A K A L A E T . G . K W H V I M A C R F L

atPORA V V V T C A S S C I G L A T A K A L A E T . G . K W H V I M A C R F L

csPORA V V I T C A S S C I G L A T A K A L A E T . G . K W H V I M A C R F L

atPORC A V I T C A S S C I G L A T A K A L A D T . G . K W H V I M A C R F L

hvpORB A I I T C A S S C I G L A T A K A L A E S . G . K W H V I M A C R Y L

asPOR V V I T C A S S C I G L A A A K A L A E T . G . K W H V M A C R F L

hvpORA V V I T C A S S C I G L A A A K A L A E T . G . K W H V M A C R F L

taPORA V V I T C A S S C I G L A A A K A L A E T . G . K W H V M A C R F L

mpPOR C I I T C A S S C I G L A T A K A L A D T . G . E W H V I M A C R F L

crPOR A I I T C A S S C I G L A A A K A L A E T . G . E W H V M A C R F L

pbPOR V V I T C A S S C I G L Y A A K A L V K R . G . W H V M A C R L E

spOR V I I T C A S S C I G L Y A A K A L I D K . G . W H V I M A C R L D

3wxb α2 β3 α3 β4

60 70 80 90 100 110

3wxb E T O K L A S K H P N L V I V E L E V T . D F A S I K A A A S V C E R L K G S S I N L L I N N A G . A

2jah K I R A L G D E L T A A G A . K V H V L E L D V A . D F Q G V D A A V A S T V E A L G G L D I L V N N A G I . M

3p19 R I K A L N L P N T L C A Q V D V T . D K Y T F D T A I T R A E K I Y G P A D A I V N N A G M . M

3csd G I R T T I L K E L R E A G V . E A D G R T C D V R . S V P E I E A L V A A V E R Y G P V D V L V N N A G R . L

3sju N V S A A V D G L R A A G H . D V D G S S C D V T . S T D E V H A A V A A A V E R F G P I G I L V N S A G R . N

1xu9 T I O K V V S H C L E L G A A S A H Y I A G T M E . D M T F A E Q F V A A Q K L M G G L D M L I L I N H I N T

3o26 K G H E A V E K L K N S N H E N V V F H Q L D V T D P I A T M S S L A D F I T H F G K L D I L V N N A G V . A

1wma R Q Q A A V Q L Q A E G L S P R E H Q . L D I D . D L Q S I A L R D F L E E Y G G L D L V N N A G I . A

1n5d R Q Q A A V Q L Q A E G L S P F E H Q L D I I . D L Q S I A L R D F L E E Y G G L D L V N N A A I . A

POR K A A R A A K S A G I A K E . N Y T I M H L D I A . S L D S V R Q F V D N F F R S E M P L D V L I N N A A V . Y

psPOR K A A R A A K S A G I A K E . N Y T I M H L D I A . S L D S V R Q F V D N F F R S E M P L D V L I N N A A V . Y

atPORB K A E R A A K S V G M P K D . S Y T V M H L D I A . S L D S V R Q F V D N F F R T E T P L D V L V C N A A V . Y

dcPOR K A E R A A K S A G M P K E . N Y T I M H L D I A . S L D S V R Q F V E T F F R S E R P L D V L V C N A A V . Y

atPORA K A E R A A Q S A G M P K D . S Y T V M H L D I A . S L D S V R Q F V D N F F R A E M P L D V L V C N A A I . Y

csPORA K A E R A A K S A G I T K E . N Y T I M H L D I A . S L D S V R Q F V D N F F Q S G R P L D V L V C N A A V . Y

atPORC K A E K A A R S V G M S K E . D Y T V M H L D I A . S L E S V R Q F V E N F F R T E Q P L D V L V C N A A V . Y

hvpORB K T A R A A R A A G M P K G . S Y T I V H L D I A . S L D S V R Q F V E N V F Q L D M P I D V V V C N A A V . Y

asPOR K A S K A A K A A G M A D G . S Y T V M H L D I A . S L D S V R Q F V D A F F R A E M P L D V L V C N A A I . Y

hvpORA K A S K A A K A A G M A D G . S Y T V M H L D I A . S L D S V R Q F V D A F F R A E M P L D V L V C N A A I . Y

taPORA K A S K A A K A A G M A D G . S Y T V M H L D I A . S L D S V R Q F V D A F F R A E M P L D V L V C N A A I . Y

mpPOR K A E R A A R S V G I P K D . S Y T V I H C D I A . S F D S V R A F V D N F F R T E R Q L D V L V C N A A V . Y

crPOR K A E C A A K K V G M P A G . S Y S T I L H D I L S . S L E S V R Q F V Q N F R A S G R R L D A L V C N A A V . Y

pbPOR K A D S A A K S L G M S P D . S Y T L M H I D I G . S L D S V R K F V T Q F F E S G K S L D A L V C N A A V . Y

spOR K T O R V A D E L G E P K D . S Y T I L R L D I G . Y L D S V R F V M A Q F E L G R E L K A L V C N A A V . Y

3wxb α4

120 130

3wxb R A N T I D N E T L R D M S E V Y I T N T I A F L

2jah L L G P V E D A D T T D W T R M I D T N L L G L M

3p19 L L G Q I D T Q E A N E W Q R M F D V V L G L L

3csd G S G A T A E L A D E L W L D V E T N I T G V E

3sju S I G E T A D L D A L W A D V L D T N T G V F

1xu9 S I N L F . H D D I H V R K S M E V N E L S V V

3o26 G E S V D A D R F K A M I S D I G E D S E E L V K I Y E K P E A Q E L M S E T Y E L A E E C L K I N V N G V K

1wma F K V A D E T P F H I Q A E V T M K T N E F F G T R

1n5d F O L D N E T P F H I Q A E L T M K T N E M G T R

POR F F T A K E F S F T A D G F E I S V G T N H L G H F

psPOR F F T A K E F S F T A D G F E I S V G T N H L G H F

atPORB F F T A K E F T Y S A E G F E L S V A T N H L G H F

dcPOR F F T A K E F T A D G F E L S V G T N H L G H F

atPORA F F T A N Q T F T A E G F E L S V G T N H L G H F

csPORA F F T A K E F T F T A E G F E L S V G T N H L G H F

atPORC Q F T A K E F S F T A E G F E I S V G T N H L G H F

hvpORB Q F T A K E F S F T A D G F E M S V G V N H L G H F

asPOR R F T A R K E T F T A E G F E M S V G V N H L G H F

hvpORA R F T A R T E T F T A D G H E M S V G V N H L G H F

taPORA R F T A R T E T F T A D G H E M S V G V N H L G H F

mpPOR F F T D K E F K S A E G F E L S V G T N H M G H F

crPOR L E T A K E F T A D G F E L S V G T N H L G H F

pbPOR M F L L K E F M R F P S Y E L S V A T N H F G H F

spOR F F L L D E L W S A D O Y E L S V A T N H L G H F

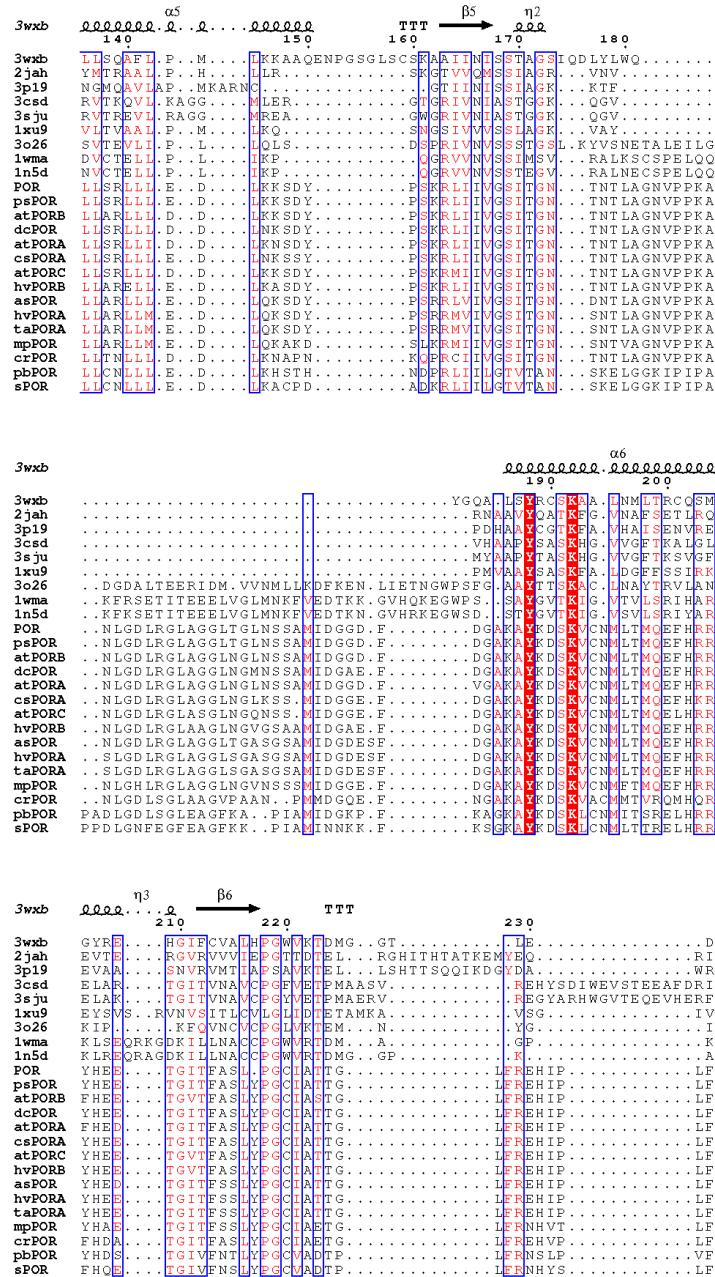


Figure S2. Multiple alignment of sequences, labelled to the left by organism name and gene name. The top line provides the secondary structure of 3WXB. Identical residues are red shaded. The blue boxes and red letters represent similar amino acids. Protein and accession numbers of the 14 UniProt sequences used are as follows: atPORA (Q42536), atPORB (P21218), atPORC (O48741), psPOR (Q01289), hvPORA (P13653), hvPORB (Q42850), mpPOR (P21218), crPOR (P21218), pbPOR (P21218), and sPOR (P21218).

taPORA (Q41578), asPOR (P15905), csPORA (Q41249), crPOR (Q39617), mpPOR (O80333), dcPOR (Q9SDT1), pbPOR (O66148), sPOR (Q59987). Species of origin is indicated with the following prefixes: at, *Arabidopsis thaliana*; ps, *Pisum sativum* (pea); hv, *Hordeum vulgare* (barley); ta, *Triticum aestivum* (wheat); as, *Avena sativa* (oat); cs, *Cucumis sativus* (cucumber); cr, *Chlamydomonas reinhardtii*; mp, *Marchantia paleacea* (liverwort); dc, *Daucus carota* (carrot); pb, *Plectonema boryanum*; s, *Synechocystis sp. strain PCC 6803*.

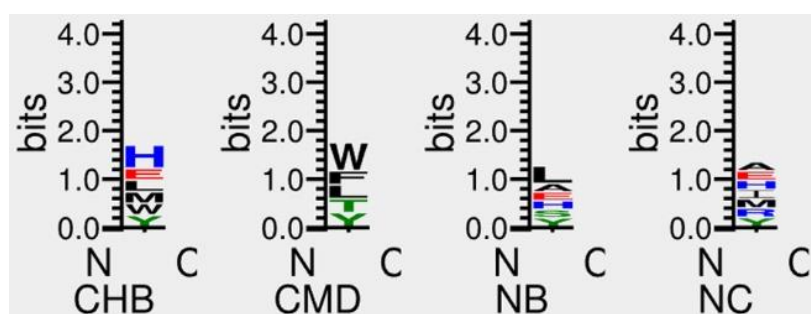


Figure S3. WebLogo of amino acids that interact with the four atoms of the porphyrin-containing ligand structures.

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