Mitochondrial DNA Poly-C length heteroplasmy as a marker for risk of critical COVID-19

Eliecer Coto¹, Daniel Vázquez-Coto¹, Guillermo M. Albaiceta¹, Laura Amado-Rodríguez¹, Mar González-Fernández¹, Claudia García-Lago¹, Lucinda Velázquez-Cuervo¹, Elías Cuesta-Llavona¹, and Juan Gómez¹

¹Hospital Universitario Central de Asturias

April 18, 2023

Abstract

Mitochondria play a central role in the innate and acquired response against viral infections. Common mtDNA variants have been associated with severe COVID-19 and mtDNA depletion. A poly C length variation has been associated with mtDNA instability and increased risk for several diseases. We studied 482 patients who required treatment in the intensive care unit and age matched population controls. The 16184-16193 poly-C and 514-523 CA-repeats were determined by fluorescent capillary electrophoresis and Sanger sequencing of PCR fragments. We found a significantly higher frequency of 16184-16193 mtDNA poly-C heteroplasmy in patients aged [?]60 compared to patients aged >60 years. Poly-C heteroplasmy did not differ between the age control groups. Poly-C heteroplasmy was associated with the presence of the 16223 T allele, that was associated with the risk of critical COVID-19 at [?]60 years. In Conclusion, heteroplasmy in the poly-C tract of the mtDNA control region might be a marker for critical COVID-19. The 16184-16193 heteroplasmy was linked to the 16223 T allele, that was significantly increased among patients aged [?]60 years. This finding requires validation in other cohorts and to determine the functional link between length variation in the mitochondrial DNA control sequence and risk of severe SARS-CoV-2 disease.

Mitochondrial DNA Poly-C length heteroplasmy as a marker for risk of critical COVID-19

Daniel Vázquez-Coto¹, Guillermo M. Albaiceta^{2,3,4,5,6}, Laura Amado-Rodríguez^{2,3,4,5,6}, Mar González-Fernández¹, Claudia García-Lago^{1,3}, Lucinda Velázquez-Cuervo^{1,3}, Elías Cuesta-Llavona^{1,3}, Juan Gómez^{1,3,5}, Elicer Coto^{1,3,4}

¹Genética Molecular, Hospital Universitario Central Asturias, Oviedo, Spain.

²Unidad de Cuidados Intensivos Cardiológicos, Hospital Universitario Central Asturias, Oviedo, Spain.

³Instituto de Investigación Sanitaria del Principado deAsturias, ISPA, Oviedo, Spain.

⁴Universidad de Oviedo, Oviedo, Spain.

⁵CIBER-Enfermedades Respiratorias. Instituto de Salud Carlos III. Madrid, Spain.

 6 Instituto Universitario de Oncología del Principado de Asturias. Oviedo, Spain.

 $^7 \mathrm{Neumologia},$ Hospital Universitario Central Asturias, Oviedo, Spain.

Corresponding author:

Eliecer Coto, PhD

Genética Molecular-HUCA, Oviedo, Spain

eliecer.coto@sespa.es

Abstract.

Mitochondria play a central role in the innate and acquired response against viral infections. Common mtDNA variants have been associated with severe COVID-19 and mtDNA depletion. A poly C length variation has been associated with mtDNA instability and increased risk for several diseases. We studied 482 patients who required treatment in the intensive care unit and age matched population controls. The 16184-16193 poly-C and 514-523 CA-repeats were determined by fluorescent capillary electrophoresis and Sanger sequencing of PCR fragments. We found a significantly higher frequency of 16184-16193 mtDNA poly-C heteroplasmy in patients aged [?]60 compared to patients aged >60 years. Poly-C heteroplasmy did not differ between the age control groups. Poly-C heteroplasmy was associated with the presence of the 16223 T allele, that was associated with the risk of critical COVID-19 at [?]60 years.

In Conclusion, heteroplasmy in the poly-C tract of the mtDNA control region might be a marker for critical COVID-19. The 16184-16193 heteroplasmy was linked to the 16223 T allele, that was significantly increased among patients aged [?]60 years. This finding requires validation in other cohorts and to determine the functional link between length variation in the mitochondrial DNA control sequence and risk of severe SARS-CoV-2 disease.

Key words: COVID-19; mitochondria; haplogroups; heteroplasmy; ageing

1. Introduction

Mitochondria contain their own genome, a circular double-strand DNA molecule (mtDNA) of 16,569-bp that encodes for proteins of the mitochondrial respiratory chain and mitochondrial tRNAs and rRNAs. Because mtDNA is inherited from the mother, germ-line mutations in the mtDNA are associated with rare maternal inherited diseases. The mtDNA is prone to acquire nucleotide changes that accumulate with age or under exposure to environmental toxics [Lee et al., 1998; Wallace 2010; Bratic and Larsson , 2013; Ziadaet al., 2019].

In addition to rare pathogenic variants the mtDNA contains many common variants that originated in individuals from particular populations and spread with worldwide migrations. Specific combinations of these variants classify the mitochondrial haplogroups, with frequencies that are characteristic of each human population [Wilson and Allard, 2004; Torroni et al., 2006; Brotherton et al., 2013]. For instance, haplogroup H is defined by 7028C (among other nucleotide changes) and is the most common among Europeans while is absent among individuals of African or East Asian ascent. These mtDNA haplogroups are transmitted from mother to offspring and their worlwide distribution permitted to trace the migration of humans outside Africa, raising the concept of a *mitochondrial Eve* [Pakendorf andStoneking, 2005].

Mitochondrial DNA variants/haplogroups have been associated with differences in physiological processes such as energy production, ageing, regulation of apoptosis or pathogen immune-mediated responses [Gomez-Duran et al., 2010; Chen et al., 2012;Kenney et al., 2014; Krzywanski et al., 2016; Friedrich et al., 2022]. As a consequence, these haplogroups have been associated with adaptation to exercise or susceptibility to develop several traits such as diabetes, cardiovascular disease, or infectious diseases [Castro et al., 2007; Yonova-Doing et al., 2022]. In reference to infections these variants might be associated with the risk of sepsis or the severity of HIV and herpex disease, among others [Hendrickson et al., 2008; Yang et al., 2008; Hart et al., 2013; Levinson et al, 2016]. Haplogroups might also play a role in the risk for severe COVID-19, the disease caused by SARS-CoV-2 [Wu et al., 2021; Dirican et al., 2022; Vazquez-Coto et al., 2022; Kumari et al. 2023].

Each cell has a variable number of mitochondria and each mitochondria contains several copies of the mtDNA. For a particular nucleotide position, the mitochondria from each individual may exhibit the same variant (homoplasmy) or different alleles (heteroplasmy) [Santos et al., 2008; Li et al., 2010;Klutsch et al., 2011]. Heteroplasmy is commonly inherited from the mother and for disease-related variants the degree of heteroplasmy in the different tissues determines the extent of the symptoms. Rare highly penetrant

mutations cause monogenic disorders that often affect the nervous system, muscles, heart, and endocrine organs, and many healthy individuals carry low levels of heteroplasmy (<1% of mtDNA with the mutation) either inherited from the mother or acquired. An increased burden of heteroplasmy contributes to increased risk for diseases such as MELAS (mitochondrial encephalomyopathy, lactic acidosis and stroke-like episodes), diabetes mellitus, and others [Avital et al., 2012; Folmes et al., 2013; Chae et al., 2020].

Heteroplasmy is common in poly-cytosine tracts in the mtDNA control region [Bendall et al., 1995; Lagerstrom-Fermer et al., 2001; Shin et al., 2006; Zhao et al., 2010;Mueller et al., 2011; Shen et al., 2015]. One of these is located between nucleotides 16184-16193, that contains the origin for replication of the mtDNA heavy (H) chain. These poly-C tracts are prone to length instability that would increase the risk of mtDNA loss and impairment of mitochondrial regulated processes [Chiaratti et al., 2022]. Among others, the T16189C mtDNA polymorphism increases the risk for poly-C instability and has been associated with diabetes, cancer, and coronary artery disease (CAD), among other diseases [Zhao et al., 2010; Mueller et al., 2011; Shen et al., 2015]. Length heteroplasmy might be at low level in resting while increases in cells subjected to extensive division, such as the immune cells under chronic inflammation. Increased heteroplasmy might thus represent a marker of a deleterious immune-response that would increase the risk of developing severe infectious diseases [Stefano et al., 2022; Ren et al., 2020; Elesela et al., 2021; Li et al., 2021].

Due to the overactivation of the immune system among individuals infected by SARS-CoV-2 we hypothesised that blood leukocytes from patients with severe COVID-19 might exhibit a different profile of poly-C heteroplasmy. To address this issue we characterised the mtDNA region containing the 16184-16193 poly-C tract in patients with critical COVID-19 and age-matched population controls.

2. Patients and Methods.

Study participants. We obtained the demographic and clinical data of 482 COVID-19 patients (age range 24-95 years) who required admission in the intensive care unit (ICU). The study was approved by the Ethics Committee of Principado de Asturias (Oviedo, Spain; approval id project ISCIII-PI21/00971), and all the patients (or their next of kin) gave informed consent to participate in the study. These patients were hospitalised between March-2020 and December-2021, a period with four SARS-CoV-2 pandemic waves in our community. We did not determine the SARS-CoV-2 variant in all the patients, but the study period was characterised by the Wuhan (pandemic waves 1-2), alpha (wave 3) and delta (wave 4) variants.

The inclusion criteria was a severe pneumonia in need for ICU admission with SARS-CoV-2 confirmed by PCR test, and exclusion criteria were age <18, respiratory failure due to condition other than COVID-19, or refusal to participate. These patients were followed till disease remission with hospital discharge or death. Body-mass index (BMI) and preexisting cardiovascular comorbidities (hypertension, diabetes, hypercholesterolemia) were obtained from the clinical history at ICU admission. Based on previous reports we compared COVID-19 patients aged [?]60 and >60 years [Nakanishi et al., 2021]. At the time of inclusion, none of the patients had a recorded diagnostic of mitochondrial disease.

All the participants were of European ancestry from the region of Asturias (Northern Spain, total population 1 million). The controls (N=363) were individuals from the general population recruited with the main purpose of defining the DNA variant frequencies. These controls were not serologically studied (presence of anti-SARS-CoV-2 antibodies) to exclude previous asymptomatic infection. In order to exclude the possibility of age-bias in the genotype frequencies we compared patients and controls within the same age-range.

Genetic analysis. Patients and controls were studied for mtDNA single nucleotide polymorphisms (SNPs) C7028T and C16223T by Sanger sequencing of PCR fragments. The 7028 polymorphism differentiates the major European haplogroup H (7028C) from no-H (7028T), while 16223 differentiates the ancestral African macro-haplogroup N (16223T) from the out-of-Africa macro-haplogroup R (16223C) (*www.mitomap.org*). This method allowed to determine the presence of heteroplasmy at the 7028 and 16223 nucleotides.

To define the degree of length heteroplasmy at the 16184-16193 poly C tract we per-

formed a fluorescent capillary electrophoresis of PCR fragments generated with primers FAM-5'CTGCCAGCCACCATGAATATTGTACGG and 5'GTGGCTTTG GAGTTGCAGTTGATGTGTGA (annealing at 65oC). The forward primer was labelled with FAM and fragments were thus visualised as fluorescent peaks after capillary electrophoresis (**Figure 1A**). Homoplasmic fragments corresponded to single peaks while the presence of additional peaks was considered as heteroplasmy. We also characterised the degree of heteroplasmy at the CA-repeat between nucleotides 514-523 by fluorescent capillary electrophoresis of PCR fragments with primers fam-5' CACTTTTAACAGTCACCCCCCAACTAAC and 5'TTCGGGGGTATGGGGTTA GCAGCG (annealing at 65oC) (**Figure 1B**). To discrimitate true from PCR stutter peaks (characteristic of amplicons containing nucleotide repeats) we condidered heteroplasmy when the additional peaks had a heigh >10% relative to the main fluorescent peak [**Walsh et al., 2016**]. The accuracy of this method was validated by sequencing PCR fragments with different degrees of heteroplasmy (**suppl figure**).

Statistical analysis. All the values were collected in an excell file. The statistical analysis was performed with the R-software (*www.r-project.org*). Logistic regression (linear generalized model, LGM) was used to compare mean values and frequencies between the groups.

3. Results.

In **table 1** we show the main characteristics of the critical COVID-19 patients aged [?]60 years or older. We found a higher frequency of male in the two age groups. As expected the frequency of hypertensives, diabetics, and hyperlipaemics was higher among the patients >60 years. There were a total of 103 deaths (**table 1**). The frequency of mtDNA 7028C that characterises haplogroup H was significantly lower in the [?]60 years patients (38% vs 48%). The frequency of 16223T was significantly higher in the [?]60 years group (18% vs 10%). The 7028C and 16223T did not differ between the two age control groups (**table 2**). Thus, we concluded that the difference between pagtients aged [?]60 and >60 years would not be a consequence of a survival-effect in the general population, and these mtDNA markers could thus represent risk factors for age-dependent critical COVID-19.

Control region length heteroplasmy. Fluorescent capillary electrophoresis of the amplified fragments showed multiple peaks (heteroplasmy) at the 16184-16193 poly-C tract significantly more frequent among the [?]60 years patients compared to age matched controls (19% vs 9%; p=0.002). Heteroplasmy was also more common among the [?]60 years compared to the older patients (19% vs 10%; p=0.02), without differences between patients and controls >60 years (table 2). We found a maximum frequency of length heteroplasmy among patients aged <50 years, suggesting that this mitochondrial genomic event was significantly associated with critical COVID-19 at younger age (Figure 2; suppl. Table 1).

The 16184-16193 poly-C tract that increases length heteroplasmy has been associated with several multifactorial disorders, including metabolic and cardiovascular. Because critical COVID-19 was significantly associated with hypertension, diabetes, dyslipidaemia, and male sex, we determined whether length heteroplasmy was higher among patients with these conditions. We did not find statistical differences between the groups (**suppl. table 2**). We also compared death and survivors, without significant difference. Moreover, after correcting by multiple variables advanced age was the only significantly associated with the risk of death (p<0.001).

We also determined the frequencies of the control region CAn-repeat. We did not find significant differences between patients and controls in the two age-groups. Heteroplasmy at this region was found in 3-6% of the study cohorts, without significant difference between the groups.

Poly-C heteroplasmy was linked to 16223 T. We found a higher frequency of poly-C heteroplasmy among patients 16223 T compared to 16223 C, in the two age groups (**suppl. table 3**). The 16223 T is the ancestral African allele and is present in less than 10% of current Europeans with the rare IXW haplogroups (**suppl. table 4**). This pointed to the possibility that the higher frequency of 16189 T and 162184-16193 poly-C heteroplasmy were a consequence of the linkage between 16223 T and 16189 C, a variant that increases the risk of poly-C instability (**suppl. table 5**).

4. Discussion.

The main finding of our study was a significantly increased frequency of length heteroplasmy at the poly-C tract in the mtDNA region that contains the replication start site of the H strand. This sequence, between nucleotides 16184-16193, contains a poly-C tract interrupted by C>T changes, and the presence of C could results in replication instability with multiple poly-C sequences.

The poly-C heteroplasmy might be inherited from the mother with cellular heterogeneity between different tissues in the same individual. In addition, the degree of heteroplasmy might increase with age or under exposure to environmental toxics, drugs, bacterial and viral infections, among others. An increased length heteroplasmy at this poly-C region has been reported among cancer, diabetes and coronary artery disease patients [Zhao et al., 2010; Mueller et al., 2011;Shen et al., 2015]. This associations might be explained by a mechanism that links the poly-C tract with an impaired replication of the mtDNA that could result in a reduction of the number of mtDNA copies and the loss of mitochondrial functions. In a study with blood samples from 837 healthy adults Liou et al. found that the number of mtDNA copies was significantly reduced among individuals with uninterrupted poly-C [Liou et al, 2010]. Thus, variants that increased the amount of length heteroplasmy might cause alteration of mtDNA copy number in human blood cells compared to T-interrupted tracts [Amo et al., 2017]. Interestingly, length heteroplasmy at the control region was also associated with significantly lower copy number of mtDNA in leukocytes from breast cancer patients [Zhao et al., 2010].

The number of mitochondria and mtDNA copies vary between cells and tissues and might be critical for a proper physiological response to situations that require a high energy demand and other mitochondrialmediated responses [Stefano et al., 2022;He et al., 2010; Stefano et al., 2017]. A high degree of heteroplasmy might impairs the maintenance of ATP levels in response to strong physiological demand, such as the required by the immunological cells after infections [Koshiba 2013;Moore and Ting, 2008; Schilf et al., 2021;Shenoy 2020]. The effect of heteroplasmy in mtDNA copy number and mitochondrial function has been validated in mouse strains with different mtDNA variants [Hu et al., 2019]. These engineered mice showed a non-random segregation of mtDNA copy numbers that suggested a pressure to reduce the degree of heteroplasmy when this was detrimental for covering the cellular demands. Situations that increase the degree of heteroplasmy, such as aging, exposure to environmental compounds, infections, etc, might result in an impaired capacity to respond through mitochondrial pathways, increasing the risk for disease.

Our study was based on whole blood cells and would thus be representative of the degree of heteroplasmy among leukocytes in a disease (COVID-19) characterised by an exacerbated inflammatory response with over-activation of immune-cells to respond to the infection. These cells are thus under strong metabolic demand that would require an increase in the number of mitochondria and mtDNA copies. It is tempting to speculate that a similar dynamic of mtDNA length heteroplasmy occurs in other cells and tissues such as lung, brain, heart. This possibility might be investigated in the context of persistent (long-COVID) symptoms.

The role of mitochondria in the pathophysiology of acute and chronic inflammation has been extensively studied. Immune-competent cells such as monocytes, macrophages, antigen-presenting cells, are under programmed changes in mitochondrial bioenergetics in response to innate and adaptive immunological processes [Angajala et al., 2018;Zuo et al., 2019]. The degree of mtDNA length heteroplasmy might contribute to define the extent of these immune-mediated responses, making some individuals more susceptible to an impaired capacity to fight infection or to regulate the extent of the proinflammatory stimuli [Lechuga-Vieco et al., 2020;Pollara et al., 2018; Zhu et al., 2018]. Several studies have shown the capacity of viruses, including SARS-CoV-2, to hijack oxidative phosphorylation, ATP production, and other mitochondrial functions [Stefano et al., 2022;Stefano et al., 2020]. Key mitochondrial mediated processes might be also affected by viral proteins that reduced the antiviral innate immune responses, thus promoting the extent of infection and disease severity [Chen et al., 2007; Yoshizumi et al., 2014; Choi et al., 2018]. Some authors have suggested that SARS-CoV-2 infection and replication was improved by the takeover of mitochondrial processes, and viral proteins would suppress mitochondrial functions reducing the innate and adaptive immune responses

[Singh et al., 2020]. In this context, heteroplasmy at the poly-C tract might reduce the individual's capacity to exhibit a proper mitochondria mediated immune response increasing the risk for severe COVID-19.

A functional link between 16189C and increased risk for disease has been suggested by some authors. Park et al. identified 16189C as a risk factor for diabetes and used chromatin immunoprecipitation in cybrid cells to identify the mitochondrial single-stranded DNA-binding protein (mtSSB) as a candidate protein bound to the 16189 region [**Park** et al., 2008]. MtSSB has a lower binding affinity for the 16189C variant and because this nucleotide lies in the control region of mtDNA replication and transcription the variant might affect mtDNA replication and recovery after mitochondrial damage [**Takamatsu** et al., 2002; **Park** et al., 2008]. In this context, several studies have reported increased circulating mtDNA as a marker of COVID-19 severity and mortality [**Scozzi** et al., 2021; **Valdes-Aguayo** et al., 2022; **Streng** et al., 2022]. Studies to determine whether mtDNA variants and leght heteroplasmy were associated with reduced number of mtDNA copies and increased circulating mtDNA would be of special interest.

A limitation of our study was the absence of over-time comparison of the degree of poly-C heteroplasmy within the same individual. Heteroplasmy is commonly inherited and could thus be present in all the individual's cells, but it is well known that de degree of heteroplasmy might increase with age or in response to environmental exposures. It is possible that SARS-CoV-2 proteins that target the mitochondria enhance the miss-replication of mtDNA promoting impairment of innate and acquired immunity and increasing the risk for critical COVID-19. To verify this hypothesis it is necessary to determine the degree of heteroplasmy before and after infection.

Finally, the risk of developing severe COVID-19 has been associated with some mitochondrial haplogroups [Wu et al., 2021;Vazquez-Coto et al., 2022]. We previously reported a decreased frequency of the common European haplogroup H (7028 C) that could be protective for critical disease at younger age. In addition, 16223 T might be associated with an increased risk of severe disease. Interestingly, 16189 C is more common among individuals with haplogroups with 16223 T (such as the European X) than among haplogroup H (characterised by 16223 C) (supplementary table) [Laricchia et al., 2022]. Thus, the association of haplogroups with several diseases might be explained by its linkage to 16189 C and other variants that could increase the risk for poly-C length heteroplasmy.

In conclusion, we report an increased frequency of poly-C length heteroplasmy at the mtDNA control region that contains the replication start site of the H-strand among patients with critical COVID-19. Length heteroplasmy at this region has been associated with a higher risk of cancer, diabetes, coronary artery disease, and viral diseases, among others. The poly-C heteroplasmy could increase mtDNA instability and reduction of the mtDNA copies per cell, that might contribute to manifest an exacerbated inflammatory response and impaired activation of the innate and acquired immunological response to viral infection. Further research to confirm the association are necessary, as well as functional studies to uncover the linkage between mtDNA length heteroplasmy and viral disease.

Acknowledgements. This work was supported by a grant from the Spanish Plan Nacional de I+D+I Ministerio de Economia y Competitividad and the European FEDER, grants ISCIII-PI21/00971 (E.C.), RICOR2040- RD21/0005/0011 (E.C.), and PI22/00705 (J.G.).

Competing interests. None of the authors have competing interests related to this work.

AUTHOR CONTRIBUTIONS . Lead researchers : EC, GMA, JG;study design : EC, GMA, JG; patient assessment and data acquisition : GMA, LAR, MGC, ECLL, JG; database: DVC, EC, GMA, JG; genotyping: DVC, CGL, MGF, LVC, EC; data filtering and analysis: EC, DVC; statistical analysis: EC, DVC;analysis of results: EC, DVC; writing the manuscript:EC; revision of manuscript: all the authors.

ETHICS AND CONSENT. This study was approved by the clinical research ethics committee of Hospital Universitario Central Asturias (HUCA) (project approval id ISCIII-PI21/00971). All the participants or they next of kin gave written or verbal consent. Data were handled in observance of Spanish legislation on data

protection. The study complies with the principles of the Declaration of Helsinki ("Recommendations guiding doctors in biomedical research involving human subjects").

DATA AVAILABILITY STATEMENT. The data that support the findings of this study are available from the corresponding author upon reasonable request. An Excel file with the raw data would be available for meta-analysis research.

	[?]60 years N=164	>60 years N=318	p-value
Male Female	118 (72%) 46 (28%)	235 (73%) 83 (27%)	0.85
Age Median years	53 (18-60)	71 (61-92)	
(range)			
BMI median (range)	29(19-54)	31(19-55)	0.01
BMI >30	69 (42%)	151 (47%)	0.01
Diabetes	21 (13%)	76 (24%)	0.005
Hypercholesterolemia	54 (33%)	162(51%)	0.0004
Hypertension	66 (40%)	203 (64%)	$<\!0.0001$
Death	17 (10%)	86 (28%)	$<\!0.0001$

 Table 1. Main values in the critical COVID-19 patients aged [?]60 years and older.

Table 2. Frequency of the mtDNA variants in the critical COVID-19 patients and population controls. All them were genotyped for the 7028 variant (C=haplogroup H) and the 16223 variant (T=haplogroup N; C=haplogroup R).

	[?]60 years	[?]60 years	>60 years	>60 years
	Covid-19	Controls	Covid-19	Controls
	N=164	N = 182	N=318	N=181
7028 C (H)	62 (38%)	86 (47%)	153~(48%)	82~(45%)
7028 T (no-H)	102(62%)	96(53%)	165(52%)	99(55%)
p-value (T) OR	p=0.11 OR=1.43	p=0.11 OR=1.43	p=0.19 OR=0.75	p=0.19 OR=0.75
(95%CI)	(0.92 - 2.22)	(0.92 - 2.22)	(0.49 - 1 - 15)	(0.49 - 1 - 15)
16223 T (N)	27~(16%)	9(5%)	31 (10%)	9~(5%)
16223 C (R)	133 (81%)	170 (93%)	283~(89%)	169 (93%)
$16223 \ T/C$	4(3%)	3(2%)	4 (1%)	3(2%)
Heteroplasmy				
p-value (T) OR	p=0.002 OR=3.36	p=0.002 OR=3.36	p=0.39 OR=1.47	p=0.39 OR=1.47
(95%CI)	(1.49-7.54)	(1.49-7.54)	(0.60 - 3.59)	(0.60 - 3.59)
514-523 CAn	21 (13%)	20 (11%)	32~(10%)	21 (12%)
CA_4				
CA_5	128~(79%)	140 (77%)	258~(81%)	145~(80%)
CA_6	6~(3%)	9~(5%)	12 (4%)	7~(4%)
CA ₇	0	2(1%)	4(1%)	4(2%)
$\mathrm{CA}_{\mathrm{heteroplasmy}}$	9~(5%)	11 (6%)	12 (4%)	4(3%)
16184-16193	32~(19%)	16 (9%)	31~(10%)	24~(13%)
Heteroplasmy				
Homoplasmy	132~(71%)	176~(91%)	287~(90%)	157 (87%)
	p=0.002 OR=2.67 (1.40-5.06)	p=0.002 OR=2.67 (1.40-5.06)	$\substack{\text{p=0.67 OR=0.89}\\(0.51\text{-}1.53)}$	p=0.67 OR=0.89 (0.51-1.53)

.

4. REFERENCES

Amo T, Kamimura N, Asano H, Asoh S, Ohta S. Cisplatin selects short forms of the mitochondrial DNA OriB variant (16184-16193 poly-cytosine tract), which confer resistance to cisplatin. Sci Rep. 2017 Apr 10;7:46240. doi: 10.1038/srep46240.PMID: 28393913

Angajala A, Lim S, Phillips JB, Kim JH, Yates C, You Z, Tan M. Diverse roles of mitochondria in immune responses: novel insights into immuno-metabolism. Front Immunol 2018; 9: 1605. https://doi.org/10.3389/fimmu.2018.01605**Avital** G, Buchshtav M, Zhidkov I, Tuval Feder J, Dadon S, Rubin E, Glass D, Spector TD, Mishmar D.**Mitochondrial** DNA **heteroplasmy** in **diabetes** and normal adults: role of acquired and inherited mutational patterns in twins. Hum Mol Genet. 2012 Oct 1;21(19):4214-24. doi: 10.1093/hmg/dds245. Epub 2012 Jun 26.PMID: 22736028

Bendall KE, Sykes BC. Length **heteroplasmy** in the first hypervariable segment of the human mtDNA control region. Am J Hum Genet. 1995 Aug;57(2):248-56.PMID: 7668250

Bratic A , Larsson NG. The role of mitochondria in aging. J Clin Invest. 2013 Mar;123(3): 951-7. doi: 10.1172/JCI64125. PMID: 23454757

Brotherton P, Haak W, Templeton J, Brandt G, Soubrier J, Jane Adler C, Richards SM, Der Sarkissian C, Ganslmeier R, Friederich S, Dresely V, van Oven M, Kenyon R, Van der Hoek MB, Korlach J, Luong K, Ho SYW, Quintana-Murci L, Behar DM, Meller H, Alt KW, Cooper A; Genographic Consortium. Neolithic mitochondrial haplogroup H genomes and the genetic origins of Europeans.Nat Commun. 2013;4: 1764. doi: 10.1038/ncomms2656. PMID: 23612305

Burtscher J, Cappellano G, Omori A, Koshiba T, Millet GP. Mitochondria: in the cross fire of SARS-CoV-2 and immunity. iScience 2020; https://doi.org/10.1016/j.isci.2020.101631

Castro MG, **Terrados** N , Reguero JR, Alvarez V, **Coto** E.Mitochondrial haplogroup T is negatively associated with the status of elite endurance athlete. Mitochondrion. 2007 Sep;7(5):354-7. doi: 10.1016/j.mito.2007.06.002. PMID: 17660050

Chae HW, Na JH, Kim HS, Lee YM. Mitochondrial diabetes and mitochondrial DNA mutation load in MELAS syndrome. Eur J Endocrinol. 2020 Nov;183(5):505-512. doi: 10.1530/EJE-20-0189.PMID: 33107434

Chen CY, Ping YH, Lee HC, et al. Open reading frame 8a of thehuman severe acute respiratory syndrome coronavirus not only promotesviral replication but also induces apoptosis. J Infect Dis 2007; 196: 405–415

Chen A, Raule N, Chomyn A, Attardi G. Decreased reactive oxygen species production in cells with mitochondrial haplogroups associated with longevity. PLoS One. 2012;7(10): e46473. doi: 10.1371/journal.pone.0046473. PMID: 23144696

Choi HJ, Park A, Kang S, Lee E, Lee TA, Ra EA, Lee J, Lee S, Park B. Human cytomegalovirus-encoded US9 targets MAVS and STING signaling to evade type I interferon immune responses. Nat Commun. 2018 Jan 9;9(1):125. doi: 10.1038/s41467-017-02624-8.PMID: 29317664

Chiaratti MR, Chinnery PF. Modulating mitochondrial DNA mutations: factors shaping heteroplasmy in the germ line and somatic cells. Pharmacol Res. 2022 Nov;185:106466. doi: 10.1016/j.phrs.2022.106466. Epub 2022 Sep 27.PMID: 36174964

Dirican E, Savrun ŞT, Aydın İE, Gülbay G, Karaman Ü. Analysis of mitochondrial DNA cytochromeb (CYB) and ATPase-6 gene mutations in COVID-19 patients. J Med Virol. 2022 Mar 8. doi: 10.1002/jmv.27704. Online ahead of print.PMID: 35258110

Elesela S, Lukacs NW. Role of Mitochondria in Viral Infections. Life (Basel). 2021 Mar 11;11(3):232. doi: 10.3390/life11030232.PMID: 33799853

Folmes CD, Martinez-Fernandez A, Perales-Clemente E, Li X, McDonald A, Oglesbee D, Hrstka SC, Perez-Terzic C, Terzic A, Nelson TJ. Disease-causing mitochondrial heteroplasmy segregated within induced pluripotent stem cell clones derived from a patient with MELAS. Stem Cells. 2013 Jul;31(7):1298-308. doi: 10.1002/stem.1389.PMID: 23553816

Friedrich VK, Rubel MA, Schurr TG.Mitochondrial genetic variation in human bioenergetics, adaptation, and adult disease.Am J Hum Biol. 2022 Feb;34(2): e23629. doi: 10.1002/ajhb.23629. PMID: 34146380

Gómez-Durán A, Pacheu-Grau D, López-Gallardo E, Díez-Sánchez C, Montoya J, López-Pérez MJ, Ruiz-Pesini E. Unmasking the causes of multifactorial disorders: OXPHOS differences between mitochondrial haplogroups.Hum Mol Genet. 2010 Sep 1;19(17): 3343-53. doi: 10.1093/hmg/ddq246. PMID: 20566709

Hart AB, Samuels DC, Hulgan T. The other genome: a systematic review of studies of mitochondrial DNA haplogroups and outcomes of HIV infection and antiretroviral therapy. AIDS Rev. 2013 Oct-Dec;15(4):213-20.PMID: 24322381

He Y, Wu J, Dressman DC, Iacobuzio-Donahue C, Markowitz SD, Velculescu VE, Diaz LA Jr, Kinzler KW, Vogelstein B, Papadopoulos N. Heteroplasmic mitochondrial DNA mutations in normal and tumour cells. Nature 2010; 464(7288):610–614. https://doi.org/10.1038/nature08802

Hendrickson SL, Hutcheson HB, Ruiz-Pesini E, et al. Mitochondrial DNA haplogroups influence AIDS progression. AIDS. 2008 Nov 30;22(18):2429-39. doi: 10.1097/QAD.0b013e32831940bb.PMID: 19005266

Hu M, Schulze KE, Ghildyal R, et al. Respiratory syncytial virus co-opts host mitochondrial function to favour infectious virus production.Elife. 2019 Jun 27;8:e42448. doi: 10.7554/eLife.42448.PMID: 31246170

Kenney MC, Chwa M, Atilano SR, Falatoonzadeh P, Ramirez C, Malik D, Tarek M, Del Carpio JC, Nesburn AB, Boyer DS, Kuppermann BD, Vawter MP, Jazwinski SM, Miceli MV, Wallace DC, Udar N. Molecular and bioenergetic differences between cells with African versus European inherited mitochondrial DNA haplogroups: implications for population susceptibility to diseases. Biochim Biophys Acta. 2014 Feb;1842(2): 208-19. doi: 10.1016/j.bbadis.2013.10.016. PMID: 24200652

Klütsch CF, Seppälä EH, Uhlén M, Lohi H, Savolainen P. Segregation of point mutation heteroplasmy in the control region of dog mtDNA studied systematically in deep generation pedigrees. Int J Legal Med. 2011 Jul;125(4):527-35. doi: 10.1007/s00414-010-0524-7. Epub 2010 Nov 4.PMID: 21049272

Koshiba T. Mitochondrial-mediated antiviral immunity. Biochim Biophys Acta. 2013 Jan;1833(1):225-32. doi: 10.1016/j.bbamcr.2012.03.005. Epub 2012 Mar 13.PMID: 22440325Krzywanski DM, Moellering DR, Westbrook DG, et al. Endothelial Cell Bioenergetics and Mitochondrial DNA Damage Differ in Humans Having African or West Eurasian Maternal Ancestry. Circ Cardiovasc Genet. 2016 Feb;9(1):26-36. doi: 10.1161/CIRCGENETICS.115.001308. PMID: 26787433

Kumari D, Singh Y, Singh S, Dogra V, Srivastava AK, Srivastava S, Garg I, Bargotya M, Hussain J, Ganju L, Varshney R. Mitochondrial pathogenic mutations and metabolic alterations associated with COVID-19 disease severity. J Med Virol. 2023; 95(2): e28553. doi: 10.1002/jmv.28553.PMID: 36832542

Lagerström-Fermér M, Olsson C, Forsgren L, Syvänen AC.Heteroplasmy of the human mtDNA control region remains constant during life. Am J Hum Genet. 2001 May;68(5):1299-301. doi: 10.1086/320115. Epub 2001 Mar 26.PMID: 11283795

Laricchia KM, Lake NJ, Watts NA, Shand M, Haessly A, Gauthier L, Benjamin D, Banks E, Soto J, Garimella K, Emery J; Genome Aggregation Database Consortium. Mitochondrial DNA variation across 56,434 individuals in gnomAD. Genome Res. 2022Mar;32(3):569-582. doi: 10.1101/gr.276013.121. PMID: 35074858

Lee HC, Lu CY, Fahn HJ, Wei YH. Aging- and smoking-associated alteration in the relative content of mitochondrial DNA in human lung.FEBS Lett. 1998 Dec 18;441(2): 292-6. doi: 10.1016/s0014-5793(98)01564-6. PMID: 9883902

Lechuga-Vieco AV, Latorre-Pellicer A, Johnston IG, Prota G, Gileadi U, Justo-Mendez R, Acin-Perez R, Martinez-de-Mena R, Fernandez-Toro JM, Jimenez-Blasco D, Mora A, Nicolas-Avila JA, Santiago DJ,

Priori SG, Bolanos JP, Sabio G, Criado LM, Ruiz-Cabello J, Cerundolo V, Jones NS, Enriquez JA (2020) Cellidentity and nucleo-mitochondrialgenetic context modulate OXPHOS performance and determine somaticheteroplasmydynamics. SciAdv 6(31): eaba5345. https://doi.org/10.1126/sciadv.aba5345

Levinson RT, Hulgan T, Kalams SA, Fessel JP, Samuels DC. Mitochondrial Haplogroups as a Risk Factor for Herpes Zoster. Open Forum Infect Dis. 2016 Oct 19;3(4):ofw184. doi: 10.1093/ofid/ofw184. eCollection 2016 Oct.PMID: 27807590

Li M, Schönberg A, Schaefer M, Schroeder R, Nasidze I, Stoneking M. Detecting heteroplasmy from high-throughput sequencing of complete human mitochondrial DNA genomes. Am J Hum Genet. 2010 Aug 13;87(2):237-49. doi: 10.1016/j.ajhg.2010.07.014.PMID: 20696290

Li X, Wu K, Zeng S, Zhao F, Fan J, Li Z, Yi L, Ding H, Zhao M, Fan S, Chen J. Viral Infection Modulates Mitochondrial Function. Int J Mol Sci. 2021 Apr 20;22(8):4260. doi: 10.3390/ijms22084260.PMID: 33923929

Liou CW, Lin TK, Chen JB, et al.. Association between a common mitochondrial DNA D-loop polycytosine variant and alteration of mitochondrial copy number in human peripheral blood cells. *J Med Genet* 2010; 47: 723-728

Moore CB, Ting JP. Regulation of mitochondrial antiviral signaling pathways. Immunity. 2008 Jun;28(6):735-9. doi: 10.1016/j.immuni.2008.05.005.PMID: 18549796

Mueller EE, Eder W, Ebner S, Schwaiger E, Santic D, Kreindl T, Stanger O, Paulweber B, Iglseder B, Oberkofler H, Maier R, Mayr JA, Krempler F, Weitgasser R, Patsch W, Sperl W, Kofler B. The **mitochondri**al T16189C polymorphism is associated with coronary artery disease in Middle European populations. PLoS One. 2011 Jan 26;6(1):e16455. doi: 10.1371/journal.pone.0016455.PMID: 21298061 Nakanishi T, Pigazzini S, Degenhardt F, et al. Age-dependent impact of the major common genetic risk factor for COVID-19 on severity and mortality. J Clin Invest. 2021 Dec 1;131(23):e152386. doi: 10.1172/JCI152386.PMID: 34597274

Pakendorf B, Stoneking M. Mitochondrial DNA and human evolution. Annu Rev Genomics Hum Genet. 2005; 6: 165-83. doi: 10.1146/annurev.genom.6.080604.162249. PMID: 16124858

 ${\bf Park}$, K.S., Chan, J.C., Chuang, LM. et al. A mitochondrial DNA variant at position 16189 is associated with type 2 diabetes mellitus in Asians. Diabetologia 51, 602–608 (2008). https://doi.org/10.1007/s00125-008-0933-z

Pollara J, Edwards RW, Lin L, Bendersky VA, Brennan TV. Circulating mitochondria in deceased organ donors are associated with immune activation and early allograft dysfunction. JCI Insight 2018; 3(15): e121622. https://doi.org/10.1172/jci.insight.121622

Ren Z, Zhang X, Ding T, Zhong Z, Hu H, Xu Z, Deng J. Mitochondrial Dynamics Imbalance: A Strategy for Promoting Viral Infection.Front Microbiol. 2020 Aug 21;11:1992. doi: 10.3389/fmicb.2020.01992. eCollection 2020.PMID: 32973718

Santos C, Sierra B, Alvarez L, Ramos A, Fernández E, Nogués R, Aluja MP. Frequency and pattern of heteroplasmy in the control region of human mitochondrial DNA. J Mol Evol. 2008 Aug;67(2):191-200. doi: 10.1007/s00239-008-9138-9. Epub 2008 Jul 11.PMID: 18618067

Scozzi D, Cano M, Ma L, Zhou D, Zhu JH, O'Halloran JA, Goss C, Rauseo AM, Liu Z, Sahu SK, Peritore V, Rocco M, Ricci A, Amodeo R, Aimati L, Ibrahim M, Hachem R, Kreisel D, Mudd PA, Kulkarni HS, Gelman AE. Circulating mitochondrial DNA is an early indicator of severe illness and mortality from COVID-19. JCI Insight. 2021 Feb 22;6(4):e143299. doi: 10.1172/jci.insight.143299.PMID: 33444289

Schilf P, Künstner A, Olbrich M, Waschina S, Fuchs B, Galuska CE, Braun A, Neuschütz K, Seutter M, Bieber K, Hellberg L, Sina C, Laskay T, Rupp J, Ludwig RJ, Zillikens D, Busch H, Sadik CD, Hirose M, Ibrahim SM. A mitochondrial polymorphism alters immune cell metabolism and protects mice from skin inflammation. Int J MolSci 2021; 22(3):1006. https://doi.org/10.3390/ijms22031006

Shen J, Wan J, Song R, Zhao H. Peripheral blood mitochondrial DNA copy number, length heteroplasmy and breast cancer risk: a replication study. Carcinogenesis. 2015 Nov;36(11):1307-13. doi: 10.1093/carcin/bgv130. Epub 2015 Sep 10.PMID: 26363030

Shenoy S. Coronavirus (Covid-19) sepsis: revisiting mitochondrial dysfunction in pathogenesis, aging, inflammation, and mortality. Inflamm Res 2020; 69(11):1077–1085**Shin** MG, Levin BC, Kim HJ, Kim HR, Lee IK, Cho D, Kee SJ, Shin JH, Suh SP, Ryang DW. Profiling of length **heteroplasmies** in the human **mitochondrial** DNA control regions from blood cells in the Korean population. Electrophoresis. 2006 Apr;27(7):1331-40. doi: 10.1002/elps.200500551.PMID: 16502464

Singh KK, Chaubey G, Chen JY, Suravajhala P (2020) Decoding SARS-CoV-2 hijacking of host mitochondria in COVID-19 pathogenesis. Am J Physiol Cell Physiol 319(2):C258–C267. htt-ps://doi.org/10.1152/ajpcell.00224.2020

Stefano GB, Bjenning C, Wang F, Wang N, Kream RM. Mitochondrial heteroplasmy. Adv Exp Med Biol 2017; 982:577–594. https://doi.org/10.1007/978-3-319-55330-6 30

Stefano GB, Kream RM. Mitochondrial DNA Heteroplasmy as an Informational Reservoir Dynamically Linked to Metabolic and Immunological Processes Associated with COVID-19 Neurological Disorders. Cell Mol Neurobiol. 2022 Jan;42(1):99-107. doi: 10.1007/s10571-021-01117-z. Epub 2021 Jun 12.PMID: 34117968

Streng LWJM, de Wijs CJ, Raat NJH, Specht PAC, Sneiders D, van der Kaaij M, Endeman H, Mik EG, Harms FA. In Vivo and Ex Vivo Mitochondrial Function in COVID-19 Patients on the Intensive Care Unit. Biomedicines. 2022 Jul 20;10(7):1746. doi: 10.3390/biomedicines10071746.PMID: 35885051

Takamatsu C, Umeda S, Ohsato T et al (2002) Regulation of mitochondrial D-loops by transcription factor A and single-stranded DNA-binding protein. EMBO Rep 3:451–456. PMCID: PMC1084112. DOI: 10.1093/embo-reports/kvf099

Torroni A, Achilli A, Macaulay V, Richards M, Bandelt HJ. Harvesting the fruit of the human mtDNA tree. Trends Genet. 2006 Jun;22(6): 339-45. doi: 10.1016/j.tig.2006.04.001. PMID: 16678300

Valdés-Aguayo JJ, Garza-Veloz I, Vargas-Rodríguez JR, Martinez-Vazquez MC, Avila-Carrasco L, Bernal-Silva S, González-Fuentes C, Comas-García A, Alvarado-Hernández DE, Centeno-Ramirez ASH, Rodriguez-Sánchez IP, Delgado-Enciso I, Martinez-Fierro ML. Peripheral Blood Mitochondrial DNA Levels Were Modulated by SARS-CoV-2 Infection Severity and Its Lessening Was Associated With Mortality Among Hospitalized Patients With COVID-19 .Front Cell Infect Microbiol. 2021 Dec 16;11:754708. doi: 10.3389/fcimb.2021.754708. eCollection 2021.PMID: 34976854

Vázquez-Coto D , Albaiceta GM, Amado-Rodríguez L, Clemente MG, Cuesta-Llavona E, Gómez J, Coto E. Common mitochondrial haplogroups as modifiers of the onset-age for critical COVID-19. Mitochondrion. 2022 Nov;67:1-5. doi: 10.1016/j.mito.2022.09.001. Epub 2022 Sep 15.PMID: 36115538

Wallace DC.Mitochondrial DNA mutations in disease and aging.Environ Mol Mutagen. 2010 Jun;51(5): 440-50. doi: 10.1002/em.20586. PMID: 20544884

Walsh PS, Fildes NJ, Sequence analysis and characterization of stutter products at the tetranucleotide repeat locus vWA. Reynolds R.Nucleic Acids Res. 1996 Jul 15;24(14):2807-12. doi: 10.1093/nar/24.14.2807.PMID: 8759015

Wilson MR, Allard MW. Phylogenetics and Mitochondrial DNA. Forensic Sci Rev. 2004 Jan;16(1): 37-62. PMID: 26256812

Wu Y, Wang XH, Li XH, et al. Common mtDNA variations at C5178a and A249d/T6392C/G10310A decrease the risk of severe COVID-19 in a Han Chinese population from Central China. Mil Med Res. 2021 Nov 1;8(1):57. doi: 10.1186/s40779-021-00351-2.PMID: 34724985

Yang Y, Shou Z, Zhang P, et al. Mitochondrial DNA haplogroup R predicts survival advantage in severe sepsis in the Han population. Genet Med. 2008 Mar;10(3):187-92. doi: 10.1097/GIM.0b013e318163c343.PMID: 18344708

Yonova-Doing E, Calabrese C, Gomez-Duran A, et al. An atlas of mitochondrial DNA genotype-phenotype associations in the UK Biobank. Nat Genet. 2021 Jul;53(7):982-993. doi: 10.1038/s41588-021-00868-1. PMID: 34002094

Yoshizumi T, Ichinohe T, Sasaki O, et al. Influenza A virus protein PB1-F2 translocates into mitochondria via Tom40 channels and impairs innate immunity. Nat Commun. 2014 Aug 20;5:4713. doi: 10.1038/ncomms5713.PMID: 25140902

Zhao H, Shen J, Medico L, Platek M, Ambrosone CB. Length heteroplasmies in human mitochondrial DNA control regions and breast cancer risk. Int J Mol Epidemiol Genet. 2010 Apr 5;1(3):184-92.PMID: 21537390

Zhu M, Barbas AS, Lin L, Scheuermann U, Bishawi M, Brennan TV (2018) Mitochondria released by apoptotic cell death initiate innate immune responses. Immunohorizons 2(11):384–397. htt-ps://doi.org/10.4049/immunohorizons.1800063

Ziada AS, Lu MY, Ignas-Menzies J, Paintsil E, Li M, Ogbuagu O, Saberi S, Hsieh AYY, Sattha B, Harrigan PR, Kalloger S, Côté HCF; CIHR team grant on cellular aging, HIV comorbidities in women, children (CARMA). Mitochondrial DNA somatic **mutation** burden and heteroplasmy are associated with chronological age, **smoking**, and HIV infection.Aging Cell. 2019 Dec;18(6): e13018. doi: 10.1111/acel.13018. PMID: 31407474

Zuo H, Wan Y (2019) Metabolic reprogramming in mitochondria of myeloid cells. Cells 9(1):5. https://doi.org/10.3390/cells9010005

Figure 1. Capillary electrophoresis of the 16184-16193 poly-C tract (A) and the 514 CAn repeat (B). Heteroplasmy (A2, A3, A4, B3): in addition to the main alleles all the fragments showed an additional peak known as DNA polymerase slippage product or stutter band. This peak would have a height <10% relative to the main peak (85). To confirm the accuracy of the capillary electrophoresis we confirmed the homoplasmy/heteroplasmy of PCR fragments by Sanger sequencing (see supplementary figures).

Figure 2. Frequencies of the poly-C lenght heteroplasmy and 16223 T patients in four age groups. Frequency of poly-C heteroplasmy and 16223 T were higher in the youngest (<50 years) group.



