

# Association of CFTR activity in sweat test, NPD, and ICM with ivacaftor and lumacaftor serum levels in Phe508del homozygous patients with cystic fibrosis

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August 28, 2022

## Abstract

Combination therapy with the cystic fibrosis (CF) transmembrane conductance regulator (CFTR) corrector lumacaftor and the CFTR potentiator ivacaftor has demonstrated significant impact on clinical parameters in Phe508del homozygous people with CF. Whether these changes under treatment are correlated to serum levels of both drugs had yet to be investigated. We therefore analyzed data from our previous study (OrkambiFacts, ClinicalTrials.gov Identifier: NCT02807415). In summary, we did not find statistically significant correlations between serum drug levels and changes in clinical parameters and biomarkers of CFTR function such as FEV1, BMI, sweat chloride, nasal potential difference (NPD) and intestinal current measurement (ICM). Absolute blood levels of lumacaftor or ivacaftor do not seem to be informative biomarkers to predict clinical improvement or the attenuation of the basic defect.

## Introduction

Combination therapy with the cystic fibrosis (CF) transmembrane conductance regulator (CFTR) corrector lumacaftor and the CFTR potentiator ivacaftor has demonstrated reduction of pulmonary exacerbations and a gain of body weight and lung function in Phe508del homozygous people with CF [1, 2]. We could previously show that lumacaftor-ivacaftor improves CFTR function determined by nasal potential difference (NPD) and intestinal current measurement (ICM) to 10 to 20% of normal CFTR activity. This effect on CFTR function was observed even in the absence of short term effects on lung function [9] Whilst the effects of lumacaftor/ivacaftor therapy on numerous facets of basic defect and clinical disease have been reported [3], published information about the pharmacokinetics of the drugs, the association of drug concentration in tissues and body fluids with clinical outcome and the relationship between serum concentrations and the effect on CFTR function remain scarce [4-7, 12].

Lumacaftor and ivacaftor are subject to clinically relevant drug-drug interactions between the CFTR modulators themselves and other concomitant medications used by people with CF. Lumacaftor is a CYP3A4/2C8/2C9/2C19 inducer and thus influences the turnover of ivacaftor, a CYP3A4 substrate [8]. Moreover, the two CFTR modulators are highly hydrophobic compounds. Consequently the tissue distribution of the drugs will depend on the body composition of fat, muscles and water which is highly variable

among CF patients depending on the extent of malabsorption and its nutritional management and physical activity.

In a prospective observational study we have examined clinical outcomes and biomarkers of CFTR function in Phe508del homozygous CF patients before and 8-16 weeks after initiation of lumacaftor-ivacaftor therapy [9]. Here we report on the association of serum concentrations of lumacaftor and ivacaftor with biomarkers of CFTR function and clinical outcomes obtained on the same day. Here we report on the association of serum concentrations of lumacaftor and ivacaftor with biomarkers of CFTR function and clinical outcomes obtained on the same day.

## Materials and Methods

This prospective observational post-approval multicenter study (ClinicalTrials.gov Identifier: NCT02807415) was conducted at three CF centers of the German Center for Lung Research (DZL) in Phe508del homozygous CF patients aged 12 years and older. Prior to and 8 – 16 weeks after the initiation of lumacaftor-ivacaftor therapy anthropometry, lung function, sweat chloride concentrations, NPD and ICM were measured in all participants [9, 11]. Collection of blood and rectal biopsies for ICM, sweat test and NPD measurements were performed within 2 to 3 hours after the administration of the morning dose of lumacaftor/ivacaftor. Serum samples were stored at -30°C until further analysis.

Serum levels of lumacaftor and ivacaftor were analyzed by a validated liquid chromatography method with UV and mass detection. Briefly, 200 µl serum was deproteinised by the addition of 600 µl methanol and subsequent centrifugation at 13,000 rpm for 15 min. The supernatant was separated by an UltiMate 3000 UHPLC system (Thermo, Waltham, Ma) on a Reprosil Pur Basic C18 (3µm, 100x2mm) column with 0.1% formic acid (v/v) / methanol as mobile phase. Lumacaftor was quantified by UV-spectrophotometry at 254 nm. Ivacaftor was quantified by quadrupole time-of-flight mass spectrometry (Bruker Daltonik, Bremen, Germany) using D18 Ivacaftor as an internal standard (ivacaftor  $m/z= 391.2$ , D18-Ivacaftor  $m/z= 409.3$ ). The injected volume was 4 µl. The detection limit was 2 µg/ml for lumacaftor and 0.2 µg/ml for ivacaftor.

The study protocol was approved by the ethics committees of all participating centers. Written informed consent was obtained from all patients included in the study, their parents or legal guardians.

## Results

In total, 51 p.Phe508del homozygous CF patients took lumacaftor and ivacaftor within this post-approval study. Eight to sixteen weeks after the initiation of treatment with CFTR modulators blood was taken to determine the serum concentrations of lumacaftor and ivacaftor (Supplementary Table S1). The median serum levels were 0.5 µg/ml for ivacaftor and 24 µg/ml for lumacaftor (Table 1), comparable to data from the EMA assessment report [8]. The serum samples contained on average 50-fold more lumacaftor than ivacaftor. Figure 1 shows the ivacaftor and lumacaftor concentrations for the 49 samples with less than 2 µg/ml ivacaftor, the remaining two samples contained high levels of ivacaftor of 9 and close to 4 µg/ml, respectively (Supplementary Table S1). The data in Figure 1 demonstrate the lack of any correlation between ivacaftor and lumacaftor levels.

The serum levels of lumacaftor, ivacaftor and the lumacaftor/ivacaftor ratio were tested for association with anthropometry, spirometry and CFTR activity determined within 0.1 – 2.5 hours past blood sampling. In 14 of 15 tests no significant correlations between drug serum levels and clinical parameters. Neither body mass index, FEV1, sweat chloride concentration, Sermet score [10] of the NPD and the chloride secretory response [9] in the ICM (Table 2) correlated. One exception from this general finding was noted, i.e. the serum level of ivacaftor correlated with the decrease of sweat chloride concentration ( $P = 0.02$ ). The correlation of the ratio of lumacaftor to ivacaftor levels with the increase of the Sermet score towards the normal range in the NPD showed a trend ( $P = 0.06$ ).

## Discussion

Our multicenter study revealed that combination therapy of Phe508del homozygous individuals with CF with

lumacaftor/ivacaftor leads to highly variable serum concentrations of the two drugs that are not correlated with each other. Ivacaftor and lumacaftor have half-lives of 12 and 23 hours, respectively [8]. Since our study participants administered the morning dose according to protocol and blood was subsequently collected after a defined time lag with low inter-patient variability, different time points of blood sampling after dosage can hardly explain the large patient-to-patient variation of drug concentrations. We therefore speculate that the individual genetic repertoire of the polymorphic cytochrome P450 superfamily, drug-drug interactions and a variable body composition, particularly of the fat compartment, may give rise to a broad distribution of absorption, residence time, metabolism and excretion of these hydrophobic drugs.

According to our knowledge three groups have published protocols to determine the concentration of lumacaftor or ivacaftor by LC-MS in primary cells [6], sputum [4] and serum [4 -7] from in total 13 CF patients [4-7]. Our data on samples from 51 Phe508del homozygotes CF patients corroborate the previous reports that serum concentrations of ivacaftor during concomitant medication with lumacaftor are substantially lower than during ivacaftor monotherapy confirming the manufacturer's report [8] that lumacaftor is a CYP3A4 inducer that leads to a more rapid turnover of the CYP3A4 substrate ivacaftor.

With the exception of the correlation between ivacaftor levels and the change in sweat chloride, the absolute drug levels of lumacaftor or ivacaftor did not correlate with any improvement in anthropometry, lung function or CFTR activity in sweat gland, respiratory or intestinal epithelium. The remaining ivacaftor levels in presence of lumacaftor might therefore be of importance. However, our data indicate that the absolute blood levels of lumacaftor or ivacaftor seem to be no informative biomarkers to predict clinical improvement or the attenuation of the CF basic defect by combination therapy with these CFTR modulators. These findings are consistent with previous reports containing smaller numbers of ICM and NPD [12].

Nevertheless, drug monitoring of lumacaftor/ivacaftor may still be helpful to assess the patient's adherence to the prescribed treatment. Further studies are required to assess whether the CFTR correctors elxacaftor and tezacaftor [13-17] exert a similar pharmacokinetic behavior as observed for ivacaftor and lumacaftor.

#### Ethics approval

The study protocol was approved by the ethics committee of Hannover Medical School (No. 2846-2015) and subsequently by the ethics committees of the University of Heidelberg and the University of Giessen.

#### Sources of funding

Supported by the German Federal Ministry of Education and Research (82DZLE12A1) within the Clinical Trials Program of the German Center for Lung Research

#### Declaration of Competing Interest

C.D., R.T., A.M.D, S.Y.G, J.M.C., and C.D.E have nothing to disclose

M.J.H. received honoraria from Amgen GmbH, Baxter Deutschland GmbH, Biotest Pharma GmbH, CSL-Behring GmbH, Fresenius Kabi GmbH, Leo Pharma GmbH, Merck-Serono GmbH, Novartis Pharma GmbH, Pfizer PFE GmbH, Roche AG, Sun Pharmaceuticals

L.N. reports grants from German Center for Lung Research, during the conduct of the study; personal fees from Vertex Pharmaceuticals, personal fees from Boehringer Ingelheim, outside the submitted work; .

M.S. is a central multiple-breath washout overreader in the Vertex sponsored trial VX16-809-121 and principal investigator and subinvestigator in a number of Vertex sponsored trials on CFTR modulators in patients with CF

B.T. reports grants from German Federal Ministry of Education and Research , during the conduct of the study; personal fees and other from Vertex Pharmaceuticals, outside the submitted work;

M.A.M. reports grants from German Federal Ministry of Education and Research , grants from Einstein Foundation Berlin, during the conduct of the study; personal fees and other from Boehringer Ingelheim,

personal fees from Arrowhead Pharmaceuticals, personal fees and other from Vertex Pharmaceuticals, personal fees from Santhera, personal fees from Sterna Biologicals, personal fees from Enterprise Therapeutics, personal fees from Antabio, personal fees from Kither Biotech, personal fees from Prieris Pharmaceuticals, outside the submitted work;

### Acknowledgements

The authors thank the patients with cystic fibrosis for their participation in this study; A. Fichtner and J. Pfeil for performing rectal biopsy procedures; C. Rueckes-Nilges, C. Herth, S. Tamm, C. Berger, and S. Engelhardt for excellent technical assistance; and S. Barth, S. Hämmerling, S. Junge, A. Sauer-Heilborn, F. Ringshausen, O. Sommerburg, and J. Schäfer-Feterowski for clinical care of study participants

### References

- [1] Wainwright CE, Elborn JS, Ramsey BW, Marigowda G, Huang X, Cipolli M, Colombo C, Davies JC, De Boeck K, Flume PA, Konstan MW, McColley SA, McCoy K, McKone EF, Munck A, Ratjen F, Rowe SM, Waltz D, Boyle MP; TRAFFIC Study Group; TRANSPORT Study Group. Lumacaftor-Ivacaftor in Patients with Cystic Fibrosis Homozygous for Phe508del CFTR. *N Engl J Med* 2015;373(3):220-31. doi: 10.1056/NEJMoa1409547.
- [2] Konstan MW, McKone EF, Moss RB, Marigowda G, Tian S, Waltz D, Huang X, Lubarsky B, Rubin J, Millar SJ, Pasta DJ, Mayer-Hamblett N, Goss CH, Morgan W, Sawicki GS. Assessment of safety and efficacy of long-term treatment with combination lumacaftor and ivacaftor therapy in patients with cystic fibrosis homozygous for the F508del-CFTR mutation (PROGRESS): a phase 3, extension study. *Lancet Respir Med*. 2017;5(2):107-118. doi: 10.1016/S2213-2600(16)30427-1.
- [3] Bell SC, Mall MA, Gutierrez H, Macek M, Madge S, Davies JC, Burgel PR, Tullis E, Castaños C, Castellani C, Byrnes CA, Cathcart F, Chotirmall SH, Cosgriff R, Eichler I, Fajac I, Goss CH, Drevinek P, Farrell PM, Gravelle AM, Havermans T, Mayer-Hamblett N, Kashirskaya N, Kerem E, Mathew JL, McKone EF, Naehrlich L, Nasr SZ, Oates GR, O'Neill C, Pypops U, Raraigh KS, Rowe SM, Southern KW, Sivam S, Stephenson AL, Zampoli M, Ratjen F. The future of cystic fibrosis care: a global perspective. *Lancet Respir Med*. 2020 Jan;8(1):65-124. doi: 10.1016/S2213-2600(19)30337-6.
- [4] Schneider EK, Reyes-Ortega F, Wilson JW, Kotsimbos T, Keating D, Li J, Velkov T. Development of HPLC and LC-MS/MS methods for the analysis of ivacaftor, its major metabolites and lumacaftor in plasma and sputum of cystic fibrosis patients treated with ORKAMBI or KALYDECO. *J Chromatogr B Analyt Technol Biomed Life Sci* 2016;1038:57-62. doi: 10.1016/j.jchromb.2016.10.026.
- [5] Schneider EK, Reyes-Ortega F, Li J, Velkov T. Optimized LC-MS/MS Method for the High-throughput Analysis of Clinical Samples of Ivacaftor, Its Major Metabolites, and Lumacaftor in Biological Fluids of Cystic Fibrosis Patients. *J Vis Exp* 2017;(128):56084. doi: 10.3791/56084.
- [6] Guimbellot JS, Ryan KJ, Anderson JD, Liu Z, Kersh L, Esther CR, Rowe SM, Acosta EP. Variable cellular ivacaftor concentrations in people with cystic fibrosis on modulator therapy. *J Cyst Fibros*. 2020;19(5):742-745. doi: 10.1016/j.jcf.2020.01.011.
- [7] Vonk SEM, van der Meer-Vos M, Bos LDJ, Neerincx AH, Majoor CJ, Maitland-van der Zee AH, Mathôt RAA, Kemper EM. A Quantitative Method for the Analysis of Ivacaftor, Hydroxymethyl Ivacaftor, Ivacaftor Carboxylate, Lumacaftor, and Tezacaftor in Plasma and Sputum Using LC-MS/MS and Its Clinical Applicability. *Ther Drug Monit* 2020 Nov 4. doi: 10.1097/FTD.0000000000000829.
- [8] EMA. Assessment report ORKAMBI (ivacaftor/lumacaftor). European medicines agency. 24 September 2015 EMA/686121/2018
- [9] Graeber SY, Dopfer C, Naehrlich L, Gyulumyan L, Scheuermann H, Hirtz S, Wege S, Mairbäurl H, Dorda M, Hyde R, Bagheri-Hanson A, Rueckes-Nilges C, Fischer S, Mall MA, Tümmler B. Effects of Lumacaftor-Ivacaftor Therapy on Cystic Fibrosis Transmembrane Conductance Regulator Function in

Phe508del Homozygous Patients with Cystic Fibrosis. *Am J Respir Crit Care Med* 2018;197(11):1433-1442. doi: 10.1164/rccm.201710-1983OC.

[10] Sermet-Gaudelus I, Girodon E, Sands D, Stremmler N, Vavrova V, Deneuille E, Reix P, Bui S, Huet F, Lebourgeois M, Munck A, Iron A, Skalicka V, Bienvenu T, Roussel D, Lenoir G, Bellon G, Sarles J, Macek M, Roussey M, Fajac I, Edelman A. Clinical phenotype and genotype of children with borderline sweat test and abnormal nasal epithelial chloride transport. *Am J Respir Crit Care Med*. 2010;182(7):929-36. doi: 10.1164/rccm.201003-0382OC.

[11] Hirtz S, Gonska T, Seydewitz HH, Thomas J, Greiner P, Kuehr J, et al. CFTR Cl<sup>-</sup> channel function in native human colon correlates with the genotype and phenotype in cystic fibrosis. *Gastroenterology* 2004; 127:1085–1095.

[12] Masson A, Schneider-Futschik EK, Baatallah N, Nguyen-Khoa T, Girodon E, Hatton A, Flament T, Le Bourgeois M, Chedevergne F, Bailly C, Kyrilli S, Achimastos D, Hinzpeter A, Edelman A, Sermet-Gaudelus I. Predictive factors for lumacaftor/ivacaftor clinical response. *J Cyst Fibros*. 2019 May;18(3):368-374. doi: 10.1016/j.jcf.2018.12.011. Epub 2018 Dec 28. PMID: 30595473

[13] Mall MA, Mayer-Hamblett N, Rowe SM. Cystic Fibrosis: Emergence of Highly Effective Targeted Therapeutics and Potential Clinical Implications. *Am J Respir Crit Care Med*. 2020 May 15;201(10):1193-1208.

[14] Middleton PG, Mall MA, Dřevínek P, Lands LC, McKone EF, Polineni D, Ramsey BW, Taylor-Cousar JL, Tullis E, Vermeulen F, Marigowda G, McKee CM, Moskowitz SM, Nair N, Savage J, Simard C, Tian S, Waltz D, Xuan F, Rowe SM, Jain R; VX17-445-102 Study Group. Elexacaftor-Tezacaftor-Ivacaftor for Cystic Fibrosis with a Single Phe508del Allele. *N Engl J Med*. 2019 Nov 7;381(19):1809-1819

[15] Heijerman HGM, McKone EF, Downey DG, Van Braeckel E, Rowe SM, Tullis E, Mall MA, Welter JJ, Ramsey BW, McKee CM, Marigowda G, Moskowitz SM, Waltz D, Sosnay PR, Simard C, Ahluwalia N, Xuan F, Zhang Y, Taylor-Cousar JL, McCoy KS; VX17-445-103 Trial Group. Efficacy and safety of the elexacaftor plus tezacaftor plus ivacaftor combination regimen in people with cystic fibrosis homozygous for the F508del mutation: a double-blind, randomised, phase 3 trial. *Lancet*. 2019 Nov 23;394(10212):1940-1948.

[16] Griese M, Costa S, Linnemann RW, Mall MA, McKone EF, Polineni D, Quon BS, Ringshausen FC, Taylor-Cousar JL, Withers NJ, Moskowitz SM, Daines CL. Safety and Efficacy of Elexacaftor/Tezacaftor/Ivacaftor for 24 Weeks or Longer in People with Cystic Fibrosis and One or More F508del Alleles: Interim Results of an Open-Label Phase 3 Clinical Trial. *Am J Respir Crit Care Med*. 2021 Feb 1;203(3):381-385.

[17] Nichols DP, Paynter AC, Heltshe SL, Donaldson SH, Frederick CA, Freedman SD, Gelfond D, Hoffman LR, Kelly A, Narkewicz MR, Pittman JE, Ratjen F, Rosenfeld M, Sagel SD, Schwarzenberg SJ, Singh PK, Solomon GM, Stalvey MS, Clancy JP, Kirby S, Van Dalen JM, Kloster MH, Rowe SM; PROMISE Study group. Clinical Effectiveness of Elexacaftor/Tezacaftor/Ivacaftor in People with Cystic Fibrosis: A Clinical Trial. *Am J Respir Crit Care Med*. 2022 Mar 1;205(5):529-539.

### Supplementary Materials

Table 1. Serum levels of lumacaftor and ivacaftor in 51 p.Phe508del homozygous individuals with CF

	Median (IQR)	Range
Ivacaftor [µg/ml]	0.5 (0.3 - 0.8)	0.1 - 9.0
Lumacaftor [µg/ml]	23.5 (17.9 - 33.3)	0.0 - 52.2
Lumacaftor/Ivacaftor	50.8 (35.8 - 82.0)	0 - 390

Table 2. Correlation of ivacaftor and lumacaftor serum levels with the change to baseline in clinical features and CFTR activity\*

	$\Delta$ BMI	$\Delta$ FEV1 predicted	$\Delta$ Sweat chloride	$\Delta$ Sernet score	$\Delta$ IBMX/ Forskolin
Ivacaftor	-0.012 (0.94)	0.008 (0.96)	0.334 (0.02)	-0.004 (0.98)	-0.059 (0.71)
Lumacaftor	0.143 (0.33)	0.077 (0.60)	0.169 (0.25)	0.066 (0.67)	0.127 (0.42)
Luma/Iva	0.003 (0.98)	0.166 (0.25)	-0.001 (0.92)	0.279 (0.06)	0.077 (0.62)

\*For each entry the Table first denotes the correlation coefficient followed by the *P* value (Pearson correlation test; in brackets)

Figure 1. Serum levels of lumacaftor and ivacaftor of p.Phe508del homozygous individuals with CF during combination therapy

