# Warming increases survival and asexual fitness in a facultatively sexual freshwater cnidarian with winter diapause

Jácint Tökölyi<sup>1</sup>

<sup>1</sup>University of Debrecen

December 24, 2022

#### Abstract

Temperature is a key abiotic factor controlling population dynamics. In facultatively sexual animals inhabiting the temperate zone, temperature regulates the switch between asexual and sexual modes of reproduction, initiates growth or dormancy and acts together with photoperiod to mediate seasonal physiological transitions. Increasing temperature due to recent global warming are likely to disrupt population dynamics of facultatively sexual animals because of the strong temperature-dependence of multiple fitness components. However, the fitness consequences of warming in these animals are still poorly understood. This is unfortunate, since facultatively sexual animals – through their ability for asexual reproduction resulting in quick population growth and sexual reproduction enabling long-term persistence – are key components of freshwater ecosystems. Here, I studied the fitness effects of warming in Hydra oligactis, a freshwater cnidarian that reproduces asexually throughout most of the year but switches to sexual reproduction under decreasing temperatures. I exposed hydra polyps to simulated short summer heatwaves or long-term elevated winter temperatures. Since sexual development in this species is dependent on low temperature, I predicted reduced sexual investment (gonad production) and elevated asexual fitness (budding) in polyps exposed to higher temperatures. The results show a complex effect of warming on sexual fitness: while gonad number decreased in response to warming, polyps exposed to high winter temperature were capable of multiple rounds of gamete production. Asexual reproduction and survival rate, on the other hand, clearly increased in response to higher temperatures, especially in males. These results predict increased population growth of H. oligactis in temperate freshwater habitats, which will likely affect the population dynamics of its' main prey (freshwater zooplankton), and through that, the whole food web.

# Introduction

Organisms living in the temperate zone are exposed to seasonal fluctuations in temperature that provide contrasting physiological environments during summer and winter. Many species inhabiting such environments have adapted to temperature variation by specializing and evolving thermal preference to either high or low temperature (Huey and Kingsolver, 1989) or scheduling their major life events to different parts of the year (McNamara and Houston, 2008). For instance, among invertebrates that combine sexual with asexual reproduction in a complex life cycle (i.e., facultatively sexual organisms, such as aphids, flatworms, crustaceans, rotifers, sponges or cnidarians), sexual and asexual reproduction events are often performed in different parts of the year (Acker and Muscat, 1976; Fuchs et al., 2014; Green, 1966; Reisa, 1973; Shaffer et al., 2020; Simon et al., 2002). Increasing or decreasing temperatures in these species can induce a switch from asexual to sexual reproduction (Acker and Muscat, 1976; Fuchs et al., 2014; Reisa, 1973; Schröder, 2005; Shaffer et al., 2020; Simon et al., 2002; Vowinckel, 1970), initiate the growth and development of embryos inside resting eggs (Cáceres and Schwalbach, 2001; Cooley, 1971; Gilbert, 2017; Gulbrandsen and Johnsen, 1990; Hairston and Kearns, 1995; Vandekerkhove et al., 2005), and act as a modulator of other cues, such as photoperiod and crowding (Decaestecker et al., 2009; Gyllström and Hansson, 2004; Innes, 1997; Reisa, 1973; Schröder, 2005).

Temperature is a key regulator of the dynamics of populations because it determines locomotor activity and

food intake rates (Angilletta et al., 2002) and acts as a physical factor regulating cellular processes, such as metabolism or energy budgets (Brown et al., 2004; Huey and Kingsolver, 2019), ultimately determining growth, reproduction and survival (Angilletta, 2009). In facultatively sexual organisms both sexual and asexual modes of reproduction critically depend on temperature and switches in the mode of reproduction are expected to have a major influence of population dynamics. Both modes of reproduction contribute to population persistence and/or growth, albeit in different ways. Asexual reproduction results in quick population growth, allowing genotypes to increase in frequency when conditions are favourable (Scheuerl and Stelzer, 2019). Sexual reproduction, by contrast, often results in the production of resting eggs that are able to survive adverse conditions. These resting eggs are able to persist for years or even decades and replenish populations after stochastic extinction events (Franch-Gras et al., 2017), provide sources of novel genetic combinations with higher fitness (McLean et al., 2022), and enable dispersal of propagules to novel habitats (Panov et al., 2004).

Recent climate change, however, is affecting temperatures worldwide and rapidly rising temperatures have negative effects on organisms that evolved under a different thermal regime (McCarty, 2001; Walther et al., 2002). Documented effects of global warming include changes in phenology (i.e. timing of major life history events such as migration in birds or flowering in plants; (Jenni and Kéry, 2003; Molnár V et al., 2012)), shifts in geographical range size (Thomas, 2010) and a widely detected decline in animal body size (Sheridan and Bickford, 2011). Facultatively sexual organisms, due to their adaptation to seasonally varying temperature regimes, should be strongly impacted by warming temperatures, although the complexity of facultatively sexual life cycles makes prediction of the expected consequences of global warming difficult. In aphids, for instance, documented population responses to warming can be either positive, neutral or negative (Blanchard et al., 2019; van Baaren et al., 2010). Since aphids are reproducing parthenogenetically during summer and switch to sexual reproduction during summer, rising temperatures are expected to yield short-term increases in population sizes (until temperature becomes stressful) and the disappearance of sexually produced offspring (Blanchard et al., 2019). Warming temperatures have been shown in mesocosm experiments to boost spring population growth of zooplankton, with a higher effect in parthenogenetically reproducing cladocerans than non-parthenogenetic copepods (Ekvall and Hansson, 2012). Due to the altered population growth and differential thermal sensitivity, zooplankton communities experience changes in dominance patterns (Ekvall and Hansson, 2012), but also experience trophic mismatches that can result in population declines (Winder and Schindler, 2004). However, despite these examples, the number of studies examining the expected consequence of temperature warming on facultatively sexual organisms is still very low. This is unfortunate, since facultatively sexual organisms – through their ability to achieve quick population growth through asexually reproduction and long-term persistance and dispersal through the production of dormant stages - are key components of the ecosystems they inhabit, both as prey and as predators. Therefore, changes in their population dynamics due to climate warming is likely to have wide-range consequences at the level of the whole ecosystem.

The freshwater cnidarian Hydra oligactis Pallas 1766 is a small, sessile predator inhabiting the temperate zone. It is a cold-adapted species that prefers low temperatures, and even short-term exposures to temperatures above 30 °C are lethal (Bosch et al., 1988). They reproduce asexually throughout much of the year through budding, however, they rapidly switch to a sexual mode of reproduction upon exposure to cold temperatures (Reisa, 1973). Males produce testes, and females produce ovaries, and upon fertilization the egg develops a into a resting embryo surrounded by a thick shell that is extremely resistant to adverse conditions (Reisa, 1973). Hence, the production of resting eggs appears to be an adaptation to survive freezing water through dormancy, and indeed, sexually reproducing individuals are found in nature during autumn, before the freezing of water surface occurs (Miklós et al., 2021; Sebestyén et al., 2018). Following sexual reproduction, hydra polyps experience a senescence-like process and increased mortality risk, although some of them survive and revert to asexual reproduction (Tökölyi et al., 2017; Yoshida et al., 2006), depending on their age, size, and genotype (Miklós et al., 2022; Ngo et al., 2021; Sebestyén et al., 2020). Because of the presence of these asexual individuals, *H. oligactis* can reach huge population densities during late winter and early summer ((Bryden, 1952); J.T. personal observation) which could influence the population dynamics of their prey (mainly cladocerans and copepods) in a manner that amplifies with climate change. However, whether warming temperatures affect sexual or asexual fitness, and thereby population dynamics in hydra is still unclear.

Here, I set out to address this question by exposing *H. oligactis* polyps belonging to six strains (three male and three female) to elevated temperature. Since climate warming is predicted to be unequal across seasons (Meehl et al., 2007), I considered two scenarios: an increased frequency of summer heatwaves and an increased average winter temperature, both of these predicted based on current climate projections for the temperate and boreal zone (Meehl et al., 2007). To this end, hydra polyps were exposed either to a short (1-week long) period of high summer temperature (23  $^{\circ}$ C, compared to 18  $^{\circ}$ C for controls), which was followed by a winter treatment consisting either of low (8  $^{\circ}$ C) or high winter temperature (12  $^{\circ}$ C) maintained for five months. Throughout the experiment sexual and asexual fitness components were recorded. Because of the sensitivity of *H. oligactis* to high temperature I predicted reduced performance (both sexual and asexual) of polyps exposed to short heatwaves. Furthermore, because of the dependence of sexual reproduction on cold temperatures in *H. oligactis* I predicted a reduced sexual fitness in groups exposed to warm winter temperatures. Conversely, a higher asexual fitness and survival might be predicted due to the negative relationship between sexual reproduction and other fitness components (Ngo et al., 2021; Roff, 1993; Stearns, 1989).

#### Materials & methods

## Strains and culture conditions

For the experiment presented here six *H. oligactis* strains were used (three male strains: C2/7, M83/4 and T3/2 and three female strains: X11/14, M26/9/10 and T3/1). All strains were established from a single individual that was collected from freshwater bodies in Eastern Hungary between 2016 and 2020 and asexually propagated in the lab under simulated summer conditions (Gergely & Tökölyi *in press*). These conditions include 18  $^{\circ}$ C temperature and 12/12 hours light/dark cycle. Polyps were kept individually in 6-well tissue culture plates containing artificial hydra medium (1mM Tris, 1mM NaCl, 1mM CaCl<sub>2</sub>, 0.1mM KCl, 0.1mM MgSO<sub>4</sub>; pH: 7.6) and they were fed and cleaned four times per week.

# Experimental design

All data recorded for this experiment comes from polyps that were kept individually. The experiment consisted of two treatments: a summer treatment and a winter treatment. The summer treatment lasted for one week and aimed at simulating a heat wave. The winter treatment lasted for five months and aimed at simulating elevated winter temperatures. For both treatments, the control values were chosen such that both summer and winter conditions are comparable to our previous studies investigating H. oligactis life history (Ngo et al. 2021; Sebestvén et al. 2020; Tökölyi et al. 2017). Hence, the control temperature for summer was set as 18 °C (cold summer, CS) and the control temperature for winter was set to 8 °C (cold winter, CW). To simulate summer heatwaves, I exposed polyps to 23 °C for one week (warm summer, WS). Water temperatures as high as 25 °C regularly occur in natural populations at the end of the summer (J. T. personal observation), however, in the laboratory larger or longer heat treatments (i.e., above 23  $^{\circ}$ C or longer than one week) result in excess mortality. For the winter treatment, I exposed experimental polyps to 12 °C temperature for five months (warm winter, WW). A four degree increase in winter temperature is in accordance with IPCC predictions for Europe by the end of the century (Meehl et al., 2007). The summer and winter treatments were applied in a factorial design, such that there were four experimental groups: cold summer – cold winter (CS-CW); cold summer – warm winter (CS-WW); warm summer – cold winter (WS-CW) and warm summer – warm winter (WS-WW).

During the one-week summer treatment (either 18 or 23  $^{\text{o}}$ C) polyps were fed four times per week. After the summer treatment, polyps were photographed on a 1-mm grid paper sheet to measure their size, since size at cooling is an important predictor of sexual reproduction and post-reproductive survival (Ngo et al., 2021). Next, they were moved to either 8 or 12  $^{\text{o}}$ C, depending on their treatment and an 8/16 hours light/dark cycle. Lowering temperature induces sexual reproduction in this species and I recorded the presence of male or female gonads four times per week as a proxy for sexual fitness. In addition, the number of detached

buds was recorded twice per week as a proxy for asexual fitness. The winter treatment lasted for five months during which polyps were fed and cleaned twice per week. A five month winter treatment is necessary and sufficient time period to quantify sexual reproduction and post-reproductive survival in most polyps in these strains. At the end of the winter treatment, polyps were scored as survived if they had an intact body with tentacles and had the ability to feed, and they were scored dead if they disappeared during the experiment or consisted only of necrotic tissue. All experimental groups were started with 30-36 polyps per strain, with a total starting sample size of N = 843, although some individuals were excluded because they were accidentally lost during the experiment, or they proved to be sex-changed individuals (see Results). The experiment was initiated in batches over a period of about two months, depending on the availability of experimental animals. Each batch consisted of animals randomly divided into four treatment groups and any batch effects were statistically taken into account (see below).

## Data analysis

The effects of treatments were analyzed for the following dependent variables: (1) body size after summer treatment; (2) sexual development time (days elapsed after the beginning of the winter treatment until the appearance of first gonads); (3) sexual fitness (total number of eggs produced by females and the maximum number of testes seen on a male); (4) asexual fitness (total number of buds produced by an individual from the start to the end of the experiment, i.e. during both the summer and winter treatments) and (5) survival rate. I used Generalized Linear Mixed Models implemented in the glmmTMB package (v1.1.4) in R version 4.2.2 (Brooks et al., 2017; R Core Team, 2020). Gaussian errors were assumed for body size and binomial errors for survival rate. For sexual development time, sexual and asexual fitness (count data), four different error structures were considered: Gaussian, Poisson, negative binomial with linear parametrization (nbinom1) and negative binomial with quadratic parametrization (nbinom2) (Brooks et al., 2017). The model best fitting to the data was selected with Akaike Information Criteria corrected for small sample size (AICc). After selecting the right distribution, I fitted a model with experimental treatment as a fixed effect and strain ID and batch ID as random effects. This model was compared to a random effects null model via Likelihood Ratio Tests (LRTs). Post-hoc treatments-versus-control comparisons with Dunnett's correction were obtained using the *emmeans* R package (Lenth, 2022). For the model analyzing body size after summer treatment only one predictor was included (summer treatment) and I used LRT to evaluate the effect of treatment on size. Finally, I generated model diagnostics plot using the DHARMa R package (v0.4.5; (Hartig, 2022)) to check that model residuals did not show outstanding deviations from model assumptions. Since the model diagnostics for sexual development time showed substantial deviations from the expectation for both males and females, I also analyzed these variables with non-parametric Kruskal-Wallis tests, followed by pairwise Wilcoxon tests with Holm's correction.

## Results

Of the total N = 843 individuals 12 were accidentally lost during routine handling and nine individuals were sex-changed. All data were removed for these individuals, yielding a sample size of N = 822, with N = 206 individuals in the CS-CW group, N = 200 individuals in the WS-CW group, N = 210 individuals in the CS-WW group and N = 206 individuals in the WS-WW group.

# Body size after summer treatment

A one-week heat treatment reduced body size in male polyps by approximately 17% and female polyps by approximately 24%, with the difference being statistically highly significant in both sexes (Gaussian GLMM, LRT males: chi-square = 117.59, p < 0.001; females: chi-square = 42.984, p < 0.001; Fig. 1.).

#### Sexual development time

In both male and female strains, sexual development time was significantly influenced by treatment (males: LRT, Gaussian GLMM,  $\lambda^2 = 257.19$ , p < 0.001; females: LRT, Negative binomial GLMM with quadratic parametrization,  $\lambda^2 = 133.34$ , p < 0.001). Both male and female polyps exposed to one week heat treatment showed delayed sexual development, while those exposed to elevated winter temperatures showed slightly,

but significantly faster sexual development (Table 1., Fig. 2., 3.). Polyps in the WS-CW group delayed their reproduction most (Fig. 2A., 3A.), implying that a large sudden drop in temperature has negative effects on sexual development. The results were corroborated by the Kruskal-Wallis test and the pairwise Wilcoxon test as *post-hoc*analysis, with the exception of males in the CS-WW group, where advancement of sexual development time was only marginally significant (p = 0.081).

#### Sexual fitness

The number of gonads produced was also significantly influenced by treatment (males: LRT, Gaussian GLMM,  $\lambda^2 = 56.43$ , p < 0.001; females: LRT, Negative binomial GLMM with linear parametrization,  $\lambda^2 = 34.66$ , p < 0.001). Male polyps exposed to a simulated summer heatwave had reduced number of testes (Table 1.; Fig. 2B.). The negative effects of a warm summer were accentuated if they were followed by cold winter: a sudden large drop in temperature was associated with markedly reduced male fitness (Table 1.; Fig. 2B.). In females, on the other hand, only the WS-CW group had significantly reduced number of eggs compared to the CS-CW group (Table 1., Fig. 3B.).

Surprisingly, polyps exposed to a warm winter treatment underwent a second round of gametogenesis after the first one. This second gametogenesis event happened in individuals that recovered from their first gametogenesis event, around 3-4 months after cooling (Fig. 2C., 3C.).

## Asexual fitness

In males, as exual fitness was significantly affected by treatment (LRT, Negative binomial GLMM with linear parametrization,  $\lambda^2 = 26.96$ , p < 0.001). The number of buds produced increased if polyps were exposed to a simulated summer heat wave and if they were cultured under elevated winter temperatures, or they received both treatments compared to a cold summer-cold winter scenario (Table 1, Fig. 4A.). In females, by contrast, treatment had no effect on as exual fitness (LRT, Negative binomial GLMM with quadratic parametrization,  $\lambda^2 = 6.11$ , p = 0.106).

# Survival rate

Male survival rate was significantly affect by treatment (LRT, Binomial GLMM,  $\lambda^2 = 44.36$ , p < 0.001). Male polyps exposed to simulated summer heatwaves, elevated winter temperatures, or both treatments had significantly higher survival than polyps in the control group (Table 1, Fig. 4C.). The highest survival rate was seen in male polyps exposed to the CS-WW treatment, where about 80% of the polyps survived compared to ~50% in the control CS-CW group. Female survival was only marginally significantly affected by treatment (LRT, Binomial GLMM,  $\lambda^2 = 7.36$ , p = 0.061), with the WS-WW group being significantly more likely to survive compared to the control group (~45% of female polyps survived compared to 30% in the control group; Table 1., Fig. 4C.).

#### Discussion

Organisms are heavily affected by warming temperatures and show short- or long-term biological responses that include advancing phenologies, shifting geographical distribution ranges, and altered physiology (Huey and Kingsolver, 2019; McCarty, 2001; Thomas, 2010; Walther et al., 2002). Warming also affects biological fitness, such that organisms have altered reproduction and/or survival, with important consequences for population dynamics and ecosystem stability (Anderson, 2016; Deutsch et al., 2008; Kingsolver et al., 2013; Sinclair et al., 2016). While the fitness consequences of warming are widely considered to be negative, detailed investigations show a much more varied picture, with both negative and positive effects observed depending e.g. on the thermal tolerance and phenotypic plasticity of the species (Chamaillé-Jammes et al., 2006; Deutsch et al., 2008; Pelini et al., 2012; Weitere et al., 2009).

Here, I studied the fitness consequences of warming across the life cycle of a facultatively sexual freshwater cnidarian H. oligactis, an important predator of zooplankton in freshwater ecosystems. This species reproduces asexually during summer, switches to a sexual mode of reproduction after the temperature drops during autumn and experiences a post-reproductive senescence with increased mortality risk (Sebestyén et

al., 2020, 2018). However, some individuals do not initiate sexual reproduction at all, or survive and continue reproducing as exually during the winter. Due to the heat-intolerance of this species (Bosch et al., 1988) and the dependence of sexual reproduction on cold temperature in *H. oligactis*, I hypothesized that warming might have negative fitness effects and an overall shift from sexual to a sexual reproduction. The results paint a more complex picture of fitness effects of warming in Hydra.

Exposure of hydra polyps to simulated summer heat waves had immediate negative effects on the body size of hydra polyps. The reduction of body size with warming in animals is a widespread phenomenon (Gardner et al., 2011; Sheridan and Bickford, 2011), and is thought to be due to altered metabolic requirements and nutrient-dependence (Audzijonyte et al., 2022; Lee et al., 2015), oxygen limitation (Verberk et al., 2021; Walczyńska et al., 2015) or temperature-dependent size-fecundity or size-mortality relationships (Arendt, 2015; Audzijonyte et al., 2022). The size declines observed here were the result of phenotypic flexibility and occurred very quickly (observed after a week of exposure to simulated heat waves), suggesting a likely involvement of heat stress in the shrinkage of hydra polyps. Although such an immediate reduction in body size due to heat stress might not occur in most animals with a relatively fixed adult body size (e.g. insects), several other groups are able to plastically change adult size in response to environmental conditions (see e.g. Thommen et al., 2019 for an example of body size fluctuations in response to food availability in planarians). and in these animals warming, and the associated heatwaves could have an immediate effect on body size, just like in hydra. Furthermore, the changes in body size in hydra are significant because body size at cooling is a strong predictor of subsequent performance in this species: small polyps have delayed sexual maturation, reduced fecundity, but a higher post-reproductive survival (Ngo et al., 2021). In accordance, I found that polyps in the "Warm Summer" groups, exposed to simulated summer heat wave, required more time to produce the first gonads, had reduced sexual fitness and at least in males, they had a higher post-reproductive survival rate and higher asexual fitness.

The effects of the summer heat wave, however, were modulated by the winter treatment in a complex way. In H. oligactis, temperatures below 12 °C degrees are required for gamete differentiation to occur (Littlefield, 1991; Littlefield et al., 1991), and researchers previously used temperatures from 4 to 10  $^{\circ}$ C to induce sex in this species (Boutry et al., 2022; Kaliszewicz, 2015; Tomczyk et al., 2020). Higher simulated winter temperatures advanced sexual maturation in both males and females, which is consistent with a hypothesis that higher temperatures are more permissive for cell proliferation and the differentiation of gametes and reproductive tissue (Alvarez and Nicieza, 2002). These results suggest that, while all temperatures below or equal to 12  $^{\circ}C$  promote sexual development in *H. oligactis*, the exact temperature can affect fine details of sexual development. The strongest effects on sexual development and fecundity were, however, observed in the WS-CW group, where simulated summer heatwaves were followed by a sudden drop in temperature. Animals in this group needed the most time to produce the first gonads and, at least in males, they had the lowest number of reproductive organs. This suggests that sudden drops in temperature are stressful for Hydra, and this stress could contribute to sex induction in this species. While temperature fluctuations as large as this (from 23 to 8  $^{\circ}$ C within one day) are unlikely to occur under natural conditions in a freshwater habitat, both heat waves and cold spells, i.e. temperature variability are predicted to be more common in the future (Meehl et al., 2007) and these are likely to have negative physiological consequences in Hydra. Remarkably. I detected an unexpected consequence of simulated winter warming: polyps cultured under 12  $^{0}$ C underwent a second round of gonadogenesis and continued to show signs of sexual reproduction five months after cooling, while all surviving polyps cultured on 8  $^{\circ}$ C were asexual. Hence, while warming might have immediate negative effects on sexual reproduction in general in this species (especially if it demonstrates in increased temperature fluctuations), these negative effects could be counterbalanced on the longer term by the positive effects of warm winters on the number of reproductive cycles.

Compared to sexual reproduction, the effects of elevated temperatures on asexual fitness were much more clearcut (although sex-dependent). Polyps exposed to simulated heatwaves produced a higher number of asexual buds in male strains, and males exposed to elevated winter temperatures produced a higher number of asexual buds. The increased asexual fitness might be explained on one hand by a shift from sexual to asexual reproduction (possibly mediated by the reduced size of polyps exposed to simulated heat waves) and by the higher survival rate of polyps exposed to higher temperatures. Since asexual reproduction allows very quick population growth, these results suggest that increased temperatures will result in higher hydra population sizes in late winter due to climate warming. Hence, temperate freshwater bodies might experience "hydra blooms" similar to the warming-induced jellyfish blooms observed in marine environments (Goldstein and Steiner, 2020; Holst, 2012; Purcell et al., 2007), with potential up- and down-stream consequences on the whole aquatic food web. Unfortunately, little is known about hydra population dynamics under natural settings. Based on the phenology of resting egg production in *H. oligactis*, which peaks before the onset of the winter, one could assume that population size collapses during winter either due to freezing, low food availability, or increased mortality due to reduced somatic maintenance of sexually reproducing polyps (Sebestyén et al., 2018). The limited number of field observations, however, seem to contradict this assumption, since all observations point to the fact that *H. oligactis* thrives during the winter and reaches very large population densities (Bryden, 1952; Ribi et al., 1985; Welch and Loomis, 1924). The experiment presented here suggests that Hydra population sizes could become even higher in the future due to climate warming. Future studies should aim to test these predictions in a more natural setting, e.g., in mesocosms simulating the complexity of an aquatic food web to gain more insight into this problem.

In parallel with the higher asexual fitness of polyps exposed to simulated warming, I also found a positive effect of summer heatwaves and warmer winters on post-reproductive survival. Indeed, the higher asexual fitness of polyps exposed to higher temperature might be at least partly the consequence of their increased survival rate, although this effect was clearly observed only in males, while in females it was much more subtle. Hence, temperature appears in a growing list of intrinsic and extrinsic factors that affect post-reproductive survival in this species (age, size, genotype; Miklós et al., 2022; Ngo et al., 2021; Sebestyén et al., 2020). Exposure to high temperature could have resulted in higher survival for at least two different reasons. First, simulated summer heatwaves caused a reduction in body size, which is known to directly influence survival rate in this species (Ngo et al., 2021), most likely through a shift of resources from reproduction to survival. However, even male polyps that were not exposed to simulated summer heatwaves experienced improved survival rate if they were cultured at 12  $^{\circ}C$  simulated winter temperatures, which points at a second, independent mechanism. The higher temperature could have caused improved survival by altering the metabolic cost of tissue maintenance in these polyps (Gillooly et al., 2001). Alternatively, it could also be the consequence of a shift from reproduction to survival functions in polyps experiencing warmer winters, although no clear evidence of that was observed, since polyps in the "Warm Winter" groups did not show evidence of reduced testes number compared to the "Cold Winter" groups. Further studies should explore the effect of temperature on post-reproductive survival in more detail.

# References

Acker, T.S., Muscat, A.M., 1976. The Ecology of *Craspedacusta sowerbii* Lankester, a freshwater Hydrozoan. The American Midland Naturalist 95, 323–336. https://doi.org/10.2307/2424397

Ålvarez, D., Nicieza, A.G., 2002. Effects of temperature and food quality on anuran larval growth and metamorphosis. Functional Ecology 16, 640–648. https://doi.org/10.1046/j.1365-2435.2002.00658.x

Anderson, J.T., 2016. Plant fitness in a rapidly changing world. New Phytologist 210, 81–87. https://doi.org/10.1111/nph.13693

Angilletta, M.J., 2009. Temperature and the life history, in: Angilletta Jr., M.J. (Ed.), Thermal adaptation: A theoretical and empirical synthesis. Oxford University Press, p. 157-180. https://doi.org/10.1093/acprof:oso/9780198570875.003.0006

Angilletta, M.J., Niewiarowski, P.H., Navas, C.A., 2002. The evolution of thermal physiology in ectotherms. Journal of Thermal Biology 27, 249–268. https://doi.org/10.1016/S0306-4565(01)00094-8

Arendt, J., 2015. Why get big in the cold? Size-fecundity relationships explain the temperature-size rule in a pulmonate snail (*Physa*). Journal of Evolutionary Biology 28, 169–178. https://doi.org/10.1111/jeb.12554

Audzijonyte, A., Jakubavičiūtė, E., Lindmark, M., Richards, S.A., 2022. Mechanistic temperature-size rule

explanation should reconcile physiological and mortality responses to temperature. The Biological Bulletin 000-000. https://doi.org/10.1086/722027

Blanchard, S., Lognay, G., Verheggen, F., Detrain, C., 2019. Today and tomorrow: impact of climate change on aphid biology and potential consequences on their mutualism with ants. Physiological Entomology 44, 77–86. https://doi.org/10.1111/phen.12275

Bosch, T.C., Krylow, S.M., Bode, H.R., Steele, R.E., 1988. Thermotolerance and synthesis of heat shock proteins: these responses are present in *Hydra attenuata* but absent in *Hydra oligactis*. Proceedings of the National Academy of Sciences 85, 7927–7931.

Boutry, J., Tissot, S., Mekaoui, N., Dujon, A.M., Meliani, J., Hamede, R., Ujvari, B., Roche, B., Nedelcu, A.M., Tokolyi, J., Thomas, F., 2022. Tumors alter life history traits in the freshwater cnidarian, *Hydra* oligactis . iScience 25, 105034. https://doi.org/10.1016/j.isci.2022.105034

Brooks, M., E., Kristensen, K., Benthem, K., J., van, Magnusson, A., Berg, C., W., Nielsen, A., Skaug, H., J., Mächler, M., Bolker, B., M., 2017. glmmTMB balances speed and flexibility among packages for Zero-inflated Generalized Linear Mixed Modeling. The R Journal 9, 378. https://doi.org/10.32614/RJ-2017-066

Brown, J.H., Gillooly, J.F., Allen, A.P., Savage, V.M., West, G.B., 2004. Toward a Metabolic Theory of Ecology. Ecology 85, 1771–1789. https://doi.org/10.1890/03-9000

Bryden, R.R., 1952. Ecology of *Pelmatohydra oligactis* in Kirkpatricks Lake, Tennessee. Ecological Monographs 22, 45–68. https://doi.org/10.2307/1948528

Cáceres, C.E., Schwalbach, M.S., 2001. How well do laboratory experiments explain field patterns of zoo-plankton emergence? Freshwater Biology 46, 1179–1189. https://doi.org/10.1046/j.1365-2427.2001.00737.x

Chamaillé-Jammes, S., Massot, M., Aragón, P., Clobert, J., 2006. Global warming and positive fitness response in mountain populations of common lizards *Lacerta vivipara*. Global Change Biology 12, 392–402. https://doi.org/10.1111/j.1365-2486.2005.01088.x

Cooley, J.M., 1971. The effect of temperature on the development of resting eggs of *Diaptomus oregonensis* Lillj (copepoda: Calanoida). Limnology and Oceanography 16, 921–926. https://doi.org/10.4319/lo.1971.16.6.0921

Decaestecker, E., De Meester, L., Mergeay, J., 2009. Cyclical parthenogenesis in *Daphnia* : sexual versus asexual reproduction, in: Schön, I., Martens, K., Dijk, P. (Eds.), Lost Sex: The Evolutionary Biology of Parthenogenesis. Springer Netherlands, Dordrecht, pp. 295–316. https://doi.org/10.1007/978-90-481-2770-2\_15

Deutsch, C.A., Tewksbury, J.J., Huey, R.B., Sheldon, K.S., Ghalambor, C.K., Haak, D.C., Martin, P.R., 2008. Impacts of climate warming on terrestrial ectotherms across latitude. Proceedings of the National Academy of Sciences 105, 6668–6672. https://doi.org/10.1073/pnas.0709472105

Ekvall, M.K., Hansson, L.-A., 2012. Differences in recruitment and life-history strategy alter zooplankton spring dynamics under climate-change conditions. PLOS ONE 7, e44614. https://doi.org/10.1371/journal.pone.0044614

Franch-Gras, L., García-Roger, E.M., Serra, M., José Carmona, M., 2017. Adaptation in response to environmental unpredictability. Proceedings of the Royal Society B: Biological Sciences 284, 20170427. https://doi.org/10.1098/rspb.2017.0427

Fuchs, B., Wang, W., Graspeuntner, S., Li, Y., Insua, S., Herbst, E.-M., Dirksen, P., Böhm, A.-M., Hemmrich, G., Sommer, F., Domazet-Lošo, T., Klostermeier, U.C., Anton-Erxleben, F., Rosenstiel, P., Bosch, T.C.G., Khalturin, K., 2014. Regulation of Polyp-to-Jellyfish Transition in Aurelia aurita. Current Biology 24, 263–273. https://doi.org/10.1016/j.cub.2013.12.003 Gardner, J.L., Peters, A., Kearney, M.R., Joseph, L., Heinsohn, R., 2011. Declining body size: a third universal response to warming? Trends in Ecology & Evolution 26, 285–291.

Gergely, R, Tökölyi, J. *in press*. Resource availability modulates the effect of body size on reproductive development. Ecology and Evolution

Gilbert, J.J., 2017. Resting-egg hatching and early population development in rotifers: a review and a hypothesis for differences between shallow and deep waters. Hydrobiologia 796, 235–243. https://doi.org/10.1007/s10750-016-2867-7

Gillooly, J.F., Brown, J.H., West, G.B., Savage, V.M., Charnov, E.L., 2001. Effects of size and temperature on metabolic rate. Science 293, 2248–2251. https://doi.org/10.1126/science.1061967

Goldstein, J., Steiner, U.K., 2020. Ecological drivers of jellyfish blooms – The complex life history of a 'well-known' medusa (*Aurelia aurita*). Journal of Animal Ecology 89, 910–920. https://doi.org/10.1111/1365-2656.13147

Green, J., 1966. Seasonal variation in egg production by Cladocera. Journal of Animal Ecology 35, 77–104. https://doi.org/10.2307/2691

Gulbrandsen, J., Johnsen, G.H., 1990. Temperature-dependent development of parthenogenetic embryos in *Daphnia pulex* de Geer. Journal of Plankton Research 12, 443–453. https://doi.org/10.1093/plankt/12.3.443

Gyllström, M., Hansson, L.-A., 2004. Dormancy in freshwater zooplankton: Induction, termination and the importance of benthic-pelagic coupling. Aquat. Sci. 66, 274–295. https://doi.org/10.1007/s00027-004-0712-y

Hairston, N.G., Kearns, C.M., 1995. The interaction of photoperiod and temperature in diapause timing: a copepod example. The Biological Bulletin 189, 42–48. https://doi.org/10.2307/1542200

Hartig, F., 2022. DHARMa: Residual diagnostics for hierarchical (Multi-Level/Mixed) Regression Models (0.4.5).

Holst, S., 2012. Effects of climate warming on strobilation and ephyra production of North Sea scyphozoan jellyfish, in: Purcell, J., Mianzan, H., Frost, J.R. (Eds.), Jellyfish Blooms IV: Interactions with Humans and Fisheries, Developments in Hydrobiology. Springer Netherlands, Dordrecht, pp. 127–140. https://doi.org/10.1007/978-94-007-5316-7\_10

Huey, R.B., Kingsolver, J.G., 2019. Climate warming, resource availability, and the metabolic meltdown of ectotherms. The American Naturalist 194, E140–E150. https://doi.org/10.1086/705679

Huey, R.B., Kingsolver, J.G., 1989. Evolution of thermal sensitivity of ectotherm performance. Trends in Ecology & Evolution 4, 131–135. https://doi.org/10.1016/0169-5347(89)90211-5

Innes, D.J., 1997. Sexual reproduction of *Daphnia pulex* in a temporary habitat. Oecologia 111, 53–60. https://doi.org/10.1007/s004420050207

Jenni, L., Kéry, M., 2003. Timing of autumn bird migration under climate change: advances in long–distance migrants, delays in short–distance migrants. Proceedings of the Royal Society of London. Series B: Biological Sciences 270, 1467–1471. https://doi.org/10.1098/rspb.2003.2394

Kaliszewicz, A., 2015. Intensity-dependent response to temperature in Hydra clones. Zoological Sciences 32(1), 72-76. jzoo 32, 72-76. https://doi.org/10.2108/zs140052

Kingsolver, J.G., Diamond, S.E., Buckley, L.B., 2013. Heat stress and the fitness consequences of climate change for terrestrial ectotherms. Functional Ecology 27, 1415–1423. https://doi.org/10.1111/1365-2435.12145

Lee, K.P., Jang, T., Ravzanaadii, N., Rho, M.S., 2015. Macronutrient balance modulates the temperature-size rule in an ectotherm. The American Naturalist 186, 212–222. https://doi.org/10.1086/682072

Lenth, R., 2022. emmeans: Estimated Marginal Means, aka Least-Squares Means. R package version 1.8.3.

Littlefield, C.L., 1991. Cell lineages in Hydra: Isolation and characterization of an interstitial stem cell restricted to egg production in Hydra oligactis . Developmental Biology 143, 378–388. https://doi.org/10.1016/0012-1606(91)90088-K

Littlefield, C.L., Finkemeier, C., Bode, H.R., 1991. Spermatogenesis in *Hydra oligactis* : II. How temperature controls the reciprocity of sexual and asexual reproduction. Developmental Biology 146, 292–300. https://doi.org/10.1016/0012-1606(91)90231-Q

McCarty, J.P., 2001. Ecological consequences of recent climate change. Conservation Biology 15, 320–331. https://doi.org/10.1046/j.1523-1739.2001.015002320.x

McLean, K.D., Gowler, C.D., Dziuba, M.K., Zamani, H., Hall, S.R., Duffy, M.A., 2022. Sexual recombination and temporal gene flow maintain host resistance and genetic diversity. Evol Ecol. https://doi.org/10.1007/s10682-022-10193-6

McNamara, J.M., Houston, A.I., 2008. Optimal annual routines: behaviour in the context of physiology and ecology. Philosophical Transactions of the Royal Society B: Biological Sciences 363, 301–319. htt-ps://doi.org/10.1098/rstb.2007.2141

Meehl, G.A., Stocker, T.F., Collins, W.D., Friedlingstein, P., Gaye, A.T., Gregory, J.M., Kitoh, A., Knutti, R., Murphy, J.M., Noda, A., Raper, S.C.B., Watterson, I.G., Weaver, A.J., Zhao, Z.C., 2007. Global climate projections. Chapter 10.

Miklós, M., Laczkó, L., Sramkó, G., Barta, Z., Tökölyi, J., 2022. Seasonal variation of genotypes and reproductive plasticity in a facultative clonal freshwater invertebrate animal (*Hydra oligactis*) living in a temperate lake. Ecology and Evolution 12, e9096. https://doi.org/10.1002/ece3.9096

Miklós, M., Laczkó, L., Sramkó, G., Sebestyén, F., Barta, Z., Tökölyi, J., 2021. Phenotypic plasticity rather than genotype drives reproductive choices in Hydra populations. Molecular Ecology 30, 1206–1222. https://doi.org/10.1111/mec.15810

Molnár V, A., Tökölyi, J., Végvári, Z., Sramkó, G., Sulyok, J., Barta, Z., 2012. Pollination mode predicts phenological response to climate change in terrestrial orchids: a case study from central Europe. Journal of Ecology 100, 1141–1152. https://doi.org/10.1111/j.1365-2745.2012.02003.x

Ngo, K.S., R-Almási, B., Barta, Z., Tökölyi, J., 2021. Experimental manipulation of body size alters life history in hydra. Ecology Letters 24, 728–738. https://doi.org/10.1111/ele.13698

Panov, V.E., Krylov, P.I., Riccardi, N., 2004. Role of diapause in dispersal and invasion success by aquatic invertebrates. J Limnol 63, 56. https://doi.org/10.4081/jlimnol.2004.s1.56

Pelini, S.L., Diamond, S.E., MacLean, H., Ellison, A.M., Gotelli, N.J., Sanders, N.J., Dunn, R.R., 2012. Common garden experiments reveal uncommon responses across temperatures, locations, and species of ants. Ecology and Evolution 2, 3009–3015. https://doi.org/10.1002/ece3.407

Purcell, J.E., Uye, S., Lo, W.-T., 2007. Anthropogenic causes of jellyfish blooms and their direct consequences for humans: a review. Marine Ecology Progress Series 350, 153–174. https://doi.org/10.3354/meps07093

R Core Team, 2020. R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. URL https://www.R-project.org/.

Reisa, J.J., 1973. Ecology of hydra, in: Biology of Hydra. Academic Press, New York and London, pp. 59–105.

Ribi, G., Tardent, R., Tardent, P., Scascighini, C., 1985. Dynamics of hydra populations in Lake Zürich, Switzerland, and Lake Maggiore, Italy. Schweiz. Z. Hydrol 47, 45–56. https://doi.org/10.1007/BF02538183

Roff, D., 1993. Evolution Of Life Histories: Theory and Analysis. Springer Science & Business Media.

Scheuerl, T., Stelzer, C.-P., 2019. Asexual reproduction changes predator population dynamics in a life predator-prey system. Population Ecology 61, 210–216. https://doi.org/10.1002/1438-390X.1017

Schröder, T., 2005. Diapause in monogonont rotifers. Hydrobiologia 546, 291–306. https://doi.org/10.1007/s10750-005-4235-x

Sebestyén, F., Barta, Z., Tökölyi, J., 2018. Reproductive mode, stem cells and regeneration in a freshwater cnidarian with postreproductive senescence. Functional Ecology 32, 2497–2508. https://doi.org/10.1111/1365-2435.13189

Sebestyén, F., Miklós, M., Iván, K., Tökölyi, J., 2020. Age-dependent plasticity in reproductive investment, regeneration capacity and survival in a partially clonal animal (*Hydra oligactis*). Journal of Animal Ecology 89, 2246–2257. https://doi.org/10.1111/1365-2656.13287

Shaffer, M.R., Davy, S.K., Maldonado, M., Bell, J.J., 2020. Seasonally driven sexual and asexual reproduction in temperate Tethya species. Biol Bull 238, 89–105. https://doi.org/10.1086/708624

Sheridan, J.A., Bickford, D., 2011. Shrinking body size as an ecological response to climate change. Nature Climate Change 1, 401–406. https://doi.org/10.1038/nclimate1259

Simon, J.-C., Rispe, C., Sunnucks, P., 2002. Ecology and evolution of sex in aphids. Trends in Ecology & Evolution 17, 34–39. https://doi.org/10.1016/S0169-5347(01)02331-X

Sinclair, B.J., Marshall, K.E., Sewell, M.A., Levesque, D.L., Willett, C.S., Slotsbo, S., Dong, Y., Harley, C.D.G., Marshall, D.J., Helmuth, B.S., Huey, R.B., 2016. Can we predict ectotherm responses to climate change using thermal performance curves and body temperatures? Ecology Letters 19, 1372–1385. https://doi.org/10.1111/ele.12686

Stearns, S.C., 1989. Trade-offs in life-history evolution. Functional Ecology 3, 259–268. htt-ps://doi.org/10.2307/2389364

Thomas, C.D., 2010. Climate, climate change and range boundaries. Diversity and Distributions 16, 488–495. https://doi.org/10.1111/j.1472-4642.2010.00642.x

Thommen, A., Werner, S., Frank, O., Philipp, J., Knittelfelder, O., Quek, Y., Fahmy, K., Shevchenko, A., Friedrich, B.M., Jülicher, F., Rink, J.C., 2019. Body size-dependent energy storage causes Kleiber's law scaling of the metabolic rate in planarians. eLife 8, e38187. https://doi.org/10.7554/eLife.38187

Tökölyi, J., Ősz, Z., Sebestyén, F., Barta, Z., 2017. Resource allocation and post-reproductive degeneration in the freshwater cnidarian *Hydra oligactis* (Pallas, 1766). Zoology 120, 110–116. https://doi.org/10.1016/j.zool.2016.06.009

Tomczyk, S., Suknovic, N., Schenkelaars, Q., Wenger, Y., Ekundayo, K., Buzgariu, W., Bauer, C., Fischer, K., Austad, S., Galliot, B., 2020. Deficient autophagy in epithelial stem cells drives aging in the freshwater cnidarian Hydra. Development 147. https://doi.org/10.1242/dev.177840

van Baaren, J., Le Lann, C., JM van Alphen, J., 2010. Consequences of climate change for aphidbased multi-trophic systems, in: Kindlmann, P., Dixon, A.F.G., Michaud, J.P. (Eds.), Aphid biodiversity under environmental change: patterns and processes. Springer Netherlands, Dordrecht, pp. 55–68. https://doi.org/10.1007/978-90-481-8601-3\_4

Vandekerkhove, J., Declerck, S., Brendonck, L., Conde-Porcuna, J.M., Jeppesen, E., Meester, L.D., 2005. Hatching of cladoceran resting eggs: temperature and photoperiod. Freshwater Biology 50, 96–104. https://doi.org/10.1111/j.1365-2427.2004.01312.x

Verberk, W.C.E.P., Atkinson, D., Hoefnagel, K.N., Hirst, A.G., Horne, C.R., Siepel, H., 2021. Shrinking body sizes in response to warming: explanations for the temperature–size rule with special emphasis on the role of oxygen. Biological Reviews 96, 247–268. https://doi.org/10.1111/brv.12653

Vowinckel, C., 1970. The role of illumination and temperature in the control of sexual reproduction in the planarian *Dugesia tigrina*(girard). The Biological Bulletin 138, 77–87. https://doi.org/10.2307/1540293

Walczyńska, A., Labecka, A.M., Sobczyk, M., Czarnoleski, M., Kozłowski, J., 2015. The Temperature–Size Rule in *Lecane inermis* (Rotifera) is adaptive and driven by nuclei size adjustment to temperature and oxygen combinations. Journal of Thermal Biology, What sets the limit? How thermal limits, performance and preference in ectotherms are influenced by water or energy balance 54, 78–85. https://doi.org/10.1016/j.jtherbio.2014.11.002

Walther, G.-R., Post, E., Convey, P., Menzel, A., Parmesan, C., Beebee, T.J.C., Fromentin, J.-M., Hoegh-Guldberg, O., Bairlein, F., 2002. Ecological responses to recent climate change. Nature 416, 389–395. https://doi.org/10.1038/416389a

Weitere, M., Vohmann, A., Schulz, N., Linn, C., Dietrich, D., Arndt, H., 2009. Linking environmental warming to the fitness of the invasive clam *Corbicula fluminea*. Global Change Biology 15, 2838–2851. https://doi.org/10.1111/j.1365-2486.2009.01925.x

Welch, P.S., Loomis, H.A., 1924. A limnological study of *Hydra oligactis* in Douglas Lake, Michigan. Transactions of the American Microscopical Society 43, 203–235. https://doi.org/10.2307/3221738

Winder, M., Schindler, D.E., 2004. Climate change uncouples trophic interactions in an aquatic ecosystem. Ecology 85, 2100–2106. https://doi.org/10.1890/04-0151

Yoshida, K., Fujisawa, T., Hwang, J.S., Ikeo, K., Gojobori, T., 2006. Degeneration after sexual differentiation in hydra and its relevance to the evolution of aging. Gene, Evolutionary Genomics 385, 64–70. https://doi.org/10.1016/j.gene.2006.06.031

Table 1. Sexual development time, sexual fitness (number of gonads), asexual fitness (number of buds) and survival rate of male and female *H. oligactis* polyps exposed to simulated summer heatwave (WS-CW), elevated winter temperature (CS-WW), or both (WS-WW), compared to polyps exposed to a cold summer – cold winter scenario (CS-CW). The table shows estimated marginal means contrasts from Generalized Linear Mixed Models (GLMMs) that included treatment as a fixed effect, and strain ID and batch ID as random effects (see Methods for more detail). The type of model is indicated above the contrasts: Gaussian ("gaussian"), Negative Binomial with linear parametrization ("nbinom1"), Negative Binomial with quadratic parametrization ("nbinom2") or Binomial ("binomial"). P-values are after Dunnett's correction for multiple comparisons. Significant differences are highlighted in bold.

	Males	Males	Males	Females	Females	Females
Contrast	Estimate	SE	P-value	Estimate	SE	P-value
Sexual development time	gaussian	gaussian	gaussian	nbinom2	nbinom2	nbinom2
(WS-CW) - (CS-CW)	6.53	0.44	$<\!0.001$	0.26	0.03	$<\!0.001$
(CS-WW) - (CS-CW)	-1.04	0.43	0.047	-0.08	0.03	0.016
(WS-WW) - (CS-CW)	3.01	0.44	$<\!0.001$	0.13	0.03	$<\!0.001$
Sexual fitness	gaussian	gaussian	gaussian	nbinom1	nbinom1	nbinom1
(WS-CW) - (CS-CW)	-2.89	0.43	$<\!0.001$	-0.61	0.12	$<\!0.001$
(CS-WW) - (CS-CW)	-0.03	0.43	0.998	-0.04	0.10	0.910
(WS-WW) - (CS-CW)	-1.32	0.43	0.007	-0.21	0.10	0.082
Asexual fitness	nbinom1	nbinom1	nbinom1	nbinom2	nbinom2	nbinom2
(WS-CW) - (CS-CW)	0.30	0.07	$<\!0.001$	0.19	0.10	0.114
(CS-WW) - (CS-CW)	0.24	0.07	0.001	0.13	0.09	0.387
(WS-WW) - (CS-CW)	0.28	0.07	$<\!0.001$	0.21	0.09	0.067
Survival	binomial	binomial	binomial	binomial	binomial	binomial
(WS-CW) - (CS-CW)	1.75	0.45	$<\!0.001$	0.49	0.37	0.408
(CS-WW) - (CS-CW)	2.65	0.48	< 0.001	0.35	0.37	0.648

	Males	Males	Males	Females	Females	Females
(WS-WW) - (CS-CW)	2.40	0.48	< 0.001	0.97	0.37	0.025

# Figure legends

Fig. 1. Body size of *H. oligactis* polyps from three male and three female strains and cultured for one week on 18  $^{\circ}$ C (cool summer, CS) or exposed to a simulated one-week heatwave of 23  $^{\circ}$ C (warm summer, WS). A simulated summer heatwave significantly reduces both male and female body size (see Results).

Fig. 2. Sexual reproduction in male *H. oligactis* polyps belonging to three distinct strains exposed to four distinct temperature regimes: CS-CW (18  $^{\circ}$ C summer – 8  $^{\circ}$ C winter; control group), WS-CW (23  $^{\circ}$ C summer – 8  $^{\circ}$ C winter), CS-WW (8  $^{\circ}$ C summer – 12  $^{\circ}$ C winter) or WS-WW (23  $^{\circ}$ C summer – 12  $^{\circ}$ C winter). The summer treatment lasted for one week, while the winter treatment lasted 21 weeks. The graph shows the time elapsed until the appearance of the first gonads after cooling (A), the maximum number of testes (B) and temporal changes in the frequency of sexual individuals (C). Photograph of a male individual with mature testes is shown on panel D for illustration.

Fig. 3. Sexual reproduction in female *H. oligactis* polyps belonging to three distinct strains exposed to four distinct temperature regimes: CS-CW (18  $^{\circ}$ C summer – 8  $^{\circ}$ C winter; control group), WS-CW (23  $^{\circ}$ C summer – 8  $^{\circ}$ C winter), CS-WW (8  $^{\circ}$ C summer – 12  $^{\circ}$ C winter) or WS-WW (23  $^{\circ}$ C summer – 12  $^{\circ}$ C winter). The summer treatment lasted for one week, while the winter treatment lasted 21 weeks. The graph shows the time elapsed until the appearance of the first gonads after cooling (A), the total number of eggs (B) and temporal changes in the frequency of sexual individuals (C). Photograph of a female individual with mature eggs is shown on panel D for illustration.

Fig. 4. As exual reproduction and survival in *H. oligactis* polyps belonging to three male and three female strains, exposed to four distinct temperature regimes: CS-CW (18 °C summer – 8 °C winter; control group), WS-CW (23 °C summer – 8 °C winter), CS-WW (8 °C summer – 12 °C winter) or WS-WW (23 °C summer – 12 °C winter). The summer treatment lasted for one week, while the winter treatment lasted 21 weeks. The graph shows the total number of buds produced during the 22 weeks of the experiment (A) and proportion of individuals surviving (C). The number of surviving animals from each strain and treatment group is shown on the bars. Photographs of an asexual individual (B) and a senescent individual (D) are shown for illustration.

# Data accessibility statement:

Raw data is available on Figshare (https://doi.org/10.6084/m9.figshare.21750188.v1).

R code to analyze the data is available on Figshare (https://doi.org/10.6084/m9.figshare.21750170.v1).

# Competing interests statement:

The author declares no competing interests.

#### Author contributions:

Jácint Tökölyi : Conceptualization (lead); data curation (lead); formal analysis (lead); funding acquisition (lead); methodology (lead); project administration (lead); visualization (lead); writing (lead).

#### Acknowledgements

The author wishes to thank Réka Gergely, Erzsébet Ágnes Nehéz, Mariann Emese Zöld and Hanna Révész for lab assistance.





